

## Evaluation of Essential Oil Composition Genus *Dittrichia* L. (Asteraceae) Plants in Aydın/Turkey

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### Abstract

The genus *Dittrichia* (Asteraceae), described by Greuter as a small genus, was previously known as a part of *Inula* and has a widespread Mediterranean distribution, marginally penetrating in the Atlantic European territories and in Middle East. The essential oil chemical compositions were derived from the genus *Dittrichia* L. plants were examined in the present study. The study material, *Dittrichia viscosa* (L.) Greuter and *Dittrichia graveolens* (L.) Greuter were collected West Anatolian (Aydın/Turkey) ecological conditions in September-October 2015. Essential oils of the leaves were extracted by Clevenger apparatus. Essential oil compositions were determined with Gas Chromatography-Mass Spectrometry (GC-MS) device. The results from the gas chromatography-mass spectrometry analysis showed that the obtained levo-bornyl acetate from *D. graveolens* was with the highest percentage (25.23%). The 2,4-dioxo-3-methyl-6-isopropyl pyrido[2,3-b]-[1,4]pyrazine in *D. viscosa* was with the highest percentage (29.02%).

**Keywords:** Essential oil, GC-MS, *Dittrichia*, Aydın/Turkey

## Aydın/Türkiye’de Yayılış Gösteren *Dittrichia* L. (Asteraceae) Cinsinin Kimyasal Kompozisyonunun Değerlendirilmesi

### Özet

Küçük bir cins olarak Greuter tarafından tanımlanan *Dittrichia* (Asteraceae) cinsi, önceden *Inula*’nın bir bölümü olarak biliniyordu ve Atlantik Avrupa ülkelerinde, Ortadoğu ve Akdeniz bölgesinde yayılış göstermektedir. Bu çalışma da *Dittrichia* cinsine ait bitkilerden elde edilen uçucu yağların kimyasal bileşimi incelenmiştir. Çalışma materyali *Dittrichia viscosa* (L.) Greuter ve *Dittrichia graveolens* (L.) Greuter Eylül-Ekim 2015 tarihinde Batı Anadolu (Aydın/Türkiye) ekolojik koşullarında toplanmıştır. Yaprakların uçucu yağları Clevenger cihazıyla ekstrakte edildi. Esansiyel yağ bileşimleri Gaz Kromatografisi-Kütle Spektrometresi (GC-MS) cihazı ile belirlendi. Gaz kromatografisi-kütle spektrometresi analizinin sonuçları, *D. graveolens* bitkisinden elde edilen uçucu yağda levo-bornyl acetate en yüksek düzeyde (% 25.23) olduğunu gösterdi. *D. viscosa* bitkisinde ise 2,4-diookso-3-metil-6-izopropil pirido [2,3-b]- [1,4] pirazin en yüksek düzeyde (% 29.02) olduğu belirlendi.

**Anahtar Kelimeler:** Uçucu yağ, GC-MS, *Dittrichia*, Aydın/Turkey

### Introduction

The use of aromatic plants has very long history in the World. According to World Health Organization (WHO), approximately 20. 000 plants have been used for medicinal proposes, 4 000 of which have been widely used (Salehi-Arjmand et al.,

2014). Recently, there has been great interest in essential oils and extracts of medicinal and edible plants, herbs, and spices for the development of alternative food additives, in order to prevent the growth of food-borne pathogens or to delay the onset of food spoilage (Çetin et al., 2011). Therefore

the use of essential oils for treatment of various inflammatory diseases like rheumatism, fever, diabetes suggest that oxidative stress play a role in human disease and intake of antioxidants may improve human health (Burits et al., 2001).

The Asteraceae family, the largest known plant family in the World (Nylinder and Anderberg, 2015), this family consists of approximately 23.000 species in 1535 genera (Öztürk and Çetin, 2013). Asteraceae family contains economically important species. The family contains food plants, raw material resources, medical and medicinal plants, tender and succulent plants, wild weeds and poisonous plants. Acquisition of esculents such as honey and acquisition of cooking oil from this family is used in many fields such as pharmaceutical industry. In addition, many of the species are cultivated as ornamental plants in Asteraceae (Süslü et al., 2010; Paksoy et al., 2016). The genus *Dittrichia*, described by Greuter (Greuter, 1973), as a small genus *Dittrichia* was previously considered as a section of *Inula* (Petropoulou et al., 2004) has a widespread Mediterranean distribution, marginally penetrating in the Atlantic European territories and in Middle East. According to literature data it is well defined and homogeneous in its proposed circumscription, showing close relationships with the genus *Inula* L. (Brullo and Marco, 2000). Genus *Dittrichia* Greuter is represented two species in Turkey. These species *Dittrichia viscosa* [Syn. *Inula viscosa* (L.) Aiton] and *Dittrichia graveolens* [Syn.

