



Important Fungal Diseases in Medicinal and Aromatic Plants and Their Control

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ABSTRACT

Aromatic plants constitute the main raw materials of the perfumery, food and cosmetics industry and in recent years with the increasing demand for therapeutic herbal medicines, interest in medicinal and aromatic plants has increased. Raw materials from medicinal and aromatic plants have recently begun to be used and spread in the food sector, especially in industrial sectors such as paint and perfumery. For this reason, growing healthy plant material is very important in terms of the protection of these crops. However, fungal diseases such as root rot, wilt, leaf spots, blight and anthracnose, which are problems during the cultivation of both medicinal and aromatic plants, negatively affect both the quantity and quality of these plants. For this reason, an integrated management practices including cultural measures, herbal products, biological control and, if necessary, chemical control methods with especially these fungal diseases are very important. In this review, 27 medicinal and aromatic plants, 37 fungal diseases, their chemical and biological control were included, and 161 references were used.

RESEARCH ARTICLE

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INTRODUCTION

The term ‘Herbal treatment’ has been used in many different countries with many names such as traditional therapy, complementary therapy, or natural therapy. The first records about herbal therapy were found in the Mesopotamian civilization in 5000 BC and it was determined that 250 herbal drugs; obtained from medicinal and aromatic plants as usually dried and sometimes fresh, whole, shredded or cut plant or plant parts were used (Demirezer, 2010). According to the World Health Organization (WHO), traditional medicine is used in the prevention of physical and mental diseases, diagnosis, healing, or treatment as well as maintaining good health. In addition, it is the whole of knowledge, skills and practices that can or cannot be explained based on theories, beliefs, and experiences specific to different cultures (WHO, 2017).

The therapeutic use of plants varies according to the development level of the countries. In developing countries, 80% of the population benefits from herbal products for therapeutic purposes. While this rate reaches up to 95% in some countries of Asia, Africa, and the Middle East, it is less in developed countries (40-50% in Germany, 42% in the USA, 48% in Australia, and 49% in France). However, the most important trade centres for medicinal plants are in Germany, USA, Japan, and England ([Titz, 2004](#)). World Health Organization predicts that the treatment with herbs will increase all over the world in the coming years.

According to the World Health Organization, 25% of the pharmaceutical drugs used today are manufactured from medicinal plants and 30% of drugs sold worldwide contain compounds derived from plant materials ([FAO, 2005](#)).

The use of herbal medicine in countries that apply traditional medical treatment varies according to the recommendations of the people practicing traditional/alternative medicine or their own experience. Also, in some countries, training is provided at universities related to complementary medicine. For example, in the universities of many countries in the Economic Community of West African States, such as Democratic Republic of Congo, South Africa and Tanzania there are complementary medicine courses in the curricula of pharmacy and medical studies ([WHO, 2014](#)). Complementary medicine is seen as a primary health care service in some African countries. For example, the ratio of traditional healers to the population in Africa is 1/40.000, while the ratio of medical doctors is 1/500 ([Abdullahi, 2011](#)).

Medicinal and aromatic plants are plants that have many uses such as food, medicine, cosmetics and spices and are known to have been used for similar purposes since the beginning of human history. While some of these plants are collected from nature, some of them have been cultivated and produced. However, most of the herbs used for therapeutic purposes are collected from nature. The most prominent and researched properties of medicinal and aromatic plants are their therapeutic uses ([Kumar, 2014](#)). Extracts of these plants in water or alcohol are also applied against pests and plant diseases due to their biological effects ([Isman, 2000](#); [Bakkali et al., 2008](#)). The aromatic parts of aromatic plants are used to extract therapeutic oils/essential oils containing allelochemical aroma of economic value ([Nagpal and Karki, 2004](#)).

More than 6 000 medicinal plant species have been identified from different tropical regions ([Khare, 2008](#)), more than 1 000 of them are classified as aromatic ([Panda, 2015](#)). Climate changes, intensive cultivation practices and market-oriented crop management have led to an increase in pests and diseases. These problems in medicinal and aromatic products have gradually increased ([Sharma, 2013](#); [Sharma et al., 2014](#)). Damages caused by pests or diseases can reduce their biomass and oil content ([Gupta et al., 2000](#); [Zadotani and Ikegami, 2002](#)). Besides changing climatic conditions, the indiscriminate and unplanned large-scale cultivation of medicinal and aromatic plants to meet the increasing demand of the pharmaceutical industries is leading to increased incidence and severity of diseases.

Losses caused by plant diseases not only reduce the yield of plant secondary metabolites but also reduce the quality of raw materials ([Singh et al., 2016](#)).

In this review, fungal diseases causing an infection on medicinal and aromatic plants, grown or cultured spontaneously and reducing plant quality and yield and even causing death, their symptoms and methods of controlling them are included.

In this context, diseases and management of important medicinal and aromatic plants listed in Table 1 and reported from Turkey and the world were presented.

Lavandula spp. (Lavender), *Humulus lupulus* L. (Hops), *Papaver somniferum* L. (Poppy), *Rosa* spp. (Rose), *Salvia officinalis* L. (Sage), *Origanum* spp. (Oregano), *Carthamus tinctorius* L. (Safflower), *Dianthus caryophyllus* L. (Clove), *Sesamum indicum* Linn. (Sesame), *Pimpinella anisum* L. (Anise), *Asparagus* spp. (Asparagus fern), *Rosmarinus officinalis* L. (= *Salvia rosmarinus*) (Rosemary), *Mentha piperita* L. (Mint), *Aloe vera* L. Burm. (= *Aloe barbadensis* Mill.), *Withania somnifera* (L.) Dunal (Indian Ginseng-Poisonous Gooseberry-Winter Cherry), *Rauwolfia serpentina* (L.) Benth. ex Kurz (Snakeroot) (Serpentine/Sarpagandha), *Ocimum sanctum* L. (= *Ocimum tenuiflorum* L.) (Holy Basil, Tulsi), *Coleus forskohlii* Briq (Coleus flower, *Chlorophytum borivilianum* Santapau & Fernandez (Musli), *Hyoscyamus* spp. (Henbane), *Plantago ovata* Forssk. (Psyllium Blond), *Catharanthus roseus* (L.) G. Don (Pink periwinkle), *Pogostemon cablin* (Blanco) Benth. (Patchouli), *Zingiber officinale* Roscoe (Ginger), *Vetiveria zizanioides* (L.) Nash (Vetiver), *Santalum* spp. (Sandalwood) and *Cymbopogon citratus* Stapf. (Lemon grass). Fungal disease agents that cause leaf spots, blight, rust, powdery mildew, root rot, damping-off and dieback frequently occur in these medicinal and aromatic plants (Avan, 2021).

