



## Apoptotic Effects of Beta-Carotene, Alpha-Tocopherol and Ascorbic Acid on PC-3 Prostate Cancer Cells

### Beta-Karoten, Alfa-Tokoferol ve Askorbik Asidin PC-3 Prostat Kanser Hücrelerine Apoptotik Etkileri

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#### ABSTRACT

Prostate cancer (PC) is one of the most commonly diagnosed cancer types being the second major reason of cancer-associated death in male particularly over the age of 50. Accumulating scientific evidences suggest the role oxidative stress and reactive oxygen species (ROS) in prostate cancer. A variety of factors including carcinogenic molecules, infectious diseases and toxic compounds can induce ROS production which turns into a strong contribution to the disturbed homeostasis and genetic mutation. Antioxidants can decrease the negative effects of ROS *in vitro*. Vitamin C (Ascorbic acid, Asc), vitamin A (beta carotenoids and retinoids,  $\beta$ -Crt) and vitamin E (alpha tocopherol,  $\alpha$ -Toc) play an important role in inhibition of oxidative stress and diminishing of free radicals in the body. The aim of this study was to determine the anticancer effect of  $\alpha$ -Toc,  $\beta$ -Crt and Asc on PC-3 prostate cancer cells *in vitro*. This was carried out by cell proliferation, ROS and Lipid Peroxidation assay, caspase-3 and propidium iodide staining experiments. The findings suggest that these agents behave as prooxidant by lowering cell viability and increasing the production of ROS and LPO in prostate cancer. These oxidants induce apoptosis as supported by caspase-3 (the enzyme playing key role in programmed cell death) staining by displaying a marked increase in the expression level of caspase-3 enzyme.

#### Key Words

Prostate cancer; reactive oxygen species; apoptosis; caspase-3.

#### ÖZ

Prostat kanseri (PC), özellikle 50 yaşın üzerindeki erkeklerde kansere bağlı ölümlerin ikinci büyük nedeni olan ve en yaygın olarak teşhis edilen kanser tiplerinden biridir. Bilimsel çalışmalar oksidatif stres ve Reaktif oksijen türlerinin (ROS) prostat kanseri üzerindeki rolünü göstermektedir. ROS, kanserojen moleküller, enfeksiyon, toksik bileşikler gibi homeostaza ve genetik mutasyona neden olabilecek bileşikler tarafından üretilir. Antioksidanlar, ROS'un olumsuz etkilerini *in vitro* olarak azaltabilir. C vitamini (Askorbik asit, Asc), A vitamini (beta karotenoidler ve retinoidler,  $\beta$ -Crt) ve E vitamini (alfa tokoferol,  $\alpha$ -Toc) oksidasyonun önlenmesinde ve vücuttaki serbest radikallerin konsantrasyonunun azaltılmasında önemli rol oynar. Bu çalışmanın amacı,  $\alpha$ -Toc,  $\beta$ -Crt ve Asc'nin PC-3 prostat kanseri hücreleri üzerindeki *in vitro* antikanser etkisini belirlemektir. Bu amaç, hücre çoğalması, ROS ve Lipid Peroksidasyon deneyi, kaspaz-3 ve propidium iyodür boyama deneyleri ile gerçekleştirildi. Bulgular, bu ajanların, prostat kanseri hücrelerinde hücre canlılığını azaltarak ve ROS ve LPO üretimini artırarak proksidan olarak davrandığını göstermektedir. Bu oksidanlar kaspaz-3 (programlı hücre ölümünde rol alan önemli bir enzim) boyamasıyla desteklediği üzere apoptozu kaspaz-3 enziminin ekspresyonunu artırarak indüklemiştir.

#### Anahtar Kelimeler

Prostat kanseri; reaktif oksijen türleri; apoptoz; kaspaz-3.

