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## EVALUATION OF ASSOCIATIVE EFFECTS OF *GLIRICIDIA SEPIUM* AND *MEGATHYRSUS MAXIMUS* COMBINATIONS ON FIBRE FRACTIONS, *IN-VITRO* CARBON DIOXIDE AND METHANE PRODUCTION

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Abstract: The voluntary feed intake and digestibility of forages are contingent upon the quality of the pasture, which in turn has an impact on ruminant productivity. There is limited research on the proximate/chemical composition, in-vitro gas production, and degradability of mixtures containing Megathyrsus maximus (MM) and Gliricidia sepium (GS). The study investigated the impact of different combinations of MM and GS on in-vitro gas production, degradability, and chemical composition. Megathyrsus maximus (MM) and Gliricidia sepium (GS) were combined as follows: T1- 100%MM + 0%GS, T<sub>2</sub>- 75%MM + 25%GS, T<sub>3</sub>- 50%MM + 50%GS, T<sub>4</sub>- 25%MM + 75%GS, and T<sub>5</sub>- 0%MM + 100%GS. The treatments were assayed using standard procedures. The proximate composition of the treatments was also determined using standard procedures. Data were analyzed using descriptive statistics and ANOVA at  $\alpha_{0.05}$ . Initial gas produced in T<sub>1</sub> and  $T_2$  and net gas volume (NGV) observed in  $T_1$  and  $T_2$  were significantly lower compared to other treatments. In addition, the CO<sub>2</sub> gas produced in  $T_3$ ,  $T_4$ , and  $T_5$  was significantly higher than that produced in  $T_1$  and  $T_2$ . However,  $T_1$  had the lowest CO<sub>2</sub> gas production. The treatment with 100% legume (T<sub>5</sub>) had the highest methane (CH<sub>4</sub>) production, followed by T<sub>4</sub> and T<sub>3</sub>. Treatments containing 100% (T<sub>1</sub>) and 75% (T<sub>2</sub>) grass had the lowest CH<sub>4</sub> production. In terms of degradability, it was observed that T<sub>5</sub> had significantly higher organic matter degradability (OMD) compared to other dietary treatments. The crude protein observed in T<sub>5</sub> was also significantly higher than other treatments. In conclusion, mixtures with a high content of soluble carbohydrates presented the lowest gas production. It was determined that a mixture of 75% Megathyrsus maximus and 25% Gliricidia sepium has increased carbohydrate, ash, lower moisture content, and in vitro gas production and can be utilized by ruminant farmers as a cheap and readily available source of nutrition for their animals.

Keywords: In-vitro gas production, Chemical composition, Organic Matter Degradability, Ruminant.							
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## 1. Introduction

Ruminant production in developing countries, including Nigeria, faces significant constraints due to the scarcity and fluctuating quality of year-round feed, particularly forages. This challenge is exacerbated by the tropical climate, where forage growth and quality closely mirror the seasonal distribution of rainfall, leading to acute feed shortages during the dry season (Preston, 1995; Babayemi and Bamikole, 2006). Compounding the issue is the high population density in Nigeria, which intensifies land-use competition between pasture for livestock and food crop cultivation. Addressing these challenges to ensure adequate, high-quality feed for ruminants is critical for sustaining livestock productivity and remains a focal concern for agricultural scientists and policymakers (Anim-Jnr et al., 2023).

Forage is herbaceous plants or herbaceous plant parts made available for animal consumption. Forages form the basal diet of ruminants, encompassing a wide array of plant materials consumed by grazing livestock, including grasses, legumes, and crop residues. It can also be defined as 'edible parts of plants, apart from separated grain, that can provide feed for grazing animals or that can be harvested for feeding' (Forage and Grazing Terminology Committee, 1991; Barnes and Baylor, 1995). Grasses, primarily from the *Poaceae* family, are essential forage components, with species such as *Megathyrsus maximus* (syn. *Panicum maximum*, commonly known as guinea grass) demonstrating high adaptability and production potential under local conditions (Ajavi and Babayemi, 2008). Guinea grass is versatile, suitable for cutting, silage, or grazing, and offers higher protein yield and dry matter content compared to other tropical grasses like elephant grass (Pennisetum purpureum) (Man and Wiktorsson, 2003). Ajayi and Babayemi (2008) stated that P. maximum is one of the most common grasses in the savanna region of Nigeria. Under adequate conditions, its nutritional value is high, having up to 12.5% crude protein, Total Digestible Nutrients (TDN) of 10.20 %, and some minerals such as calcium, phosphorus, and magnesium. Other researchers (Aderinola et al., 2014) affirmed that P. maximum has been classified among the best forage grasses due to its high nutritive value. Also, P. maximum produces a high vield of palatable fodder and is suited for grazing, but rapidly declines in nutritive value with age and could also die off if continually grazed close to the ground.

Odeyinka (2001) mentioned that ruminants cannot meet their maintenance needs on grass alone. Bamikole and Babayemi (2004) stated that although ruminants relish P. maximum, this grass becomes scarce during the dry season, and this causes nomads to travel long distances with their livestock in search of greener pastures and in the process, they cause damage to farmlands, lose their animals to snakebites and exposure to extremes of weather, and destroy lives and properties of farmers. These findings led to the search for leguminous forages, which are more palatable and well-accepted by ruminants all year round (Odeyinka, 2001). One such shrub in Nigeria is *Gliricidia sepium*, which is a medium-sized, semi-deciduous tree that grows up to between 10 and 15 m high. Gliricidia sepium, which contains an average of 22.3% crude protein, is described as a suitable feed for ruminants (Bawala et al., 2006). Meanwhile, the integration of mixed agricultural systems such as agropastoralism and silvo-pastoralism in providing suitable tropical forages for ruminants has been an effective strategy in combating feed shortages whilst reducing environmental footprint (Arango et al., 2020).