OCCURRENCE OF FUNGAL DISEASES

The cultivation of medicinal and aromatic plants has increased considerably in recent years due to their huge worldwide demands on plant-based medicines and aromatic compounds. These plants are affected by various diseases caused by fungi, bacteria, viruses and phytoplasmas. Among these diseases, especially fungal diseases are very important (Table 1).

Fungi infect leaves, stems and underground parts of medicinal and aromatic plants. Among these fungal diseases, Powdery mildew appears on the leaves and fresh stems, and as the disease progresses, it covers the entire developing surface of the plant. Rust diseases are airborne diseases that infect leaves, branches and fruits and cause pustules on leaves. Leaf spots and blights cause dead areas on the leaves with distinct spots over time, and in this way, they are separated from healthy tissues (Bhandari et al., 2014). Blights appear on the leaves, twigs or blossoms of the plant and cause sudden death of the plant (Sattar et al., 2006; Ramappa and Shivanna, 2013). Medicinal and aromatic plants are also highly affected by root rot, wilt, anthracnose and dieback caused by fungi and bacteria. These diseases manifest themselves with hard, dry, spongy, soft, watery or slimy-looking rotten tissues in plants (Singh et al., 2016).

Table 1. Important fungal diseases of medicinal and aromatic plants.

DISEASES	PLANTS	CAUSAL ORGANISMS	SYMPTOMS
Rusts	<i>Aloe vera</i>	<i>Phakopsora pachyrhizi</i> , <i>Uromyces aloes</i>	Yellowish red spots appear on the lower surface of the leaf and rust pustules form in these spots (Jones, 1972; Koike et al., 1998; Kalra et al., 2005; Saber et al., 2009; Soni et al., 2011; Afshan et al., 2012).
	<i>Asparagus</i> spp. (Asparagus fern)	<i>Puccinia asparagi</i>	
	<i>Cymbopogon citratus</i> (Lemon grass)	<i>Puccinia nakanishikii</i>	
	<i>Mentha</i> spp. (Mint)	<i>Puccinia menthae</i>	

	<i>Pelargonium</i> spp. (Geranium)	<i>Puccinia pelargonii-zonalis</i>	
	<i>Pimpinella anisum</i> (Anise)	<i>Puccinia pimpinellae</i>	
	<i>Rosa</i> spp. (Rose)	<i>Caecoma</i> spp.	
	<i>Dianthus caryophyllus</i> (Clove)	<i>Uromyces dianthi</i>	
	<i>Origanum vulgare</i> (Oregano)	<i>Puccinia menthae</i>	
Downy Mildews	<i>Coleus forskohlii</i> (Coleus flower)	<i>Peronospora belbahrii</i> , <i>P. lamii</i>	Yellow to light brown necrotic lesions, folds, and kinks occur on the leaves (Sain and Sharma, 1999 ; Garibaldi et al., 2004 ; Landa et al., 2005 ; Humphreys-Jones et al., 2008 ; López-Guisa et al., 2013).
	<i>Ocimum sanctum</i> (Holy Basil, Tulsi)	<i>Peronospora belbahrii</i>	
	<i>Plantago ovata</i> (Psyllium Blond)	<i>Peronospora plantaginis</i>	
	<i>Humulus lupulus</i> (Hops)	<i>Pseudoperonospora humuli</i>	
	<i>Papaver somniferum</i> (Poppy)	<i>Peronospora</i> spp.	
	<i>Rosa</i> spp. (Rose)	<i>Peronospora sparsa</i>	
	<i>Rosmarinus officinalis</i> (Rosemary)	<i>Peronospora lamii</i>	
Powdery Mildew	<i>Mentha</i> spp. (Mint)	<i>Erysiphe cichoracearum</i>	Chlorotic spots and brownish discolorations in the form of powder appear on the leaf surface. The disease causes the leaves to curl and the bend to stem (Valiyeva et al., 2004 ; Kalra et al., 2005 ; Humphreys-Jones et al., 2008 ; Thines et al., 2009 ; Baradaran et al., 2012 ; Venegas-Portilla et al., 2020).
	<i>Cymbopogon citratus</i> (Lemon grass)	<i>Erysiphe graminis</i>	
	<i>Humulus lupulus</i> (Hops)	<i>Podosphaera macularis</i>	
	<i>Rosa damascena</i> (Damask rose)	<i>Podosphaera pannosa</i>	
	<i>Salvia officinalis</i> (Sage)	<i>Golovinomyces neosalviae</i> <i>Peronospora lamii</i>	
	<i>Rosa</i> spp. (Rose)	<i>Sphaerotheca pannosa</i> var. <i>rosae</i>	
Alternaria Leaf Spots	<i>Aloe vera</i>	<i>Alternaria alternata</i> <i>A. brassicae</i>	Dark brown circular spots occur on infected leaves (Xiaovin, 1982 ; Kumar et al., 1984 ; Kishore et al., 1985 ; Kalra et al., 2005 ; Taba et al., 2009 ; Garibaldi et al., 2011 ; Zimowska, 2015).
	<i>Withania somnifera</i> (Indian Ginseng)	<i>Alternaria alternata</i> , <i>A. tenuis</i>	
	<i>Mentha</i> spp. (Mint)	<i>Alternaria alternata</i>	
	<i>Pelargonium</i> spp. (Geranium)	<i>Alternaria alternata</i>	
	<i>Salvia officinalis</i> (Sage)	<i>Alternaria alternata</i>	
	<i>Ocimum basilicum</i> (Sweet Basil)	<i>Alternaria alternata</i>	
	<i>Hyoscyamus</i> spp. (Henbane)	<i>Alternaria alternata</i>	
	<i>Carthamus tinctorius</i> (Safflower)	<i>Alternaria carthami</i>	
	<i>Papaver somniferum</i> (Poppy)	<i>Alternaria alternata</i>	
	<i>Dianthus caryophyllus</i> (Clove)	<i>Alternaria dianthi</i>	
<i>Origanum vulgare</i> (Oregano)	<i>Alternaria alternata</i>		