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## INTRODUCTION

Prostate cancer is one of the most common cancer types of men in the World 10% of which results in death [1]. Many studies have focused the relation between oxidative stress and prostate cancer [2-4]. The generation of reactive oxygen species (ROS) and changes in redox status are common biochemical phenomena in cancer cells. ROS can attack polyunsaturated fatty acids of lipid membranes and induce lipid peroxidation (LPO). An excessive increase in the amount of ROS and LPO induced by different prooxidants leads to oxidative stress and apoptosis leading to overcleaning of the cellular antioxidant defense system [5-6]. Antioxidants have been widely used as dietary supplements and have been investigated for their effectiveness in prevention of many diseases including various types of cancer. The vitamins C (Ascorbic acid, Asc), A (beta carotenoids and retinoids,  $\beta$ -Crt) and E (alpha tocopherol,  $\alpha$ -Toc) are known to act as antioxidants by delaying or inhibiting oxidation and reducing the concentration of free radicals in the body. In addition to their use in cancer prevention, supplemental antioxidants have usually been prescribed to cancer patients either by clinicians or patients themselves [7-8]. Vitamin E is reported as an efficient lipid soluble antioxidant that functions as a 'chain breaker' during lipid peroxidation process in cell membranes. It shows functionality by intercepting lipid peroxy radicals ( $\text{LOO}^\cdot$ ) and terminating the lipid peroxidation chain reactions resulting in stable tocopheroxy radical which is insufficiently reactive to initiate lipid peroxidation itself in normal circumstances, showing an essential criterion of a good antioxidant [9- 12]. Additionally, some studies demonstrate that vitamin E shows its antioxidant properties through scavenging  $\text{LOO}^\cdot$  in vivo as well as *in vitro* systems. Despite being good scavenger of lipid peroxy, it is not an efficient scavenger of  $\text{OH}^\cdot$  and alkoxy radicals ( $\text{OR}^\cdot$ ) in vivo [13].

Vitamin C or ascorbic acid, is a well-known water-soluble free radical scavenger. In addition to its role being a free radical scavenger, it has important function in regeneration of vitamin E in cell membranes within combination with reducing antioxidant glutathione [14-16, 9].

The precursors of vitamin A,  $\beta$ -Crt, are natural compounds with lipophilic properties. 500 different compounds of carotenoids have been reported until today. Most of the studied carotenoids have an extended

system of conjugated double bonds, which is responsible for their antioxidant activity. Epidemiologic research in humans have demonstrated that  $\beta$ -Crt has an important role in cancer prevention. In addition to this, the carotenoids have also been demonstrated to have the ability of inhibiting free radical reactions. Among these carotenoids,  $\beta$ -Crt was shown to reduce tri chloromethylperoxy radicals.  $\beta$ -Crt was also found to inhibit the oxidation of model compounds (tetralin and methyl linoleate) by peroxy radicals [17].

The aim of this study was to determine the anticancer effect of  $\alpha$ -Toc,  $\beta$ -Crt and Asc on PC-3 prostate cancer cells *in vitro* and elucidate underlying molecular mechanism of the antioxidants. In this context, the effects of these antioxidants on cell viability, production of ROS and LPO and their relationship with apoptosis were evaluated in this study.

## MATERIALS and METHODS

### Cell Culture

Human prostate cancer cells [PC-3 (ATCC® CRL1435™)] were cultured in complete endothelial growth media containing 10 % fetal bovine serum, 1 % penicillin-streptomycin (10000 units/ml, 10 mg/ml streptomycin). The cells were cultured in humidified incubator with 5 %  $\text{CO}_2$  and checked every two or three days. Cells were checked for mycoplasma contamination by using EZ-PCR mycoplasma test kit (Biological Industries).

### Cell Proliferation Assay

Water Soluble Tetrazolium-1 (WST-1) cell proliferation assay kit (Clontech Laboratories, USA) was used to investigate the effects of  $\alpha$ -Toc,  $\beta$ -Crt and Asc on PC-3 cells. Experiments were carried out according to the procedure provided by the supplier. Initially 10.000 PC-3 cells were seeded in 96-well plate. After that the cells were treated with different doses of agents between 15 to 60  $\mu\text{g}/\text{ml}$ . Subsequently, 5  $\mu\text{l}$  of WST-1 was added into each well. After 4h incubation, absorbance of each well was recorded at 450 nm (reference: 630 nm) by SpectraMax Plus 384 Microplate Reader (Molecular Devices, USA).

### Intracellular ROS Detection

The amount of ROS was measured by 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA). The cells were treated with 60  $\mu\text{g}/\text{ml}$  of  $\alpha$ -Toc,  $\beta$ -Crt and Asc.  $1 \times 10^6$  cells were harvested and incubated in the presence of 2  $\mu\text{M}$

DCFH-DA at 37°C for 1h. Fluorescence measurements were recorded by a spectrofluorometer (Perkin-Elmer LS-55, USA) at wavelengths of 485 nm (excitation) and 525 nm (emission).