Furthermore, the combined use of grasses and legumes, for instance, guinea grass and *Glyricidia sepium*, in feeding ruminants is beneficial because the legume fixes nitrogen to the soil, which can be utilized by grasses and thereby increase their crude protein content (Cook et al., 2020). It also provides fermentable nitrogen, other nutrients for rumen microbes, readily fermentable cellulose, and bypass protein. In addition, *Glyricidia sepium* is commonly used as fencing material (Mbah et al., 2024), and it grows all year round; this makes it a legume of great choice for ruminant farmers.

There are many ways in which feed/diets can be evaluated. It can be carried out by proximate or chemical analysis (AOAC, 1990), *in vitro* method (Menke et al., 1979 and Babayemi, 2007), and *in vivo* method, in which the diet is introduced to animals whilst considering feed intake. Although there are other methods used to carry

out *in vitro* fermentation assessment, the gas method is mostly used.

Altering the mix of dietary components is frequently suggested as a tactic that farmers could utilize to diminish the amount of energy expended by animals through eructated gasses (methane (CH<sub>4</sub>), carbon dioxide (CO<sub>2</sub>), nitrous oxide (N<sub>2</sub>O)) and to enhance feed and energy efficiency. Methane is one of the ruminal fermentation end-products synthesized by methanogenic archaebacteria from CO2 and H2 derived from the fermentation of carbon-containing substrates. It is now widely understood that methane is a potential greenhouse gas capable of causing even more global warming than carbon dioxide over time (IPCC, 2013). In ruminants, the production of enteric gas is mostly determined by nutrition and feed intake. Usually, several factors such as forage processing, type of carbohydrate, additives, fat source, degradability, and amount of H<sub>2</sub> produced can influence gas production (Janssen, 2010). In vitro approaches can yield substantial insights into the mechanisms of digestive interactions among different sources. Furthermore, evaluating feed forages individually and in association should yield insights into a plant's capacity to influence nutrient utilization from another plant (Zhang et al., 2017).

Despite these developments, limited research exists on the associative effects of *M. maximus* and *G. sepium* mixtures on proximate composition, in vitro gas production, and nutrient degradability. This study aims to fill this knowledge gap by investigating the impact of varying grass-to-legume ratios on fermentation dynamics, nutrient digestibility, and greenhouse gas emissions. The findings will provide critical insights into optimizing forage mixtures for sustainable ruminant production systems, especially in low-income countries.

## 2. Materials and Methods

The experiment was carried out in 2018 at the Teaching and Research farm of the University of Ibadan, Oyo State, Nigeria. It is situated in the derived savanna vegetation belt (Latitude 7° 27' N and 3° 45'E) and at an altitude between 200m and 300m above sea level; mean temperature of 25-29°C with an average annual rainfall of about 1250mm. The soil is well-drained and belongs to the alfisol (Babayemi et al., 2003). The analyses were, however, carried out in the Laboratories of the Department of Animal Science, University of Ibadan.

## 2.1. Experimental Design and Substrates

The samples used for the experiment were derived from plant materials at the Teaching and Research farm and surroundings of the University of Ibadan, Oyo State, Nigeria. Fresh blades of guinea grass and tender stems and leaves of gliricidia were cut from matured plants. Gliricidia contains approximately 25% DM, and guinea grass contains 23% DM levels. The grass regrowth was cut at 15 cm height above the ground, while the legume was cut at 30 cm from the branch tip. Each fresh sample consisting of leaves and tender stem harvested was sun dried for a week and ground. MM and GS were combined as follows:  $T_1$ - 100%MM + 0%GS,  $T_2$ - 75%MM + 25%GS,  $T_3$ - 50%MM + 50%GS, T<sub>4</sub>- 25%MM + 75%GS, and T<sub>5</sub>- 0%MM + 100%GS.

#### 2.2. Chemical Composition

The ground samples were oven-dried to constant weight at 105 °C. Further proximate analysis (ash, ether extract, crude protein, and crude fiber determination) of all the samples was carried out according to the procedures laid down by the Association of Official Analytical Chemists (AOAC, 2005). Neutral detergent fiber (NDF), acid detergent fiber (ADF), and acid detergent lignin (ADL) were assayed by the method of Van Soest et al. (1991).

## 2.3. In Vitro Fermentation Procedure and Measurement of Gas Production

The rumen fluid was collected prior to early morning feeding. In vitro gas production analyses were performed based on the procedure described by Menke and Steingass (1988). The rumen fluid was collected through the suction method from three West African Dwarf goats under the same feeding regime and condition with the use of a suction tube as described by Babayemi and Bamikole (2006). The animals were fed with 40% concentrate feed (40% corn, 10% wheat offal, 10% palm kernel cake, 20% groundnut cake, 5% soybean meal, 10% dried brewers grain, 1% common salt, 3.75% oyster shell, and 0.25% fish meal) and 60% browse plants. The fluid was then filtered through a four-layered cheesecloth into a warm flask, flushed with carbon dioxide (CO<sub>2</sub>) gas, and kept in a water bath previously heated to 39 °C. The mixture was intermittently stirred using an automatic Two hundred milligrams (200mg) DM of each dried and ground sample was carefully weighed into 100 ml calibrated syringes with pistons lubricated with Vaseline and thereafter, the syringes were filled with 30ml of medium consisting of 10ml of rumen fluid and 20ml of buffered mineral solution (NaHCO3+3Na2HPO4+KCl+NaCl+MgSO4.7H2O+CaCl2.2H 20) and each sample was replicated three (3) times. The syringes were tightly capped and carefully arranged in an incubator maintained at  $39\pm1^{\circ}$  C along with three (3) blank syringes containing 30ml of medium (inoculums and buffer) only as control. The gas production was recorded at 3, 6, 9, 12, 15, 18, 21, and 24 hours. The gas produced was read by measuring the space formed between the top of the piston and the liquid in the syringe. The net gas produced was recorded as the gas produced (in ml) at 24 hours of incubation. After every reading (every 3 hours), the content of the syringes was shaken properly to allow for proper mixing of the substrate and the liquid. After incubation time, 4ml of 10M NaOH solution was introduced to estimate methane (CH<sub>4</sub>) production according to Fievez et al. (2005). Mixing of the contents with NaOH allowed absorption of CO2, with the gas volume remaining in the syringe considered to be CH<sub>4</sub> (Demeyer et al., 1988). Graphs of the volume of gas produced every 3-hour interval of the three replicates of each sample were plotted against the incubation time.