Cercospora Leaf Spots	<i>Ocimum sanctum</i> (Holy Basil- Tulsi)	<i>Cercospora ocimicola</i>	Necrotic spots with dark brown edges are scattered on the leaves (Bubak, 1906 ; Enikuomehin, 2006 ; Bhandari et al., 2014).
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Cercospora rauwolfiae</i> , <i>Cercospora serpentinae</i>	
	<i>Emblica officinalis</i> (Indian Gooseberry)	<i>Cercospora</i> spp.	
	<i>Sesamum indicum</i> (Sesame)	<i>Cercospora sesami</i>	
	<i>Pimpinella anisum</i> (Anise)	<i>Cercospora malkoffii</i>	
Colletotrichum Leaf Spots	<i>Ocimum sanctum</i> (Holy Basil- Tulsi)	<i>Colletotrichum gloeosporioides</i> , <i>C. capsica</i>	Reddish brown circular spots appear on the leaves first. Holes and premature drying occur in the leaves due to the rupture of the infected tissue (Tekade et al., 2009 ; Gautam, 2014 ; Zimowska, 2015).
	<i>Withania somnifera</i> (Indian Ginseng)	<i>Colletotrichum gloeosporioides</i> , <i>C. dematium</i>	
	<i>Pelargonium</i> spp. (Geranium)	<i>C. gloeosporioides</i>	
	<i>Origanum vulgare</i> (Oregano)	<i>Colletotrichum fuscum</i>	
Corynespora Leaf Spot	<i>Coleus forskohlii</i> (Coleus flower)	<i>Corynespora cassiicola</i>	Yellowish-brown necrotic spots in the form of chlorotic halo occur on the leaves (Shukla et al., 2000 ; Garibaldi et al., 2007).
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>C. cassiicola</i>	
	<i>Mentha arvensis</i> (Menthol mint)	<i>C. cassiicola</i>	
	<i>Ocimum basilicum</i> (Sweet Basil)	<i>C. cassiicola</i>	
Curvularia Leaf Spots	<i>Cymbopogon citratus</i> , <i>C. flexuosus</i> (Lemon grass)	<i>Curvularia andropogonis</i> ,	Small oval and long dark brown necrotic lesions appear on the leaves (Thaung, 2008 ; Bhagat et al., 2014).
	<i>Mentha</i> spp. (Mint)	<i>C. lunata</i> ,	
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>C. trifolii</i>	
Diplocarpon Leaf Spot	<i>Rosa chinensis</i> (China rose), <i>Rosa damascena</i> (Isparta rose)	<i>Diplocarpon rosae</i>	Brown to black spots with dark purplish margins occur on the upper surface of the leaves (Margina and Zheljzkov, 1995a).
Macrophomina Leaf Spot	<i>Chlorophytum borivilianum</i> (Musli)	<i>Macrophomina phaseolina</i>	Small water-soaked lesions are formed surrounded by a dark brown border on the leaves (Dadwal and Bhartiya, 2012).
Myrothecium Leaf Spot	<i>Withania somnifera</i> (Indian Ginseng)	<i>Myrothecium roridum</i>	Small, dull yellow, brown coloured water-soaked spots appear on the leaves (Shivanna et al., 2014).
Stemphylium Leaf Spots	<i>Asparagus</i> spp. (Asparagus fern)	<i>Stemphylium vesicarium</i>	Light brown, large elliptical spots occur on the stem and branches, the lesions on the stems merge into large areas of infected tissue (Falloon and Tate, 1986 ; Zimowska, 2015).
	<i>Origanum vulgare</i> (Oregano)	<i>Stemphylium botryosum</i>	
Phoma Leaf Spot	<i>Origanum vulgare</i> (Oregano)	<i>Phoma herbarum</i>	The agent causes angular spots on the leaves. (Basavand et al., 2020).
Alternaria Leaf Blights	<i>Chlorophytum borivilianum</i> (Musli)	<i>Alternaria alternata</i>	Brown necrotic irregular lesions and surrounding

	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Alternaria tenuis</i> , <i>A. alternata</i>	chlorotic halos occur on the leaves (Rai and Tetrawal, 2010 ; Thakur and Harsh, 2014).
	<i>Mentha</i> spp. (Mint)	<i>Alternaria alternata</i>	
	<i>Pelargonium</i> spp. (Geranium)	<i>Alternaria alternata</i>	
	<i>Withania somnifera</i> (Indian Ginseng)	<i>Alternaria tenuis</i> , <i>A. alternata</i>	
	<i>Plantago ovata</i> (Psyllium Blond)	<i>Alternaria alternata</i>	
Colletotrichum Leaf Blights	<i>Chlorophytum borivilianum</i> (Musli)	<i>Colletotrichum dematium</i> , <i>C. capsici</i>	Small chlorotic spots appear on the lower leaves, which rapidly expand and merge into brown spots (Sattar et al., 2006 ; Ramappa and Shivanna, 2013).
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Colletotrichum dematium</i> , <i>C. capsici</i>	
	<i>Cymbopogon citratus</i> (Lemon grass)	<i>Colletotrichum caudatum</i>	
Curvularia Leaf Blights	<i>Cymbopogon citratus</i> (Lemon grass)	<i>Curvularia trifolii</i>	Long, reddish-brown necrotic lesions occur on the leaves (Alam et al., 1983 ; Sato and Ohkubo, 1990).
	<i>Cymbopogon nardus</i> (Lemon grass)	<i>Curvularia andropogonis</i>	
	<i>Vetiveria zizanioides</i> (Vetiver)	<i>Curvularia trifolii</i>	
Macrophomina Leaf Blights	<i>Chlorophytum borivilianum</i> (Musli)	<i>Macrophomina phaseolina</i>	Necrotic lesions appear on the edges and tips of infected leaves (Maiti and Geetha, 2013 ; Meena and Kadam, 2021).
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Macrophomina phaseolina</i>	
Sclerotinia Blights	<i>Mentha</i> spp. (Nane)	<i>Sclerotinia sclerotiorum</i>	The first symptoms are necrosis of the stem, darkening and wilting of the leaves. Then cottony soft rots occur (Garibaldi et al., 2013).
Rhizoctonia Leaf Blights	<i>Coleus forskohlii</i> (Coleus flower)	<i>Rhizoctonia solani</i>	Water-soaked irregular spots spreading inward from the leaf edge are formed (Mehrotra and Thapar, 1990 ; Shukla et al., 1993 ; Kalra et al., 2005 ; Sato et al., 2010 ; Aktaruzzaman et al., 2015).
	<i>Mentha</i> spp. (Mint)	<i>Rhizoctonia solani</i>	
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Rhizoctonia solani</i>	
	<i>Rosmarinus officinalis</i> (Rosemary)	<i>Rhizoctonia solani</i>	
	<i>Origanum vulgare</i> (Oregano)	<i>Rhizoctonia solani</i>	
Botrytis Leaf Blights	<i>Pelargonium</i> spp. (Geranium)	<i>Botrytis cinerea</i>	Concentric ring lesions on leaves, wilting and drying of flowers appeared (Kalra et al., 2008 ; Vinodkumar and Nakkeeran, 2017).
	<i>Rosa chinensis</i> , (China rose)	<i>Botrytis cinerea</i>	
	<i>Rosa damascena</i> (Isparta rose)	<i>Botrytis cinerea</i>	
	<i>Dianthus caryophyllus</i> (Clove)	<i>Botrytis cinerea</i>	
Passalora Blight	<i>Pimpinella anisum</i> (Anise)	<i>Passalora malkoffii</i>	The disease causes lesions and drying on all above-ground parts of plants, including inflorescences (Erzurum et al., 2005).
Phoma Leaf Blight	<i>Origanum vulgare</i> (Oregano)	<i>Phoma multirostrata</i> var. <i>macrospora</i>	Small, black spots are observed on the top and bottom of the infected leaves and on