*Inula graveolens* (L.) Desf.]. *D. viscosa* is well known for a wide variety of antimicrobial activities. The extracts of the plant possessed antiviral and antioxidant properties. Compounds isolated from the extracts exhibited anti-inflammatory and gastric antiulcerous effects (Al-Qudah et al., 2010). In Turkey the fresh aerial parts are used in folk medicine (Turkish name 'kokarot') for the treatment of wounds (Baytop, 1984). *D. graveolens* has been used in Asian and European traditional medicine as anti-infective, anti-inflammatory, anti-pathogenic, and sedative medication to treat UTI, hemorrhoid, wounds, and leishmaniosis (Maxia et al., 2008).

The aim of the present study is to determine the essential oil contents of the genus *Dittrichia* plants grown under West Anatolian ecological conditions. Furthermore, this study provides a scientific basis for future studies and will help to establish the scientific foundations of traditional treatment methods commonly used by humans.

## Materials and Methods

### Plant Materials

*D.viscosa* and *D.graveolens* aerial parts of the plants were collected as study materials in August and October 2015, which are their blooming periods, from Aydın/Turkey (Figure. 1). The collected plants were held in a room without sunlight in fabric bag.



Figure 1: Location of the Aydın region

### Isolation of Essential Oils

Approximately, 150-200 g of plant samples were used for the essential oil extraction process. Extraction was performed with Clevenger apparatus

(Basaran cam, Turkey and Misung Scientific Co., Korea) using water distillation.

**Table 1.** Essential oil composition of *D.viscosa* and *D.graveolens* plants.

| <i>D.viscosa</i> |   |              | <i>D.graveolens</i> |   |              |
|------------------|---|--------------|---------------------|---|--------------|
| RT               | Component   | Percent (%)  | RT                  | Component   | Percent (%)  |
| 33.39            | $\alpha$ -copaene   | 0.33         | 11.41               | camphene  | 1.35         |
| 35.21            | theaspirane   | 0.35         | 34.33               | levo-bornyl acetate   | 25.23        |
| 41.41            | naphthalene   | 0.55         | 34.55               | exo-methyl-camphenilol  | 0.33         |
| 44.29            | endo-3-butyl-exo-4-methyl<br>bicycle [4.3.0]non-1(9)-en-2-<br>one       | 0.45         | 35.05               | caryophyllene   | 1.31         |
| 45.42            | homoadamantane  | 0.33         | 38.16               | borneol   | 17.83        |
| 49.44            | caryophyllene oxide   | 3.35         | 40.06               | $\delta$ -cadinene  | 0.43         |
| 50.12            | farnesol  | 4.78         | 40.24               | $\alpha$ -amorphene   | 1.27         |
| 50.76            | $\alpha$ -copaen-11-ol  | 0.56         | 40.38               | 1H, 4H-3a,6a-methanopentalen-<br>1-one, tetrahydro-, (3aS)- (CAS) | 0.32         |
| 52.39            | 2-pentadecanone   | 0.55         | 43.25               | butanoic acid   | 0.58         |
| 53.38            | $\alpha$ -longipinene   | 3.08         | 46.96               | caryophyllene oxide   | 2.64         |
| 53.47            | 9-aristolen- $\alpha$ -ol   | 0.33         | 47.53               | nerolidol   | 0.31         |
| 53.69            | 4,5-dihydro-5-oxo-3-(p-tolyl)<br>isoxazole                              | 0.40         | 48.45               | 1-phenyl-1-deutero-2-propen-1-<br>ol                              | 0.78         |
| 53.91            | selina-6-en-4-ol  | 2.58         | 51.63               | naphthalene   | 13.84        |
| 54.28            | calarene  | 0.51         | 52.20               | carvacrol   | 0.64         |
| 54.43            | naphthalene   | 0.30         | 53.09               | $\alpha$ -cadinol   | 0.63         |
| 54.93            | carvacrol   | 0.38         | 53.80               | 7-epi-amiteol   | 1.36         |
| 55.43            | $\alpha$ -eudesmol  | 0.64         | 54.78               | adamantane  | 0.83         |
| 55.73            | $\beta$ -eudesmol   | 2.20         | 54.97               | caryophylla-4 (12), 8(13) diene 5<br>$\beta$ -ol                  | 3.53         |
| 55.90            | (+)-15-hexadecanolide   | 0.44         | 55.89               | caryophylla-3 8(13)-dien-5.beta.-<br>ol                           | 1.06         |
| 56.01            | $\beta$ -costal   | 0.54         | 56.98               | caryophyllenol-II   | 0.91         |
| 56.21            | decanoic acid   | 0.31         | 57.73               | verbenol  | 0.35         |
| 56.34            | 7-epi-amiteol   | 2.52         | 58.17               | 1,1'-bicyclopentyl-1,1'-diol                                      | 0.31         |
| 56.53            | 1. $\beta$ ,4,4-trimethyl-bicyclo (3.2.0<br>) hept-6-en-2. $\alpha$ -ol | 0.38         | 60.39               | pentacosane   | 0.60         |
| 57.50            | caryophylla-4(12),8(13) diene 5<br>$\beta$ -ol                          | 1.85         | 61.04               | cyclobutane carboxylic acid                                       | 1.65         |
| 58.46            | caryophylla-3 8(13)-dien-<br>5.beta.-ol                                 | 0.44         | 68.61               | benzoic acid  | 0.59         |
| 59.55            | vulgarol B  | 0.45         | 70.54               | 1-methyl-2-ethyl-3-oxo dihydro<br>isoindole                       | 0.42         |
| 61.56            | (+)-3-methyl-1-penten-3-ol  | 0.36         | 76.88               | palmitic acid   | 1.09         |
| 62.16            | pentacosane   | 0.33         | 88.99               | 3-(4-methylphenyl)-5,5-<br>dimethyl-4-phenyl-5h-pyrazole          | 1.86         |
| 65.69            | $\gamma$ -costol  | 0.39         | 95.88               | benzoxazole   | 1.85         |
| 65.95            | $\beta$ -costol   | 0.92         | 96.98               | isodrimenin   | 4.47         |
| 67.93            | benzoic acid  | 0.35         | 102.81              | linoleic acid   | 0.48         |
| 79.35            | palmitic acid   | 0.40         |                     |   |              |
| 91.88            | 7-tert-butyl-4-methyl-5-nitro<br>benzisoxazole                          | 4.08         |                     |   |              |
| 99.00            | 2,4-dioxo-3-methyl-6-isopropyl<br>pyrido[2,3-b]-[1,4]pyrazine           | 29.02        |                     |   |              |
| 101.86           | eudesma-5,11(13)-dien-8,12-<br>olide                                    | 24.49        |                     |   |              |
| <b>Total</b>     |   | <b>88.94</b> | <b>Total</b>        |   | <b>88.85</b> |