estimated as defined in the equation 1:

Y= a+ b (1-ect) (Orskov and Mc Donald, 1979) where				
Y= degradability at time (t)				
a= intercept (or initial gas produced)	(1)			
b= potentially degradable of b	(1)			
c= rate of degradation of b				
t= incubation time				

The asymptote represents (a+b) of the potential degradability. The intercept of the curve is represented by a and given the DMD value at a time (zero hours). The b value was calculated as the difference between the asymptotic DMD and the intercept a, i.e. (a+b) - a. To get a good estimate of c, Y was selected (DMD% at the time) when the curve changed rapidly. The gas produced on incubation, together with the levels of other chemical constituents, was used to predict the digestibility of organic matter (equations 2 and 3).

Partition Factor (PF) = 
$$OMD/Gp$$
 (3)

Where,

DM dry matter,

OMD organic matter digestibility.

CP, crude protein in percent.

XA, ash in percent.

Gp, the net gas production in ml from 200 mg dry sample after 24 h of incubation and after correction for the day-to -day variation in the activation of rumen liquor using the Hohenheim standard.

#### 2.4. Statistical Analysis

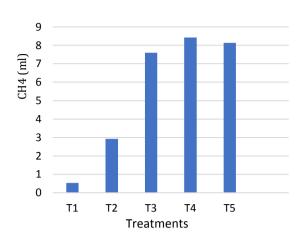
Data obtained was subjected to a one-way Analysis of Variance using SAS software (1990), and the significant differences among the means were separated using the Duncan Multiple Range Test.

## 3. Results

## 3.1. In vitro Gas Production

The in vitro gas production characteristics of forage mixtures are presented in Table 1. A graph showing methane gas production as affected by different proportions of grass-legume mixtures is also presented in Figure 1.

From the graph, the degradation characteristics were



**Figure 1.** Methane gas production as affected by different proportions of forage mixtures. T<sub>1</sub>= 100% Megathyrsus maximus +0% Gliricidia; T<sub>2</sub>= 75% Megathyrsus maximus +25% Gliricidia; T<sub>3</sub>= 50% Megathyrsus maximus +50% Gliricidia; T<sub>4</sub>= 25% Megathyrsus maximus +75% Gliricidia; T<sub>5</sub>= 0% Megathyrsus maximus +100% Gliricidia It was observed that initial gas (a) produced in T<sub>1</sub> (3.00) and T<sub>2</sub> (2.50) were similar and significantly (P<0.05)

lower compared to other treatments. The potentially degradable fraction (b) observed in  $T_3$  (7.67) and  $T_5$ (10.33) were significantly (P<0.05) higher compared to other treatments. However, the lowest (P<0.05) potentially degradable fraction was observed in  $T_1$  (1.67). Higher (P<0.05) potential gas production (a+b) was observed in T<sub>3</sub> (17.00), T<sub>4</sub> (16.33), and T<sub>5</sub> (17.67) compared to other treatments. The lowest (P<0.05) potential gas production was observed in  $T_1$  (1.33). The  $T_4$ (0.13) had a significantly (P<0.05) higher rate of degradation compared to other treatments. However, T<sub>1</sub> (0.00) and T<sub>3</sub> (0.01) were significantly (P<0.05) lower compared to other treatments. Higher (P<0.05) incubation time (t) was observed in T<sub>2</sub> (16.50) compared to  $T_1$  (7.50) and  $T_3$  (10.00) but did not differ significantly (P<0.05) from T<sub>4</sub> (11.00) and T<sub>5</sub> (15.00). The degradability at t (Y) was significantly (P<0.05) affected by different proportions of forage mixtures. It was observed that degradability in  $T_4$  (12.66) and  $T_5$  (15.22) was significantly (P<0.05) higher compared to  $T_1$  (4.67) and  $T_2$ (4.17).

Table 1. In vitro Gas production characteristics of the forage mixtures

Parameters	T1	T2	Т3	T4	T5	SEM	P value
inital gas produced	3.00 <sup>b</sup>	2.50 <sup>b</sup>	9.33ª	11.00 <sup>a</sup>	11.00 <sup>a</sup>	2.80	0.02
potentially degradable fraction	1.67c	3.33 <sup>bc</sup>	7.67ª	5.35 <sup>b</sup>	10.33ª	1.50	0.04
potential gas production	1.33c	5.83 <sup>b</sup>	17.00 <sup>a</sup>	16.33ª	17.67ª	0.83	0.01
rate of degradation	0.00 <sup>c</sup>	0.04bc	0.01c	0.13ª	$0.07^{b}$	0.02	0.02
incubation time	7.50 <sup>b</sup>	16.50ª	10.00 <sup>b</sup>	11.00 <sup>ab</sup>	15.00 <sup>ab</sup>	3.03	0.04
degradability at time (t)	4.67 <sup>b</sup>	4.17 <sup>b</sup>	10.22 <sup>ab</sup>	12.66ª	15.22ª	2.71	0.04

<sup>abc</sup> Means for treatments along x axis with different superscripts differed significantly (P<0.05)

 $T_1=100\% \text{ Megathyrsus maximus } +0\% \text{ Gliricidia; } T_2=75\% \text{ Megathyrsus maximus } +25\% \text{ Gliricidia; } T_3=50\% \text{ Megathyrsus maximus } +50\% \text{ Gliricidia; } T_4=25\% \text{ Megathyrsus maximus } +75\% \text{ Gliricidia; } T_5=0\% \text{ Megathyrsus maximus } +100\% \text{ Gliricidia.}$ 

#### 3.2. In- vitro Fermentation Parameters

Table 2 shows the in vitro fermentation parameters of grass-legume mixtures. The net gas volume (NGV) observed in T<sub>1</sub> (1.33) and T<sub>2</sub> (5.83) was significantly (P<0.05) lower compared to other treatments. The CH<sub>4</sub> gas produced in T<sub>1</sub> (0.53) was the lowest when compared to other treatments and was not significantly different from T<sub>2</sub> (2.93). However, T4 (8.43) had the highest (p <

0.05) CH<sub>4</sub> gas production. Different proportions of forage mixtures did not significantly (P<0.05) affect net methane (NM) to net gas (NG) ratio and ranged from 0.40 (T<sub>1</sub>) to 0.53 (T<sub>4</sub>). The CO<sub>2</sub> to organic matter digestibility (OMD) ratios observed in T<sub>3</sub> (0.25), T<sub>4</sub> (0.28), and T<sub>5</sub> (0.26) were similar and significantly (P<0.05) higher compared to other treatments. The T<sub>1</sub> (0.02) had the lowest (P<0.05) CO<sub>2</sub>/OMD.