			young shoots depending on the humidity (Garibaldi et al., 2015b)
Anthracnoses	<i>Aloe vera</i>	<i>Colletotrichum gloeosporioides</i>	Small necrotic spots on the leaves turn into typical anthracnose lesions as the disease progresses (Sattar et al., 2002 ; Singh et al., 2004 ; Ayvar-Serna et al., 2020)
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Colletotrichum gloeosporioides</i>	
	<i>Emblica officinalis</i> (Indian Gooseberry)	<i>Colletotrichum gloeosporioides</i>	
	<i>Mentha</i> spp. (Nane)	<i>Sphaceloma menthae</i>	
	<i>Salvia officinalis</i> (Sage)	<i>Colletotrichum dematium</i>	
	<i>Origanum vulgare</i> (Oregano)	<i>Colletotrichum tropicale</i>	
Diebacks	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Colletotrichum dematium</i>	Fading and drying appears in the tip buds of young branches. (Kulkarni and Ravindra, 1988 ; Kulkarni et al., 1992).
	<i>Catharanthus roseus</i> (Pink Periwinkle)	<i>Pythium aphanidermatum</i>	
Collar Rots	<i>Cymbopogon citratus</i> (Lemon grass)	<i>Fusarium moniliforme</i>	Chlorosis on the lower leaves and small brown necrotic lesions in the collar area of the plant occurs (Singh et al., 2001 ; Trivedi et al., 2006).
	<i>Mentha</i> spp. (Mint)	<i>Sclerotium rolfsii</i>	
	<i>Pogostemon cablin</i> (Patchouli)	<i>Fusarium oxysporum</i> , <i>Rhizoctonia solani</i>	
	<i>Chlorophytum borivilianum</i> (Musli)	<i>Corticium rolfsii</i>	
Fruit Rots	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Rhizopus stolonifer</i>	Wet rots appear on the fruit (Shukla et al., 2006 ; Singh et al., 2011).
	<i>Withania somnifera</i> (Indian Ginseng)	<i>Myrothecium</i> sp.	
	<i>Emblica officinalis</i> (Indian Gooseberry)	<i>Phomopsis phyllanthi</i>	
Leaf Rots	<i>Aloe vera</i>	<i>Sclerotium rolfsii</i> , <i>Colletotrichum dematium</i> , <i>Phoma</i> sp., <i>Rhizoctonia bataticola</i>	Water-soaked spots occur on the leaves (Shukla et al., 1981).
Rhizome Rot	<i>Zingiber officinale</i> (Ginger)	<i>Pythium aphanidermatum</i>	Hard or spongy yellowish brown to brown tissues formations appear (Stirling et al., 2009).
Stolon Rots	<i>Mentha</i> spp. (Mint)	<i>Macrophomina phaseoli</i> , <i>Rhizoctonia solani</i> , <i>R. bataticola</i> , <i>Thielavia basicola</i>	The agent typically causes fading in the stolons and rot in later stages (Kalra et al., 2008).
Root Rots	<i>Aloe vera</i>	<i>Fusarium oxysporum</i> <i>Phytophthora</i> spp., <i>Pythium</i> spp.	With the wilting of the plants, yellowing of the leaves, falling and white cotton-like mycelium growth in the collar area appear (Subbiah et al., 1996 ; Boby and Bagyaraj, 2003 ; Kamalakaran et al., 2006 ; Zimowska, 2008 ; Martini et al., 2009 ; Govindappa et al., 2010 ; Ziedan et al., 2010 ; Zimowska, 2015 ; Ağaner and Cere, 2017).
	<i>Asparagus</i> spp. (Asparagus fern)	<i>Fusarium oxysporum</i> f.sp. <i>asparagi</i> , <i>F. proliferatum</i> , <i>F. moniliforme</i> , <i>F. solani</i> , <i>F. redolens</i> , <i>Phytophthora asparagi</i> , <i>Phytophthora megasperma</i> var. <i>sojae</i> , <i>Phytophthora</i> spp., <i>Rhizoctonia solani</i>	