#### Lipid Peroxidation Assay

The antioxidant activity of  $\alpha$ -Toc,  $\beta$ -Crt and Asc was evaluated by LPO assay measuring malondialdehyde as a malondialdehyde-thiobarbituric acid adduct as explained by Smith [18]. Malondialdehyde bis was used for standard graph preparation [18].

#### Propidium Iodide Staining for Apoptosis

Cells were seeded and harvested in 96-well plate. Later, cells were treated with 60  $\mu$ g/ml of  $\alpha$ -Toc,  $\beta$ -Crt and Asc. Then, the cells were washed with PBS twice. Propidium iodide was added to each well and cells were imaged by inverted microscope with fluorescence attachment (Olympus, JAPAN).

#### Immunohistochemical Staining of Caspase-3

Immunohistochemical staining was performed as explained in [19]. The cells containing dark brown nuclei or cytoplasm identified as caspase-3 (+) cells.

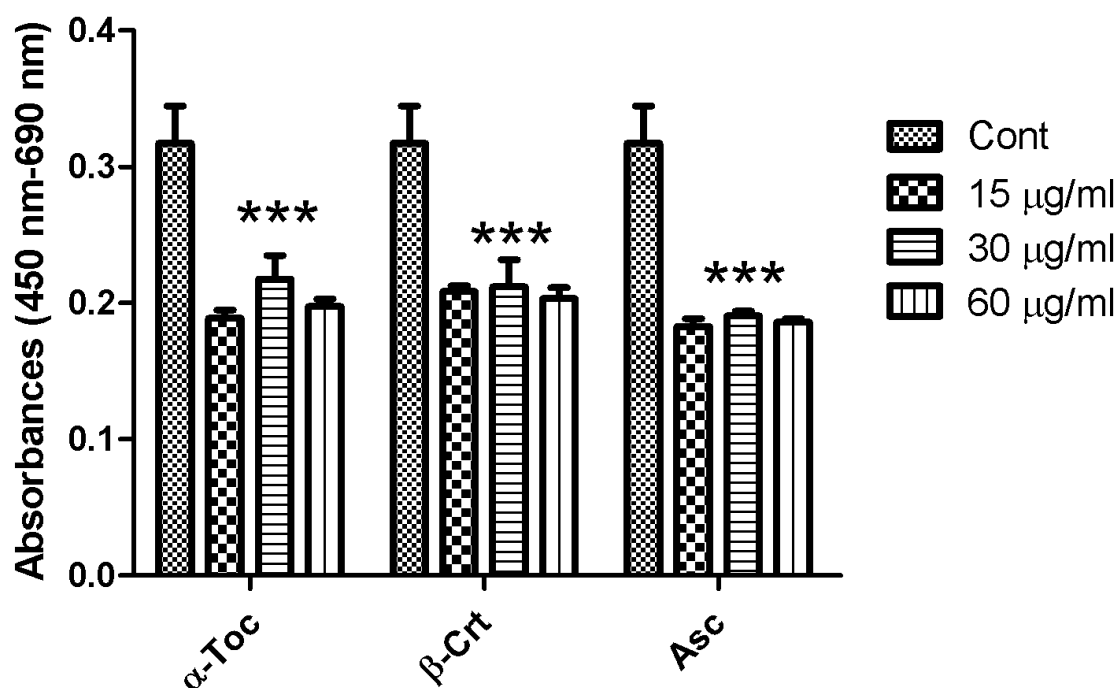
## RESULTS

#### Cell Viability

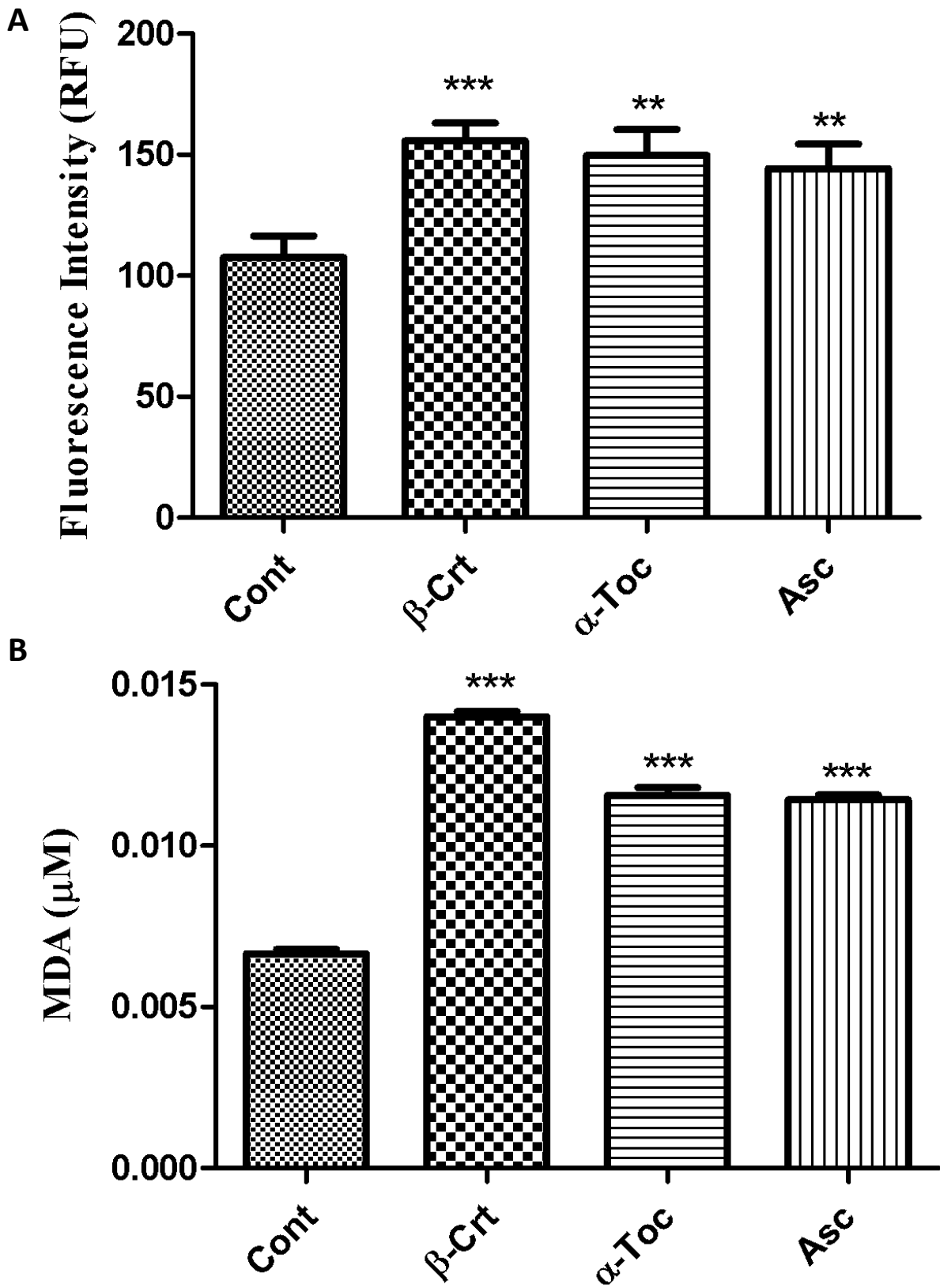
The viability of  $\alpha$ -Toc,  $\beta$ -Crt and Asc on PC 3 cell line was determined by WST-1 assay. The results of representative experiments are shown (Figure 1). In general all antioxidants tested within this study decreased cell viability at concentrations of 60  $\mu$ g/ml when compared to control.