Table 2. In vitro fermentation parameters of the grass and legume mixtures

Parameters	T1	T2	Т3	T4	T5	SEM	P value
NGV	1.33 <sup>b</sup>	5.83 <sup>b</sup>	17.00 <sup>a</sup>	16.33ª	17.67ª	2.54	0.02
CH <sub>4</sub>	0.53 <sup>b</sup>	2.93 <sup>b</sup>	7.60 <sup>a</sup>	8.43ª	8.13ª	1.29	0.04
NM/NG	0.40	0.51	0.45	0.53	0.47	0.04	0.36
OMD	30.61 <sup>b</sup>	29.99 <sup>d</sup>	29.91 <sup>d</sup>	30.49c	30.79ª	0.03	< 0.0001
CO <sub>2</sub>	0.80c	2.90c	9.40 <sup>a</sup>	7.90ª	9.53ª	0.80	0.02
CO <sub>2</sub> /OMD	0.02c	0.10 <sup>b</sup>	0.25ª	0.28ª	0.26ª	0.03	0.02
CH4/OMD	0.03 <sup>b</sup>	0.09 <sup>b</sup>	0.31ª	0.26ª	0.31ª	0.03	0.03
PF	$0.07^{a}$	0.02 <sup>b</sup>	0.01c	0.01 <sup>c</sup>	0.01 <sup>c</sup>	0.003	0.004

<sup>abcd</sup> Means of treatments along a row with different superscript differed significantly (P<0.05), NGV= net gas volume, CH<sub>4</sub>= methane, NM/NG- Net methane: Net gas. T<sub>1</sub>= 100% *Megathyrsus maximus* +0% *Gliricidia*; T<sub>2</sub>= 75% *Megathyrsus maximus* +25% *Gliricidia*; T<sub>3</sub>= 50% *Megathyrsus maximus* +50% *Gliricidia*; T<sub>4</sub>= 25% *Megathyrsus maximus* +75% *Gliricidia*; T<sub>5</sub>= 0% *Megathyrsus maximus* +100% *Gliricidia*.

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The T<sub>1</sub> (0.03) and T<sub>2</sub> (0.09) had significantly (P<0.05) lower CH<sub>4</sub> to OMD ratios compared to treatments. The CO<sub>2</sub> gas produced in T<sub>3</sub> (9.40), T<sub>4</sub> (7.90), and T<sub>5</sub> (9.53) were significantly (P<0.05) higher compared to other treatments. However, T<sub>1</sub> (0.80) had the lowest (P<0.05) CO<sub>2</sub> gas production. The net CO<sub>2</sub> to net gas ratio was not significantly (P<0.05) affected by different proportions of forage mixtures and ranged from 0.47 (T<sub>4</sub>) to 0.60 (T<sub>1</sub>).

## 3.3. Nutritional Composition of Grass-legume Mixtures

Table 3 shows the proximate composition of dietary treatments as influenced by different proportions of forage mixtures. Higher (p<0.05) Moisture Content (MC) was observed in T<sub>5</sub> (5.95%) compared to other dietary treatments. The T<sub>2</sub> (4.37%) had the lowest (p<0.05) MC. The Crude Protein (CP) observed in T<sub>5</sub> (21.48) was significantly (p<0.05) higher compared to other treatments. However, the lowest (p<0.05) CP was

observed in  $T_1$  (13.16). Higher (p<0.05) ether extract (EE) was observed in  $T_1$  (3.29) compared to other treatments. The  $T_5$  (1.98) had the lowest (p<0.05) EE but was similar to T<sub>3</sub> (2.01). The crude fiber (CF) observed in T<sub>1</sub> (25.99) was significantly (p<0.05) higher compared to other dietary treatments. Similar (p<0.05) CF values were observed in T<sub>3</sub> (22.60) and T<sub>4</sub> (22.59). However, T<sub>3</sub> (20.20) had the lowest (p < 0.05) CF. Ash content observed in  $T_1$ (9.13) was significantly (p<0.05) higher compared to other treatments. The lowest (p<0.05) ash content was observed in  $T_3$  (5.55). The NFE observed in  $T_2$  (71.46) was significantly (p<0.05) higher compared to other dietary treatments. The  $T_1$  (69.71) and  $T_3$  (69.72) had similar (p<0.05) NFE values. However,  $T_5$  (65.04) had the lowest (p<0.05) NFE value. Higher (P<0.05) partitioning factor (PF) was observed in T1 (0.07) compared to other dietary treatments.