	<i>Origanum</i> spp. (Oregano)	<i>R. solani</i> , <i>M. phaseolina</i>	
	<i>Chlorophytum borivilianum</i> (Musli)	<i>Rhizoctonia bataticola</i> , <i>Fusarium solani</i>	
	<i>Coleus forskohlii</i> (Coleus flower)	<i>Fusarium chlamydosporum</i> , <i>F. solani</i> , <i>Macrophomina phaseolina</i> , <i>Ralstonia solanacearum</i>	
	<i>Ocimum sanctum</i> (Holy Basil- Tulsi)	<i>Rhizoctonia solani</i> , <i>Pythium</i> spp.	
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Macrophomina phaseolina</i>	
	<i>Pelargonium</i> spp. (Geranium)	<i>Rhizoctonia solani</i> , <i>Macrophomina phaseolina</i> <i>Pythium</i> sp.	
	<i>Rosa chinensis</i> (China rose), <i>Rosa damascena</i> (Isparta rose)	<i>Phytophthora</i> spp., <i>Alternaria</i> spp., <i>Rhizoctonia</i> spp., <i>Sclerotinia</i> spp., <i>Pythium</i> spp.	
	<i>Salvia officinalis</i> (Sage)	<i>Phytophthora cryptogea</i> , <i>Rhizoctonia solani</i> , <i>Fusarium</i> spp., <i>Fusarium oxysporum</i> , <i>Phoma exigua</i> var. <i>exigua</i>	
	<i>Lavandula</i> spp. (Lavender)	<i>Phytophthora nicotianae</i> , <i>P. palmivora</i> , <i>P. cinnamomi</i> , <i>P. cactorum</i>	
	<i>Carthamus tinctorius</i> (Safflower)	<i>Macrophomina phaseolina</i>	
	<i>Sesamum indicum</i> (Sesame)	<i>Macrophomina phaseolina</i>	
	<i>Origanum vulgare</i> (Oregano)	<i>Phytophthora tentaculata</i>	
	<i>Origanum dubium</i> (Oregano)	<i>Boeremia exigua</i> var. <i>exigua</i> , <i>Rhizoctonia solani</i> , <i>Fusarium</i> spp.	
Stem Rots	<i>Asparagus</i> spp. (Asparagus fern)	<i>Fusarium oxysporum</i> f.sp. <i>asparagi</i> , <i>F. proliferatum</i> , <i>F. moniliforme</i> , <i>F. solani</i> , <i>F. redolens</i>	Pale green water-soaked lesions appear. The fleshy tissue becomes weak and the water in the tissue comes out quickly, a slight odor is felt and this part turns brown (Trujillo et al., 1988 ; Burns and Benson, 2000 ; Elena, 2006 ; Oogi et al., 2009 ; Martini et al., 2009 ; Zimowska, 2015 ; Samouel et al., 2016)
	<i>Lavandula</i> spp. (Lavender)	<i>Phytophthora nicotianae</i> , <i>P. palmivora</i> , <i>P. cinnamomi</i> , <i>P. cactorum</i>	
	<i>Salvia officinalis</i> (Sage)	<i>Phomopsis sclarea</i>	
	<i>Rosmarinus officinalis</i> (Rosemary)	<i>Sclerotinia sclerotiorum</i>	
	<i>Dianthus caryophyllus</i> (Clove)	<i>Rhizoctonia solani</i>	
	<i>Origanum vulgare</i> (Oregano)	<i>Phytophthora tentaculata</i>	

	<i>Origanum dubium</i> (Oregano)	<i>Boeremia exigua</i> var. <i>exigua</i> , <i>Rhizoctonia solani</i> , <i>Fusarium</i> spp.	
Crown Rots	<i>Asparagus</i> spp. (Asparagus fern)	<i>Phytophthora asparagi</i> , <i>Phytophthora megasperma</i> var. <i>sojae</i> , <i>Phytophthora</i> spp.	Infected crowns first turn yellowish orange and as the disease progresses, rots appear (Garibaldi et al., 2015a ; Mondal et al., 2018).
	<i>Rosa chinensis</i> (China rose), <i>Rosa damascena</i> (Isparta rose)	<i>Phytophthora</i> spp., <i>Alternaria</i> spp., <i>Rhizoctonia</i> spp., <i>Sclerotinia</i> spp., <i>Pythium</i> spp.	
	<i>Papaver somniferum</i> (Poppy)	<i>Pleospora papaveracea</i>	
	<i>Salvia officinalis</i> (Sage)	<i>Phytophthora cryptogea</i>	
Wilts	<i>Coleus forskohlii</i> (Coleus flower)	<i>Ralstonia solanacearum</i>	With the fading and falling of the plants, cottony growths appear around the main root. (Nelson et al., 1960 ; Gupta et al., 2004 ; Dung et al., 2010 ; Ziedan et al., 2010).
	<i>Ocimum sanctum</i> (Holy Basil- Tulsi)	<i>Fusarium oxysporum</i> f. sp. <i>basilicum</i>	
	<i>Withania somnifera</i> (Indian Ginseng)	<i>Fusarium solani</i>	
	<i>Emblica officinalis</i> (Indian Gooseberry)	<i>Fusarium</i> sp.	
	<i>Vetiveria zizanioides</i> (Vetiver)	<i>Fusarium</i> sp.	
	<i>Plantago ovata</i> (Psyllium Blond)	<i>Fusarium oxysporum</i> , <i>F. solani</i>	
	<i>Mentha</i> spp. (Mint)	<i>Verticillium albo-atrum</i> var. <i>menthae</i>	
	<i>Pelargonium</i> spp. (Geranium)	<i>Verticillium albo-atrum</i> , <i>Verticillium dahliae</i>	
	<i>Humulus lupulus</i> (Hops)	<i>Verticillium nonalfalfae</i> , <i>Verticillium albo-atrum</i> ,	
	<i>Lavandula</i> spp. (Lavender)	<i>Fusarium sporotrichioides</i> , <i>F. oxysporum</i> , <i>F. solani</i> , <i>Sclerotinia sclerotiorum</i> , <i>Phytophthora palmivora</i>	
	<i>Rosmarinus officinalis</i> (Rosemary)	<i>Phytophthora citrophthora</i> , <i>Rhizoctonia solani</i> , <i>Fusarium oxysporum</i> , <i>Nigrospora oryzae</i>	
	<i>Carthamus tinctorius</i> (Safflower)	<i>Fusarium oxysporum</i> f. sp. <i>carthami</i>	
	<i>Sesamum indicum</i> (Sesame)	<i>Fusarium oxysporum</i> f. sp. <i>sesami</i>	
<i>Dianthus caryophyllus</i> (Clove)	<i>Fusarium oxysporum</i> f.sp. <i>dianthi</i>		
Damping-off	<i>Withania somnifera</i> (Indian Ginseng)	<i>Rhizoctonia solani</i>	Infected seedlings first turn yellow and wilt, then the plant falls over and collapses (Kishore et al., 1985 ; Alam et al., 1996 ; Carkacı and Maden, 1998 ; Li et al., 2008 ; Barguil et al., 2009).
	<i>Cymbopogon citratus</i> (Lemon grass)	<i>Pythium aphanidermatum</i>	
	<i>Rosa chinensis</i> (China rose), <i>Rosa damascena</i> (Isparta rose)	<i>Phytophthora</i> spp., <i>Alternaria</i> spp., <i>Rhizoctonia</i> spp., <i>Sclerotinia</i> spp., <i>Pythium</i> spp.	