#### Intracellular ROS and LPO Analysis

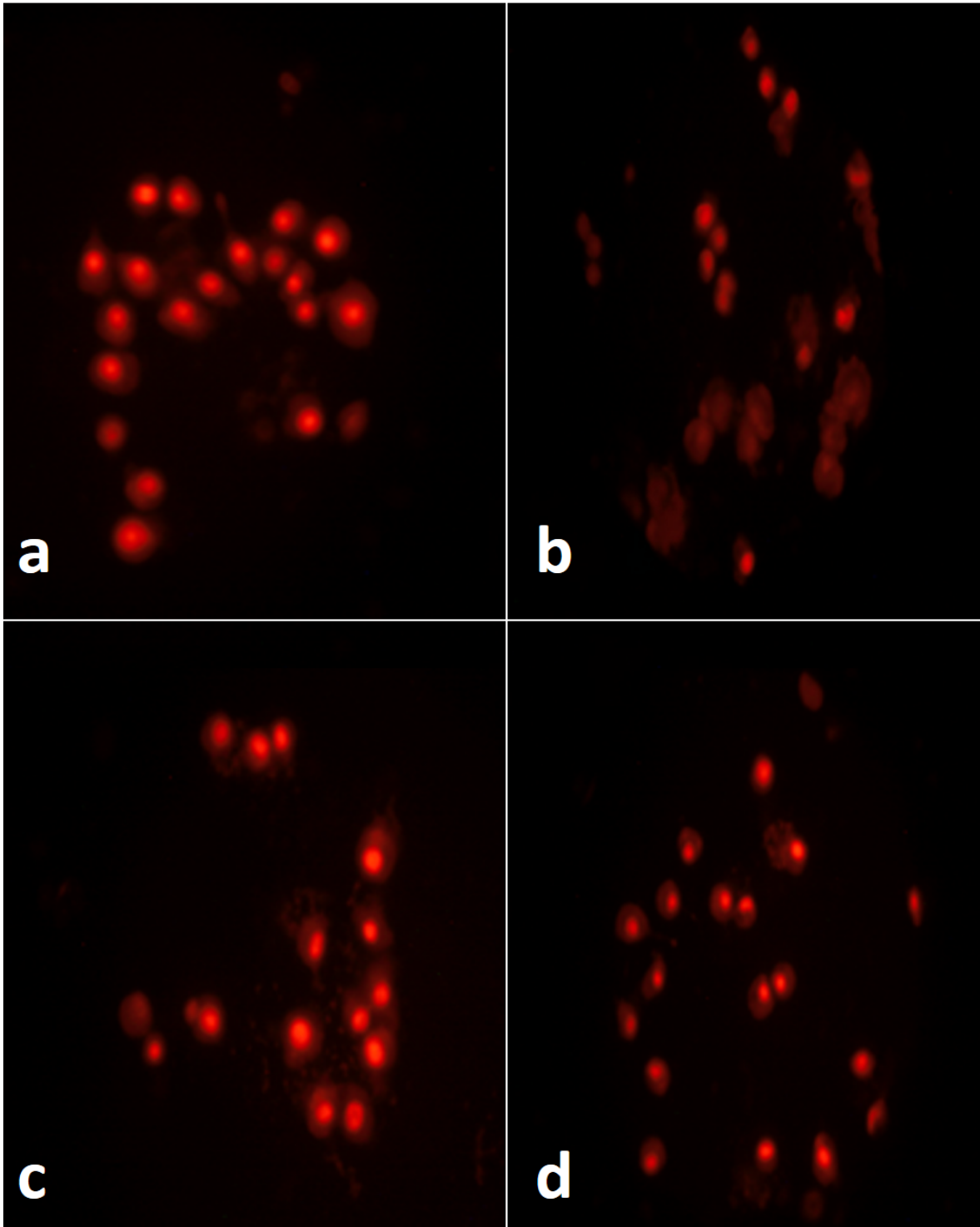
To determine whether these antioxidants could effect ROS production, a well-developed ROS assay was used. The results demonstrated that ROS production increased in comparison with non-treated prostate cancer cells (Figure 2a). Lipid peroxidation assay was used to assess oxidative stress by measuring the amount of malondialdehyde (MDA) in controls. The test results indicated a dose-dependent increase in MDA production. As shown in Figure 2b, the data revealed significant differences ( $p < 0.001$ ) in MDA production at 60  $\mu$ g/ml of  $\alpha$ -Toc,  $\beta$ -Crt and Asc compared to the control (0  $\mu$ g/ml).



**Figure 1.** The effect of  $\alpha$ -Toc,  $\beta$ -Crt and Asc in PC-3. Cell viability was examined by WST-1 assay. The data were expressed by mean  $\pm$  SEM ( $n \geq 3$ ). \*\*\* $p < 0.001$  Cont vs all antioxidants



**Figure 2.** The effect of  $\alpha$ -Toc,  $\beta$ -Crt and Asc on ROS (a) generation and LPO levels (b) in PC-3. Cells were exposed to antioxidants. Data were presented by mean  $\pm$  SEM. (n = 3).



**Figure 3.** The effect of  $\alpha$ -Toc (c),  $\beta$ -Crt (a) and Asc compared (d) to the control (0  $\mu$ g/ml) (a) on apoptosis evidenced by propidium staining.