Table 3. Proximate composition of grass and legume mixtures

Parameters	T1	T2	Т3	T4	T5	SEM	P value
Moisture content	4.70 <sup>d</sup>	4.37c	5.42 <sup>b</sup>	4.84c	5.95ª	0.02	< 0.0001
СР	13.16 <sup>c</sup>	13.99 <sup>d</sup>	15.38c	18.39 <sup>b</sup>	21.48ª	0.03	< 0.0001
EE	3.29 <sup>a</sup>	2.01c	2.01 <sup>d</sup>	2.09 <sup>b</sup>	1.98 <sup>d</sup>	0.01	< 0.0001
CF	25.99ª	24.01 <sup>b</sup>	22.60 <sup>c</sup>	22.59°	20.20 <sup>d</sup>	0.01	< 0.0001
Ash	9.13ª	8.16 <sup>b</sup>	7.47°	6.59 <sup>d</sup>	5.55c	0.03	< 0.0001
NFE	69.71 <sup>b</sup>	71.46 <sup>a</sup>	69.72 <sup>b</sup>	68.08c	65.04 <sup>d</sup>	0.05	< 0.0001

<sup>abcd</sup> Means of treatment along a row with different superscript differed significantly CP= crude protein, EE- (P<0.05) ether extract, CF= crude fiber, NFE-Nitrogen free extract.  $T_1$ = 100% Megathyrsus maximus +0% Gliricidia;  $T_2$ = 75% Megathyrsus maximus +25% Gliricidia;  $T_3$ = 50% Megathyrsus maximus +50% Gliricidia;  $T_4$ = 25% Megathyrsus maximus +75% Gliricidia;  $T_5$ = 0% Megathyrsus maximus +100% Gliricidia.

## 3.4. Fiber Fractions of Dietary Treatments

The fiber fractions of dietary treatments as affected by different proportions of forage mixtures are shown in Table 4. İt was observed that the Neutral Detergent Fiber (NDF) was not significantly (P<0.05) affected by different proportions of forage mixtures and ranged from 58.90 (T<sub>2</sub>) to 62.76 (T<sub>4</sub>). Higher (P<0.05) acid detergent fiber (ADF) values were observed in T<sub>4</sub> (31.90) and T<sub>5</sub> (30.60) compared to other dietary treatments. The T<sub>2</sub> (27.41) had

the lowest (P<0.05) ADF value compared to other dietary treatments. It was observed that the T<sub>3</sub> (16.60) had significantly (P<0.05) higher acid detergent lignin (ADL) value compared to other dietary treatments. Similarly (P<0.05) ADL values were observed in T<sub>4</sub> (15.80) and T<sub>5</sub> (15.53) and were higher (P<0.05) than in T<sub>1</sub> (14.60) and T<sub>2</sub> (14.20). However, the lowest (P<0.05) ADL value was observed in T<sub>2</sub> (14.20).

Table 4. Fiber fractions of dietary treatments as affected by different proportions of forage mixtures

Parameters	T1	T2	Т3	T4	T5	SEM	P value
NDF	60.06	58.90	60.02	62.76	60.39	1.45	0.47
ADF	28.60 <sup>c</sup>	27.41 <sup>d</sup>	31.06ª	30.90 <sup>b</sup>	30.60 <sup>b</sup>	0.08	< 0.0001
ADL	14.60 <sup>c</sup>	14.20 <sup>d</sup>	16.60 <sup>a</sup>	15.80 <sup>b</sup>	15.53 <sup>b</sup>	0.12	< 0.0001

<sup>abcd</sup> Means of treatments along a row with different superscript differed significantly (P<0.05). NDF= neutral detergent fiber, ADF= acid detergent fiber, ADF= acid detergent fiber, ADL= acid detergent lignin. T<sub>1</sub>= 100% Megathyrsus maximus +0% Gliricidia; T<sub>2</sub>= 75% Megathyrsus maximus +25% Gliricidia; T<sub>3</sub>= 50% Megathyrsus maximus +50% Gliricidia; T<sub>4</sub>= 25% Megathyrsus maximus +75% Gliricidia; T<sub>5</sub>= 0% Megathyrsus maximus +100% Gliricidia.

## 4. Discussion

*Megathyrsus maximus* is one of the most common grasses in the savanna region of Nigeria. Under adequate conditions, its nutritional value is high, having up to 12.5% crude protein, Total Digestible Nutrients (TDN) of 10.20%, and some minerals such as calcium, phosphorus, and magnesium (Ajayi and Babayemi, 2008). Aderinola et al. (2014), affirmed that *Megathyrsus maximus* has been classified among the best forage grasses due to its high nutritive value and produces high yield of palatable fodder suitable for grazing, but rapidly declines in nutritive value

with age and could also die off if continually grazed close to the ground. These reports are consistent with the findings of the current study, as observed in  $T_1$  with a crude protein content of 13.16%.

The result of the current study confirms the report of Bawala et al. (2006), who described *Gliricidia sepium* as a suitable feed for ruminants and that the combined use of grasses and legumes, for instance, guinea grass and *Gliricidia sepium* in feeding ruminants is beneficial because the legume fixes nitrogen to the soil which can be utilized by the grass and thereby increase its crude protein. It also provides fermentable nitrogen, other nutrients for the rumen microbes, readily fermentable cellulose, and bypass protein.

Kearny (2005), noted that fiber plays a fundamental role in ruminant nutrition, and it is the component in a feed that is not digested by mammalian enzymes. Some of these components are soluble under mild extraction procedures and thus result in 'soluble' and 'insoluble' fiber. Most constituents of soluble fiber (pectin, fructans, and  $\beta$ glucans) are readily fermented in the rumen and may even be readily fermented in the large intestine of monogastric animals (Righi et al., 2008). From the current study, the NDF was not significantly affected by different proportions of forage mixtures and ranged from 58.90  $(T_2)$  to 62.76  $(T_4)$ . Higher ADF values were observed in  $T_3$ (31.06) and T<sub>4</sub> (30.90) compared to other dietary treatments. The  $T_2$  (27.41) had the lowest ADF value. Merten (1997), stressed that the chemical definition of dietary fiber, such as neutral detergent fiber (NDF) or acid detergent fiber (ADF) content, was an inadequate description of the fiber content of a diet. The ADF fraction of feedstuffs includes cellulose and lignin as the primary components. Concentrations of ADF and lignin are correlated more with digestibility than with intake. It is an indicator of digestibility; as the ADF increases, digestibility decreases. NDF is a measure of cellulose, hemicelluloses ADF, and lignin fractions of feeds. NDF is more highly correlated with feed volume and chewing activity than ADF or crude fiber (CF) (Oba et al., 1999).