	<i>Santalum</i> spp. (Sandalwood)	<i>Fusarium</i> spp., <i>Phytophthora</i> spp., <i>Rhizopus</i> spp.	
	<i>Lavandula</i> spp. (Lavender)	<i>Rhizoctonia solani</i> , <i>Botrytis cinerea</i> , <i>Alternaria alternata</i> , <i>Colletotrichum</i> spp.	
	<i>Papaver somniferum</i> (Poppy)	<i>Fusarium solani</i>	
	<i>Sesamum indicum</i> (Sesame)	<i>Rhizoctonia solani</i> , <i>Fusarium</i> spp., <i>Alternaria tenuis</i>	
	<i>Pimpinella anisum</i> (Anise)	<i>Rhizoctonia solani</i> , <i>Fusarium</i> spp., <i>Alternaria tenuis</i>	
	<i>Rosa</i> spp. (Rose)	<i>Fusarium oxysporum</i>	
	<i>Dianthus caryophyllus</i> (Clove)	<i>Rhizoctonia solani</i>	
	<i>Salvia officinalis</i> (Sage)	<i>Fusarium oxysporum</i> , <i>F. solani</i> , <i>F. moniliforme</i> , <i>Rhizoctonia solani</i>	
Gray Mold	<i>Ocimum sanctum</i> (Holy Basil- Tulsi)	<i>Botrytis cinerea</i>	The agent causes dense gray-brown hairy growth on the stems and leaves, falling of leaves, damping-off of plants, severe lesions on the stem and death on the plant (Edney, 1967 ; Moreira et al., 2015).
	<i>Rosa chinensis</i> , <i>Rosa damascena</i> (China rose) (Isparta rose)	<i>Botrytis cinerea</i>	
	<i>Dianthus caryophyllus</i> (Clove)	<i>Botrytis cinerea</i>	
Blue Mold	<i>Emblica officinalis</i> (Indian Gooseberry)	<i>Penicillium citrinum</i> , <i>P. islandicum</i>	Soft, wet and colourless-looking spots occur on infected fruits. Blue green spores are appeared in these parts (Saini, 2017).
Wet Rots	<i>Withania somnifera</i> (Indian Ginseng)	<i>Choanephora cucurbitarum</i>	The infected area appears wet and these parts turn into signs of rot (Shukla et al., 2006)
	<i>Rauwolfia serpentina</i> (Snakeroot)	<i>Rhizopus stolonifer</i>	

CONTROL OF FUNGAL DISEASES

Essential oils obtained from aromatic herbs are used in the perfume and food industry. Therefore, healthy plant material is very important for maintaining product quality. However, there are major problems in the cultivation of both medicinal and aromatic plants. The damages caused by fungal diseases cause negative effects on both the quality and quantity of the plant's biomass and the limitation of its successful cultivation in large areas and different places. Chemical applications are a form of management that is often used by producers. However, toxic pesticide residues in chemicals cause serious concerns as they pose serious dangers to human health. For this reason, cultural practices, products obtained from plants and biological control methods have been used along with chemical control (Table 2).

The methods of cultural control against fungal diseases on medicinal and aromatic plants include proper field cleaning and irrigation, use of resistant varieties, use of

compost, mulch and fertilizers that strengthen plant growth, avoiding close planting, pruning regularly, removing diseased plants and destroying them are especially recommended. Avoiding close planting, pruning regularly, removing diseased plant debris and destroying them are especially recommended. Various studies show that the management of diseases with biological control is more effective in controlling multiple diseases.

By reducing the chemicals used in agriculture, it seems possible to obtain quality products by preventing the yield loss in the soil with organic and biological solutions, which are alternative methods of control ([Avan and Kotan, 2021](#)).

Table 2. Chemical and biological control methods reported on fungal diseases of medicinal and aromatic plants.

Fungal Diseases in Medicinal and Aromatic Plants	Chemical Control	Biological Control
Rusts	<ul style="list-style-type: none"> - Sulfur, Copper oxychloride (Singh, 2006). - Chlorothalonil^{1,2,6} (Douglas, 2003; Moorman, 2017) - Azoxystrobin, Myclobutanil^{1,3}, Propiconazole^{1,2,3,4,7} (Mueller et al., 2004) - Tebuconazole^{5,7}+Triadimenol^{2,7}, Triadimenol^{2,7}, Flutriafol (Margina and Zhelezkov, 1995b) - Trifloxystrobin+ Tebuconazole^{5,7}, Propiconazole^{1,2,3,4,7} (Mekonnen and Manahlie, 2018) - Triadimefon^{2,7} (Tamuli et al., 2012) 	<ul style="list-style-type: none"> - <i>Bacillus subtilis</i> and <i>Trichoderma harizianum</i> (Saber et al., 2009). - <i>Datura stramonium</i>, <i>Maesa lanceolata</i> ve <i>Milletia ferruginea</i> extracts (Mekonnen et al., 2014). - <i>Vernonia amygdalina</i>, <i>Artemisia annua</i> (Mekonnen et al., 2015)
Downy Mildews	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6} (Jat et al., 2015) - Metalaxyl (Yadav et al., 2010) - Acibenzolar-S-methyl, Azoxystrobin, Cyazofamid, Mandipropamid (McGrath and LaMarsh, 2013, 2015) - Metalaxyl-M + Copper hydroxide², Mineral fertilizer "Alexin", Mandipropamid, Azoxystrobin, Glucohumates activator complex and Acibenzolar-S-methyl (Gilardi et al., 2013) 	<ul style="list-style-type: none"> - <i>Streptomyces lydicus</i>, <i>Bacillus amyloliquefaciens</i> strain D747, <i>Reynoutria sachalinensis</i> extract, neem oil, potassium bicarbonate and hydrogen dioxide (Wyenandt et al., 2015)
Powdery Mildews	<ul style="list-style-type: none"> - Sodium bicarbonate (Salamone et al., 2009). - Azoxystrobin, Boscalid⁷+Pyraclostrobin, Metalaxyl M+ Copper oxychloride, Mandipropamid and copper-based fungicides (Minuto et al., 2012) - Boscalid⁷, Monopotassium phosphate and vegetable oils (NTI 3404, NTI 3412) (Amoretti et al., 2005) 	<ul style="list-style-type: none"> - Thyme and clove essential oil (Salamone et al., 2009).
<i>Alternaria</i> Leaf Spots	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6} (Sharma et al., 2010a). - Bordeaux mixture (Smitha et al., 2014). - Propineb^{1,2} (Parashurama and Shivanna, 2013). - Propiconazole^{1,2,3,7}, Difenoconazole⁷ (Chauhan and Ravi, 2020). - Mancozeb^{1,2,3,6}+Propiconazole^{1,2,3,4,7} (DMAPR, 2012). - Penconazole (Qazi et al., 2006) 	<ul style="list-style-type: none"> - <i>Trichoderma viride</i> (Chauhan and Ravi, 2020). - Garlic oil, ginger oil and tulsi oil, turmeric rhizome extract (Sharma et al., 2010a) - Neem extract (Guleria and Kumar, 2006).