### GC-MS Analysis

Essential oil analysis were conducted at Eskisehir Anadolu University Medicinal Plants, Drugs and Scientific Research Center (AUBİBAM) by HP 6890 Series Gas Chromatograph 5973 Mass Selective Dedector System instrument equipped with an Agilent HP-Innowax colon (60m X 0.25 mm film, 0.25 µm thickness). Helium (He) was used as a carrier gas. Conditions were as follows; from 50 °C to 240 °C by an increase of 4 °C / minutes. At 240 °C, 40 minutes of waiting time were implemented. Injection port and detector temperature were 250°C and 280°C respectively. Characterization of essential oil components was based on the library (Wiley and NIST) comparison with the mass spectra of the injected essential oil samples.

### Results and Discussion

Essential oils obtained from plants are important natural products used as raw materials in many fields such as perfumes, cosmetics, aromatherapy, and food industry (Yüce et al., 2016). In our study, totaly 121 components were detected as *D.viscosa* aerial parts essential oil composition. 88.94% of the total essential oils in 35 components (components which are ≥0.3% in total ratio) were given in Table 1. The essential oils obtained from the *D.viscosa* plant were detected to contain 2,4-dioxo-3-methyl-6-isopropyl pyrido[2,3-b]-[1,4] pyrazine (29.02%) at most (Table 1). Previous studies detected the highest amounts obtained from essential oils as follows; borneol (25.2%) (Perez-Alonso et al., 1996), fokienol (20.87%) (Al-Qudah et al., 2010), isocostic acid (56.83%) (Madani et al., 2014), 12-carboxyeudesma-3,11 (13) diene (28.88%) (Haoui et al., 2015) and azulene (9.2%) (Alalan et al., 2015). For *D. graveolens*, 110 component were detected aerial parts essential oil composition. 88.85% of the total essential oils in 31 components (components which are ≥0.3% in total ratio) were given in Table 1. The essential oils obtained from the *D.graveolens* plant were detected to contain levo-bornyl acetate (25.23%) at most (Table 1). Past studies detected the highest amounts obtained from essential oils as follows; epi-α-cadinol (30.2%) (Petropoulou et al., 2004), bornyl acetate (43.1–73.1%) (Blanc et al., 2004), 1,8-cineole (%22.4) (Kılıç, 2014), isobornylacetate (50.8 %) (Boudouda et al., 2013), 1,8 cineole (54.89%) (Angel et al., 2011), borneol (43.6%) (Mitic et al., 2016). When checked the old studies results, their results was different then our results both as substances and proportional. This is could be explained by the fact that essential oil composition may have different quality and quantity under different geographical and environmental

conditions, and also during the different periods of the plant growth (Mazandarani et al., 2013).

### Conclusion

According to the results obtained in this study, these results can be a valuable resource for future biotechnological, biological diversity, pharmaceutical and medical studies. At the same time, biodiversity and conservation biology will help you understand the importance of efforts.

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