Some of the NDFs are highly digestible, but forage NDF is the best indicator of an animal's voluntary feed intake (VFI). As the NDF content of forage increases, the VFI decreases. Also, it has been shown several times that the digestibility of plant material in the rumen is related to the proportion and lignification of plant cell walls (NDF). Forage with high lignin contents is often of low digestibility (Norton, 2010). Merry et al. (2006), reported that when animals are fed a grass and legume mixture, digestive interactions can occur in the rumen between substrates contained in the different plants, and the response of the animal to the combination of forages can differ from the balanced median values of their components considered individually. These interactions, named associative effects, can modify the metabolic processes in the digestive tract, particularly in the rumen, so that the response of an animal to a combination of forages can differ from the balanced median values of its components considered individually. This kind of response can be synergistic or antagonistic with a possible impact in terms of nutrient use by the animal, N excretion, and methane emissions. Dhiman and Satter (1997) noted that the nutritional complementarities of plant species could contribute to integrating productivity and environmental requirements, as observed from the current study. The net gas volume produced was greatest in the mixtures ranging from 50%-100% Gliricidia. This may be due to two factors. One is the higher crude protein content in these combinations, as the greater availability of crude protein in the diet allows for greater microbial activity as it is not limiting in the diet.

Johnson et al. (1995) reported that the production of methane represents a loss of between 2 and 15% of the gross energy in the feed. The authors increased feed efficiency by production and inhibition of rumen methanogenesis because methane plays a role in the global warming phenomenon and the destruction of the ozone layer. Chynoweth (1996) reported that compared to other greenhouse gases, methane is an excellent candidate to reduce global warming in the near term. However, Houghton (1997) reported that because of the shorter lifetime of methane in the atmosphere (about 12 years compared with 100-200 years for CO2), only a relatively small reduction in the anthropogenic emission of CH4, about 8% would be required to stabilize its concentration at the current level. Leng (1993) concluded that enteric methane emission is one of the few global sources of methane that can be reduced relatively. The author further stated that it is easier to manipulate, for instance, methane produced from marshes or in rice production. Furthermore, methane reduction strategies from livestock will directly benefit the farmers by improving animal productivity. The result of the current study on methane production has shown that 100% Megathyrsus maximus or a combination of 75% Megathyrsus maximus and 25% Gliricidia sepium had the least methane production.

Furthermore, Beauchemin et al. (2009) reported that improving forage quality (i.e., increasing dietary starch content) through the supplementation of alternative forages, such as leguminous and non-leguminous shrubs, has the potential to reduce CH<sub>4</sub> emissions per kg animal products as a result of increased diet digestibility and a shortened duration of feeding. The authors further stated that dietary strategies such as this have been successful in manipulating methanogenesis, at least in the short term, through either direct inhibition of methanogens, reducing the production of hydrogen in the rumen, or providing alternative sinks for the disposal of hydrogen. Carulla et al. (2005) observed a similar report and stated that secondary plant compounds (e.g., condensed tannins and saponins) have been shown to reduce enteric CH<sub>4</sub> emissions through the direct inhibition of methanogens. Similarly, including high starch feedstuffs favors propionate production and reduces ruminal pH, thus inhibiting methanogen and protozoal growth (Boadi et al.,

2004). From the current study, the degradability of forage mixtures was affected by different proportions of inclusion. The T<sub>5</sub> (30.79) had significantly higher OMD compared to other dietary treatments. Similar OMD values were observed in T<sub>2</sub> (29.99) and T<sub>3</sub> (29.91) and were significantly lower compared to other dietary treatments. The result of the present findings corroborates the report of Swan et al. (2006), who noted that the degradation of feed ingredients also depends on the distribution of starch granules within the substance. Qin et al. (2012) also added that starch granules of wheat endosperms seem to be floury and have a relatively small particle size. Consequently, the smaller starch granules have a larger surface area available for microbial and enzymatic starch hydrolysis, which results in rapid degradation. Bonhomme (1990) reported that increased methane emission can be observed as a result of the optimum symbiotic relationship between bacteria and protozoans and the efficient exchange of hydrogen between these microorganisms. Furthermore, methane production is affected by the type of carbohydrate fed to the animals (Moe and Tyrrell, 1979). Qin et al. (2012) reported that wheat had relatively higher effective degradability of dry matter (EDDM), which was more rapidly fermented by ruminal microbes. Thus, having a higher methane production of wheat may be attributed to EDDM. Moreover, low methane production of other feed ingredients might also be attributed to low EDDM and, thus, slow fermentation of ruminal microbes. Low EDDM might be due to the thickness of the protein matrix, which coats starch granules, and this matrix is relatively difficult to hydrolyze with water and enzymes (McAllister et al., 1996). The result of the present study contradicts the findings of Shibata (1992), who noted that providing ruminants with feed containing carbohydrates and high protein levels had a negative effect on methane emission while providing a diet rich in fiber resulted in an elevated volume of methane being produced. It was observed that T<sub>5</sub> has the highest methane production, followed by T<sub>4</sub> and  $T_3$ .  $T_1$  and  $T_2$  had the lowest methane production.  $T_1$  had the lowest degradability, whereas T<sub>5</sub> had the highest degradability.

## 5. Conclusion

The study demonstrated that varying proportions of *Megathyrsus maximus* and *Gliricidia sepium* significantly influence nutrient composition, in vitro gas production, and organic matter degradability. Treatments with higher proportions of *Gliricidia sepium* exhibited elevated crude protein levels, methane, and  $CO_2$  production, with T<sub>5</sub> (100% G. sepium) achieving the highest crude protein content (21.48%) and methane production (8.13 mL). Conversely, treatments with higher *M. maximus* content, such as T<sub>1</sub> (100% *M. maximus*), had higher crude fiber (25.99%) but lower methane production (0.53 mL), indicating reduced fermentability.

The degradability of organic matter significantly improved with increasing *G. sepium* inclusion, with  $T_5$ 

recording the highest value (30.79%). However, methane emissions also increased with higher legume proportions, highlighting a trade-off between enhanced digestibility and environmental impact. The study suggests that a 75% *M. maximus* and 25% *G. sepium* mixture offers an optimal balance between nutrient availability, low methane emissions, and adequate degradability, making it a suitable option for sustainable ruminant feeding systems. This proportion minimizes greenhouse gas emissions while ensuring efficient nutrient utilization, supporting productivity in tropical livestock systems.