	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6}, Propiconazole^{1,2,3,4,7}, Difenconazole⁷, Azoxystrobin (Sharma et al., 2010a). - Benomyl^{1,2,7}, Mancozeb^{1,2,3,6}, Carbendazim^{2,7} (Singh, 2006) 	
Cercospora Leaf Spots	<ul style="list-style-type: none"> - Carbendazim^{2,7} (DMAPR, 2014) - Chlorothalonil^{1,2,6}, Iprodione^{1,2,7}, Copper oxychloride, Maneb^{1,2,3,6}, Mancozeb^{1,2,3,6}, Thiophanate-methyl^{1,2,6}, Benomyl^{1,2,7} (Singh, 2006). - Zineb^{1,2} (Mondal et al., 2018) 	- Soil application of neem cake + leaf waste of eucalyptus, <i>Millettia</i> (=Pongamia) pinnata+Madhuca longifolia cake; Neem oil or Neem seed extract+Neem cake and <i>Pseudomonas fluorescens</i> (Arumugam et al., 2010).
Colletotrichum Leaf Spots	<ul style="list-style-type: none"> - Tebuconazole^{5,7} (Sharma et al., 2010a,b). - Dithane (DMAPR, 2014). - Mancozeb^{1,2,3,6}, Copper oxychloride (Mondal et al., 2018) 	<ul style="list-style-type: none"> - <i>Trichoderma viride</i>, <i>T. harzianum</i>, <i>T. koningii</i>, <i>T. virens</i>, <i>T. hamatum</i> (Musheer and Ashraf, 2017). - Gentsyl alcohol obtained from <i>Phoma herbarum</i> (Gupta et al., 2016).
Corynespora Leaf Spot	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6} (DMAPR, 2014). 	- <i>Pseudomonas</i> sp.+Salicylic acid+ <i>Clerodendron inerme</i> leaf powder (DMAPR, 2014).
Curvularia Leaf Spots	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6}, Bordeaux mixture (Smitha et al., 2014) 	
Diplocarpon Leaf Spot	<ul style="list-style-type: none"> - Trifloxystrobin+Tebuconazole^{5,7} (IIHR, 2016). 	
Macrophomina Leaf Spot		<ul style="list-style-type: none"> - <i>T. viride</i> + <i>P. fluorescens</i> (Senthamarai et al., 2008) - <i>T. viride</i> and neem based product (Kulkarni et al., 2007)
Stemphylium Leaf Spot	<ul style="list-style-type: none"> - Maneb^{1,2,3,6}, Mancozeb^{1,2,3,6}, Chlorothalonil^{1,2,6}, Iprodione^{1,2,7} (Gindrat et al., 1984). - Mancozeb^{1,2,3,6}, Carbendazim^{2,7}, Propiconazole^{1,2,3,4,7} (Mondal et al., 2018) 	
Alternaria Blights	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6} (Sharma et al., 2010a; Jat et al., 2015). - Bordeaux mixture (Smitha et al., 2014). - Copper oxychloride, Carbendazim^{2,7} (Singh, 2006) - Mancozeb^{1,2,3,6}+Propiconazole^{1,2,3,4,7} (DMAPR, 2012). 	<ul style="list-style-type: none"> - <i>Ocimum sanctum</i>, <i>Zingiber officinale</i>, <i>A. sativum</i> or neem extracts and <i>Datura metel</i> or <i>Mentha spicata</i> extracts (Sharma et al., 2010a). - <i>T. asperellum</i> (Gatak et al., 2020)
Colletotrichum Blights	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6}, Carbendazim^{2,7}, Bordeaux mixture (Shukla et al., 2010; Smitha et al., 2014). - Hexaconazole⁷, Propiconazole^{1,2,3,4,7}, Tricyclazole, Thiophanate methyl^{1,2,6} and Carbendazim^{2,7} + Mancozeb^{1,2,3,6} (Kadam et al., 2014). 	
Curvularia Leaf Blights	<ul style="list-style-type: none"> - Mancozeb^{1,2,3,6}, Bordeaux mixture (Smitha et al., 2014). - Copper oxychloride (Mondal et al., 2018) - Propineb^{1,2}, Hexaconazole⁷ and Epoxiconazole^{1,2,6} (Lakpale, 2011). 	- Neem oil, <i>Kalanchoe heterophylla</i> , <i>Curcuma amada</i> and <i>Adhatoda vasica</i> extracts, <i>T. viride</i> and <i>P. fluorescens</i> (Lakpale, 2011).
Macrophomina Blight	<ul style="list-style-type: none"> - Metalaxyl + Mancozeb^{1,2,3,6} (Meena and Kadam, 2021) 	- <i>Pseudomonas fluorescence</i> (Meena and Kadam, 2021)
Sclerotinia Blight	<ul style="list-style-type: none"> - Tebuconazole^{5,7} (Sharma et al., 2010a, b) 	<i>Trichoderma harzianum</i> , <i>Gliocladium virens</i> (Mondal et al., 2018)