#### **Propidium iodide Staining**

Propidium iodide staining of cellular nuclei was used as a marker for cell death during the 24 h. Cells are characterized by the typical nuclear modifications. In apop-

totic stages, the cells are characterized by a shrink in nucleus. Apoptotic cells were less colored as shown in (Figure 3)

### Caspase-3 Staining

To further investigate that the cell death indicated by propidium iodide staining was arisen from apoptotic cascade, immunohistochemical staining of the cells was carried for the active form of caspase 3—a downstream effector of apoptosis—using mouse monoclonal caspase-3 p11 antibody (diluted at 1:500; Santa Cruz Biotechnology, USA). The results indicate the presence of caspase 3 positive represented by black dot shown by arrows (Figure 4). This further demonstrates that the effect of these antioxidants is through apoptotic pathway.

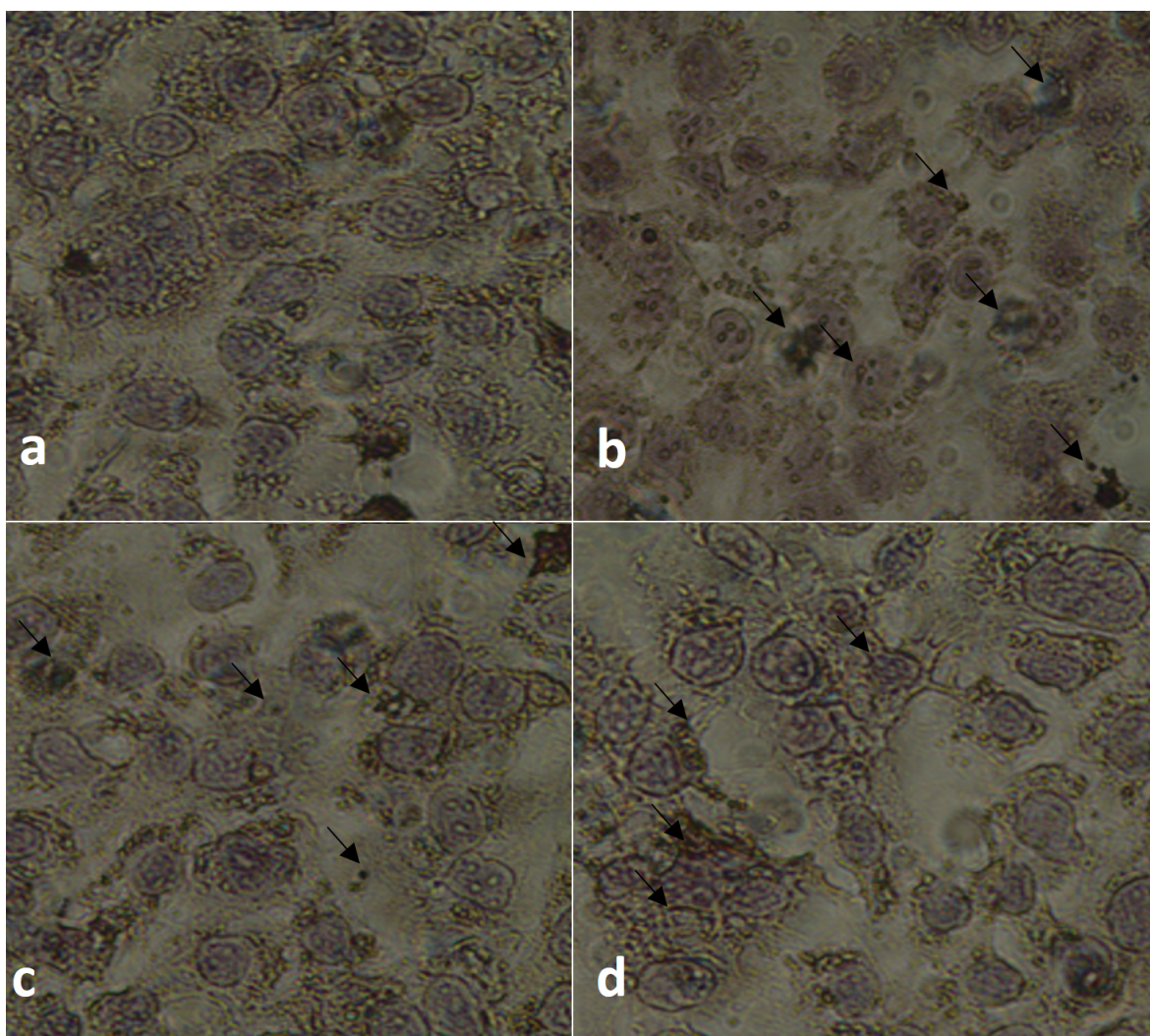
### DISCUSSION

Cancer, is one of the major causes of mortality in the world. Recently, it was reported that cancerous cells

are linked with high ROS levels, as a result of metabolic, genetic and microenvironment-dependent changes. ROS are generated through the metabolism of oxygen during mitochondrial aerobic respiration. High amount of ROS in cells damage or mutate the DNA of the cells [20].