#### **Author Contributions**

The percentage of the authors' contributions is presented below. All authors reviewed and approved the final version of the manuscript.

	G.A.I.	0.0.	U.Ş.
С	30	70	
D	30	70	
S		100	
DCP	70	30	
DAI	50	50	
L	90	10	
W	80		20
CR	30	40	30
SR	30	40	30
РМ	50	50	
FA	100		

C= concept, D= design, S= supervision, DCP= data collection and/or processing, DAI= data analysis and/or interpretation, L= literature search, W= writing, CR= critical review, SR= submission and revision, PM= project management, FA= funding acquisition.

## **Conflict of Interest**

The authors declared that there is no conflict of interest.

## **Ethical Consideration**

Ethics committee approval was not required for this study because there was no study on animals or humans.

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## References

- Aderinola OA, Lateef OA, Binuomote RT, Adeeyo A, Jekayinfa OA. 2014. Nutritional and microbial contents of varied combinations of ensiled Megathyrsus maximus and Vetiveria nigritana grass. Intr J Food Agri Vet Sci 4(1): 141- 148
- Ajayi FT, Babayemi OJ. 2008. Comparative evaluation of mixtures of Panicum maximum cv Ntchisi with stylo (Stylosanthes guianensis), lablab (Lablab purpureus), centro (Centrosema pubescens) and tridax. Livestock Research for Rural Development, 20(6).
- Anim-Jnr AS, Sasu P, Bosch C, Mabiki FP, Frimpong YO, Emmambux MN, Greathead HMR. 2023. Sustainable small ruminant production in low- and middle-income African Countries: Harnessing the potential of agroecology. Sustainability, 15(21): 15326.

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- AOAC (Association of Official Analytical Chemists) 1990. Official methods of analysis. 15<sup>th</sup> Ed. Washington DC, USA, pp: 2.
- AOAC, 2005. Official methods of analysis. Assoc Offic Analy Chem Washington, DC, USA, pp: 48.
- Arango J, Ruden A, Martinez-Baron D, Loboguerrero AM, Berndt A, Chacón M, et al. 2020. Ambition meets reality: achieving GHG emission reduction targets in the livestock sector of Latin America Front Sustain Food Syst, 4: 65.
- Babayemi OJ, Bamikole MA. 2006. Effect of Tephrosia candida DC leaf and its mixtures with guinea grass on the in vitro fermentation changes in feed for ruminants in Nigeria. Pakistan J Nutr 5(1): 14-18.
- Babayemi OJ, 2007. In vitro fermentation characteristic and acceptability by West African dwarf goats of some dry season forages. African J Bio, 6(10): 1260-1265.
- Bamikole MA, Babayemi OJ. 2004. Feeding goats with Guinea grass, veruno stylo and nitrogen fertilized grass with energy concentrate, Archi Zoot, 53: 13-23.
- Barnes RF, Baylor JE. 1995. Forages in a changing world. In: Barnes RF, Miller DA, Nelson CJ. (eds) Forages, Vol. 1. An introduction to grassland agriculture. Iowa State University Press, Ames, Iowa, USA, pp: 3-13.
- Bawala TO, Isah OA, Akinsoyinu AO. 2006. Studies on milk mineral composition of lactating West African Dwarf goats. J Anim Vet advances 5(10): 805-809.
- Beauchemin KA, McGinn SM, Benchaar C, Holtshausen L. 2009. Crushed sunflower, flax, or canola seeds in lactating dairy cow diets: effects on methane production, rumen fermentation, and milk production. J Dairy Sci, 92(5): 2118-2127.
- Boadi D, Benchaar C, Chiquette J, Masse D. 2004. Mitigation strategies to reduce enteric methane emissions from dairy cows: Update review. Canadian J Anim Scie, 84: 319-335.
- Bonhomme A. 1990. Rumen ciliates: Their metabolism and relationships with bacteria and their hosts. Anim Feed Sci Tech, 30: 203-266.
- Carulla J, Kreuzer M, Machmueller A, Hess H. 2005. Supplementation of acacia mearnsii tannins decrease methanogenesis and urinary nitrogen in forage-fed sheep. Australian J Agri Research, 56. 10.1071/AR05022.
- Chynoweth DP. 1996. Environmental impact of biomethanogenesis. Envi Moni Asses, 42: 3-18.
- Cook BG, Pengelly BC, Schultze-Kraft R, Taylor M, Burkart S, Cardoso Arango JA, et al. 2020. Tropical forages: An interactive selection tool, 2nd and revised Edn. Cali; Nairobi: International center for tropical agriculture (CIAT); Colombia and international livestock research institute (ILRI). Available online at: www.tropicalforages.info (accessed date: January 15, 2025).
- Demeyer D, De Meulemeester M, De Graeve K, Gupta BW. 1988. Effect of fungal treatment on nutritivevalue of straw. Med Fac Landbouww Rijks univ Gent 53: 1811-1819.
- Dhiman TR, Satter LD. 1997. Yield response of dairy cows fed different proportions of alfafa silage and corn silage. J Dairy Sci, 80: 2069-2082.
- Fievez V, Babayemi OJ, Demeyer D. 2005. Estimation of direct and indirect gas production in syringes: a tool to estimate short chain fatty acid production requiring minimal laboratory facilities. Anim Feed Sci Tech, 128-124, 197-210.
- Forage and Grazing Terminology Committee 1991. Terminology for grazing Lands and grazing animals. Pocahontas Press, Blacksburg, Virginia, USA, pp: 45-60.
- Getachew G, Makkar HPS, Becker K. 1999. Stochiometric relationship between short chain fatty acid and in vitro production in presence and absence of polyethylene glycol for tannin containing browses. In: FAAP satellite symposium gas

production. Fermentation Kinetics for feed evaluation and to assess microbial activity. August 18-19 Wageningen, Netherlands.