Rhizoctonia Blight	- Mancozeb ^{1,2,3,6} , Carbendazim ^{2,7} (Mondal et al., 2018)	- Trichoderma+ Organic fertilizer (Mondal et al., 2018)
Anthracnose	- Carbendazim ^{2,7} (Prakash, 2012), - Chlorothalonil ^{1,2,6} (Parameswaran et al., 2000). - Mancozeb ^{1,2,3,6} , Bordeaux mixture (Mondal et al., 2018)	
Collar Rot	- Carbendazim ^{2,7} , Thiophanate-methyl ^{1,2,6} (TNAU, 2013) - Mancozeb ^{1,2,3,6} (Mondal et al., 2018)	- <i>T. harzianum</i> (Singh and Singh, 2004).
Rhizome Rot	- Copper oxychloride, Mancozeb ^{1,2,3,6} , Carbendazim ^{2,7} (Mondal et al., 2018) - Tebuconazole ^{5,7} (Sharma et al., 2010a,b).	
Stolon Rot	- Mancozeb ^{1,2,3,6} , Carbendazim (Mondal et al., 2018) - Captan ^{1,6} (Szezeponek and Mazur, 2006)	- <i>Trichoderma harzianum</i> , <i>Gliocladium virens</i> (Mondal et al., 2018) - <i>T. viride</i> , <i>P. fluorescens</i> and <i>B. subtilis</i> (Kamalakaran et al., 2003)
Root Rots	- Mancozeb ^{1,2,3,6} , Copper oxychloride (Mondal et al., 2018) - Carbendazim ^{2,7} (DMAPR, 2006) - Carbendazim ^{2,7} +Mancozeb ^{1,2,3,6} (Ingle et al., 2014)	- <i>Trichoderma harzianum</i> (Govindappa et al., 2010) - <i>T. viride</i> + <i>P. fluorescens</i> (Ingle et al., 2014). - <i>T. viride</i> (DMAPR, 2006) - <i>P. fluorescens</i> (Govindappa et al., 2010; Ingle et al., 2014) - <i>Glomus fasciculatum</i> , <i>G. mosesae</i> (Mondal et al., 2018). - <i>T. viride</i> , <i>P. fluorescens</i> , <i>Bacillus subtilis</i> , Neem cake and Mahua cake, <i>T. viride</i> + Neem cake (Gnanaprakash et al., 2015). - <i>Allium schoenoprasum</i> , <i>Annona squamosa</i> , <i>A. indica</i> , <i>Calendula officinalis</i> , <i>Cinnamomum verum</i> , <i>Eucalyptus</i> sp., <i>Lawsonia inermis</i> , <i>O. sanctum</i> , <i>Piper nigrum</i> , <i>Z. officinale</i> aqueous extract sprays or extracts in 50% ethanol (Chathuri et al., 2011) - <i>Bacillus subtilis</i> (Elewa et al., 2011) - Neem seed powder+Carbofuran, Carbofuran+Carbendazim, Neem seed powder + Carbendazim (Kahkashan, 2003)
Stem Rot	- Carbendazim ^{2,7} , Mancozeb ^{1,2,3,6} (Mondal et al., 2018)	- <i>T. viride</i> , <i>P. fluorescens</i> and <i>B. subtilis</i> (Kamalakaran et al., 2003) - <i>Trichoderma</i> spp., <i>Glomus fasciculatum</i> and <i>G. mosesae</i> (Mondal et al., 2018)
Crown Rot	- Carbendazim ^{2,7} , Mancozeb ^{1,2,3,6} (Mondal et al., 2018)	- <i>Trichoderma</i> spp. (Mondal et al., 2018)
Wilts	- Carbendazim ^{2,7} (Singh, 2006; Bhat et al., 2014) - Benomyl ^{1,2,7} (Szezeponek and Mazur, 2006). - Copper oxychloride (Ramadevi et al., 2005). - Mancozeb ^{1,2,3,6} , Carbendazim ^{2,7} (Mondal et al., 2018)	- <i>T. viride</i> + <i>P. fluorescens</i> (Senthamarai et al., 2008) - <i>Glomus fasciculatus</i> + <i>P. fluorescens</i> (Singh et al., 2009) - Mangiferin (Ghosal et al., 1977) - <i>Bacillus subtilis</i> (Elewa et al., 2011) - Vascular arbuscular mycorrhiza (Sahab et al., 2001). - <i>Trichoderma viride</i> , <i>Pseudomonas fluorescens</i> , <i>Glomus fasciculatum</i> , <i>G. mosesae</i> (Mondal et al., 2018)

Damping-off	- Mancozeb ^{1,2,3,6} , Copper oxychloride (Mondal et al., 2018) - Copper oxychloride, Mancozeb ^{1,2,3,6} , Carbendazim ^{2,7} (Mondal et al., 2018)	- <i>Trichoderma</i> spp. (Mondal et al., 2018) - Azotobacter ve <i>Trichoderma</i> sp. (Bhat et al., 2014)
Gray Mold	- Mancozeb ^{1,2,3,6} , Zineb ^{1,2} (TNAU, 2013)	- <i>Aloe vera</i> cake, Cassava starch, gelatin and thyme oil, chitosan (Romero et al., 2017)
Blue Mold	- Pre-storage - Sodium hypochlorite, Borax, During-storage – Carbendazim ^{2,7} , Thiophanate methyl ^{1,2,6} (Prakash, 2012)	
Wet Rot	- Mancozeb ^{1,2,3,6} , Copper oxychloride (Mondal et al., 2018)	

¹: PAN Bad Actor, ²: Highly Hazardous Pesticide, ³: Development or Reproductive Toxin, ⁴: Acute Toxicity, ⁵: Acute Toxicity Moderate, ⁶: Carcinogen, ⁷: Carcinogen Possible (Pesticideinfo, 2021)

CONCLUSION

Since some of these chemicals listed in Table 2 are banned and/or restricted plant protection products in the world, including our country, extreme care should be taken in their use in medicinal and aromatic plant growing. Some environmental factors, climate changes, market-oriented crop production and management lead to an increase in the number of pests and diseases in particular. With the frequent use of synthetic pesticides in plant production, their damages to health, food and the environment have increased considerably. The use of chemicals can alter the qualitative and quantitative composition of the active ingredients in plants, which reduces their therapeutic value. For this reason, as a control method, the use of chemicals requires great care and expertise. People have become more interested in traditional and complementary medicine practices. With the reason that our country has a rich flora in terms of plant diversity, the production of medicinal and aromatic plants that can be an alternative to the use of chemical pesticides is supported. These plants, which are widely used in public health services, food and cosmetics sectors globally, continue to increase their agenda every year with the increase in market demand. Biopesticides and bioactive substances have been used instead of synthetic pesticides to prevent deterioration of the quality and increase the yield of the crop. Also, cultural practices and the use of durable varieties are preferred by the producers to reduce the application of synthetic pesticides. The most appropriate and effective control method plan should be combined with the integrated controls. The emergence of fungal diseases, which are frequently appeared in these medicinal and aromatic plants that grow spontaneously or are cultured, causes product and quality losses, creating a commercial and economic threat. For this reason, it is very important to identify, detect and control these diseases.

DECLARATION OF COMPETING INTEREST

The author declares that she has no conflict of interest.

CREDIT AUTHORSHIP CONTRIBUTION STATEMENT

The author contributed 100% to the article.

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