Despite being studied widely, there has been no precise therapy for prostate cancer to date. Therefore, novel treatment strategies are needed. Regular consumption of natural flavonoids may be associated with a decreased risk of the cancers.

One of the most important antioxidant that fights against lipid peroxidation of cell membranes is Vitamin E. It can alter the oxidative stress biomarkers positively [21].



**Figure 4.** The effect of  $\alpha$ -Toc (c),  $\beta$ -Crt (a) and Asc compared (d) to the control (0  $\mu$ g/ml) (a) on apoptosis related active caspase-3 in PC-3. The cells were imaged under the inverted light microscope and caspase-3 expressing cells were clearly visible with their dark brown color.

Previous studies demonstrated that,  $\beta$ -CRT and retinol supplementation had no effect on the prostate cancer. It was reported that daily supplementation with selenium (200  $\mu$ g), vitamin E (400 IU) or both for decreased incidence of prostate cancer in men aged over 50 years. On the other hand, supplementation with 400 IU vitamin E every other day, 500 mg vitamin C every day or a combination of the two were reported to decrease the incidence of cancer in male US physicians aged 50 years or more [6, 22-24].

Recent studies reported that these antioxidants can be behaved as an antioxidant or prooxidant, depending on the redox potential of the biological environment in which it involves [25]. At low concentrations, the antioxidants are reported to act as an antioxidant, inhibiting free radical production, while at high concentrations, they presumably behave as a prooxidant, propagating free radical-induced reactions, consuming endogenous antioxidants and inducing DNA damage. Herein the focus of this study was causing oxidative stress and apoptosis of PC-3 by  $\alpha$ -Toc,  $\beta$ -Crt and Asc. It was found that these antioxidants led to an increase in the LPO marker MDA and ROS production at high concentration of the agents tested suggesting it behaves as a prooxidant in this study. ROS have high chemical activity and they play important roles in the regulation of cell proliferation and apoptosis. The level of ROS and lipid peroxidation was highly increased in PC3 cells in response to antioxidants tested within this study in comparison with untreated control cells, as shown in Figure 2 ( $p < 0.001$ ). These results suggest that  $\alpha$ -Toc,  $\beta$ -Crt and Asc induced mitochondrial dysfunction through the depolarization of the mitochondrial membrane potential and the generation of ROS, which resulted in lipid peroxidation in PC-3 cells. Apoptosis and necrosis are the two important types of cell death for which the molecular mechanisms have been widely studied [26]. Apoptosis, programmed cell death, is a normal developmental process that is characterized by nuclear condensation and cleavage of critical cellular proteins [27]. Apoptosis and necrosis represent two different mechanisms during cellular death. The dynamics of cellular lesions in these two processes are different. It was previously reported that plasma membrane damage, occurring as a primary event during necrosis represents, on the contrary, it is a delayed but massive phenomenon during apoptosis. One way to detect whether the antioxidants induced

apoptosis or necrosis is to carry out caspase 3 staining. Therefore, in this research, in addition to investigation of cell viability and ROS and LPO production in PC-3 cell line, the expression of apoptotic caspase-3 enzyme (protease enzyme that play a key role in apoptosis) was studied by immunohistochemical staining. As shown in Figure 4, treatment of PC-3 cells with  $\alpha$ -Toc,  $\beta$ -Crt and Asc displayed a marked increase in the expression level of caspase-3 suggesting the elevation in apoptotic enzyme by the antioxidants may account for the anti-apoptotic effect.

## CONCLUSION

This study investigated *in vitro* anticancer effect of  $\alpha$ -Toc,  $\beta$ -Crt and Asc on PC-3 prostate cancer cells by cell proliferation, ROS and Lipid Peroxidation assay, caspase-3 and propidium iodide staining experiments. The results confirmed that these molecules behaved as prooxidant by decreasing cell viability and increasing ROS and LPO levels in PC-3 cells. It was found that they induced apoptotic pathway as they increased expression level of apoptotic gene caspase-3.

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