- Houghton J. 1997. Global Warming. The complete briefing, 2nd ed. xv + 251 pp. Cambridge, New York, USA, Port Chester, Melbourne, Sydney: Cambridge University Press. ISBN 0 521 62089 9; 0 521629322(pb). GeologicalMagazine. 1998; 135(6): 819-842.
- Intergovernmental Panel on Climate Change (IPCC). 2013. The physical science basis. In Working group i contribution to the fifth assessment report of the intergovernmental panel on climate change; Stocker, T.F., Qin, D., Plattner, G.K., Tignor, M.M.B., Allen, S.K., Boschung, J., Nauls, A., Xia, Y., Bex, V., Midgley, P.M., Eds.; Cambridge University Press: Cambridge, UK, 2013.
- Janssen PH. 2010. Influence of hydrogen on rumen methane formation and fermentation balances through microbial growth kinetics and fermentation thermodynamics, Anim Feed Scie Tech, 160(1–2): 1-22.
- Johnson KA, Johnson DE. 1995. Methane emissions from cattle. J Anim Sci, 73: 2483-92.
- Jouany JP. 1994. Les fermentations dans le rumen et leur optimisation. INRA Prod Anim 7(3): 207-225.
- Kearny CC. 2005. Effects of dietary physical form and carbohydrate profile on captive giraffe. Masters thesis. Gainesville, Florida, USA: University of Florida.
- Leng RA. 1993. Quantitative ruminant nutrition-a green science. Australian J Agri Research, 44(3): 363-380.
- Man NV, Wiktorsson H. 2003. Forage yield, nutritive value, feed intake, and digestibility of three grass species as affected by harvest frequency. Trop Grassland, 37: 101-110.
- Mbah CN, Mbah EC, Orji JE, Igberi C, Abam P, Awere SU. 2024. Using *Gliricidia sepium* prunings as green manure in a degraded ultisol; effects on soil physical properties and yield of okra (*Abelmuschus esculentus*) in Abakaliki, southeast Nigeria. Biol Agri Hort, 40(4): 257-266.
- McAllister TA, Cheng KJ, Okine EK, Mathison GW. 1996. Dietary, environmental, and microbiological aspects of methane production in ruminants. Canadian J Anim Sci 76(2): 231-243.
- Menke KH, Raab L, Salewski A, Steingass H, Fritz D, Schneider W. 1979. The estimation of the digestibility and metabolizable energy content of ruminant feedstuffs from the gas production when they are incubated with rumen liquor. J Agri Sci, 93: 217.
- Menke KH. Steingass. 1988. Estimation of the energy of the energetic feed value obtained from chemical analysis and in vitro gas production using rumen fluid. Anim Res Dev, 28: 7-1.
- Merry RJ, Lee MRF, Davies DR, Dewhurst RJ, Moorby JM, Scollan ND, Theodorou MK. 2006. Effects of high- sugar ryegrass silage and mixtures with red clover silage on ruminant digestion. I. In vitro and in vivo studies of nitrogen utilization. J Anim Sci, 84: 3049-3060.
- Merten DR. 1997. Creating a system for meeting the fibre requirement of Dairy cattle. J Dairy Sci, 80: 1463-1482.
- Moe PW, Tyrrell HF. 1979. Methane production in dairy cows. J Dairy Sci, 62(10): 1583-1586.
- Norton BW. 2010. The Nutritive value of tree legumes, http://www.fao.org/ag/AGP/doc/Publication/Guttshel/×55 56e0.html (accessed date: January 15, 2025).
- Oba M, Allen MS. 1999. Evaluation of the importance of the digestibility of neutral detergent fibre from forage: Effects on dry matter intake and milk yield of dairy cows. J Dairy Sci, 82: 529-596.
- Odeyinka SM. 2001. Effect of feeding varying levels of Leucaena leucocephala and Gliricidia sepium on the performance of West African Dwarf goats. Nigerian. J Anim Prod, 28(1): 61-64.

- Orskov ER, McDonald I. 1979. The estimation of protein degradability in rumen from incubation measurements weighted according to the rate of passage. J Agri Sci Cambridge, 92: 449-503.
- Preston TR. 1995. Tropical animal feeding. A manual for research workers. FAO Anim Prod Heal Paper 126, pp: 305.
- Qin WZ, Li CY, Kim JK, Ju JG, Song MK. 2012. Effects of defaunation on fermentation characteristics and methane production by rumen microbes in vitro when incubated with starchy feed sources. Asian-Australasian J Anim Scie, 25(10): 1381-8. http://doi:10.5713/ajas.2012.12240.
- Righi F, Ruini P, Romanelli S, Renzi M, Rossi F, Quarantelli A. 2008. Study of alternative field systems for the evaluation of total mixed ration physical form. Corso di Vet med - Università degli Studi di Parma Vol. 28: pp: 181- 190.
- Shibata M. 1992. Methane production in heifers, sheep and goats consuming diets of various hay-concentrate ratios. J Anim Sci

Tech, 63: 1221-1227.

- Statistical Analysis System Institute Inc 1990 SAS/STAT® user's guide Int Volume 1, version 6, Fourth Edition, Cary, NC, USA.
- Swan CG, Bowman JGP, Martin JM, Giroux MJ. 2006. Increased puroindoline levels slow ruminal digestion of wheat (Triticum aestivum L.) starch by cattle. J Anim Sci Tech. 84: 641-650.
- Theodorou MK, Williams BA, Dhanoa MS, McAllen AB, France J. 1994. A simple gas production method using a pressure transducer to determine the fermentation kinetics of ruminant feeds. Anim Feed Sci Tech, 48: 185-197.
- Van Soest PJ, Robertson JB, Lewis BA. 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. J Dairy Sci, 74(10): 3583-97.
- Zhang J, Shi H, Wang Y, Li S, Cao Z, Ji S, He Y, Zhang H. 2017. Effect of dietary forage to concentrate ratios on dynamic profile changes and interactions of ruminal microbiota and metabolites in holstein heifers. Fronti in Microb. 8: 2206.