

Evaluation of Notch1 gene expression in prostate carcinoma

Prostat kansinomunda Notch1 gen ekspresyonunun değerlendirilmesi

Zeynep Bayramoğlu¹, Betül Ünal², Sema Sezgin Göksu³, Cumhuri İbrahim Başsorgun²

¹ Department of Pathology, Konya Training and Research Hospital, Turkey

² Department of Pathology, Akdeniz University School of Medicine, Antalya, Turkey

³ Department of Medical Oncology, Akdeniz University School of Medicine, Antalya, Turkey

ORCID ID of the author(s)

ZB: 0000-0001-7075-8819

BÜ: 0000-0002-9572-3601

SSG: 0000-0002-1222-0444

CİB: 0000-0003-2440-511X

Corresponding author/Sorumlu yazar:

Zeynep Bayramoğlu

Address/Adres: Konya Eğitim ve Araştırma Hastanesi, Patoloji Kliniği, Konya, Türkiye
e-Mail: drzeynepbayramoglu@hotmail.com

Ethics Committee Approval: This study was approved by Akdeniz University Ethics committee (Date: 11/30/2016, number: 625). All procedures in this study involving human participants were performed in accordance with the 1964 Helsinki Declaration and its later amendments.

Etik Kurul Onayı: Bu çalışma Akdeniz Üniversitesi Etik Kurulu tarafından onaylanmıştır (Tarih: 30.11.2016, sayı: 625). İnsan katılımcıların katıldığı çalışmalarda tüm prosedürler, 1964 Helsinki Deklarasyonu ve daha sonra yapılan değişiklikler uyarınca gerçekleştirilmiştir.

Conflict of Interest: No conflict of interest was declared by the authors.

Çıkar Çatışması: Yazarlar çıkar çatışması bildirmemişlerdir.

Financial Disclosure: This study was supported by Akdeniz University Research Fund Accounting (BAP Project no: TTU-2017-2289).

Finansal Destek: Bu çalışma Akdeniz Üniversitesi Araştırma Fonu Muhasebesi (BAP Proje no: TTU-2017-2289) tarafından desteklenmiştir.

Previous presentation: 28th National Congress of Pathology, Ankara, Turkey, 27-30 October 2018
Önceki sunum: 28. Ulusal Patoloji Kongresi, Ankara, Türkiye, 27-30 Ekim 2018

Published: 6/28/2020
Yayın Tarihi: 28.06.2020

Copyright © 2020 The Author(s)
Published by JOSAM

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND 4.0) where it is permissible to download, share, remix, transform, and build upon the work provided it is properly cited. The work cannot be used commercially without permission from the journal.



Introduction

Prostate carcinoma (PCa) is the most commonly diagnosed malignancy in men and its incidence is rapidly increasing. Many factors are associated with pathophysiology and progression of PCa [1]. Hormonal imbalances, genetic factors, and many factors such as carcinogens in the diet and infectious agents are all sources that have been determined to contribute to prostate carcinogenesis [1-4]. The prostatic epithelium is composed of two types of cells: Luminal epithelial cells and basal epithelial cells [4]. Primary prostate tumors are almost always characterized by a luminal phenotype suggesting a luminal origin, but recently it has been indicated that both types of these epithelial cells are possible cells of origin for prostate carcinogenesis.

The Notch was first discovered in 1917 by Thomas Hunt Morgan [6]. Later, Kidd et al. discovered that the loss of Notch expression not only affected the neurodevelopmental phenotype of this *Drosophila* strain, but also the morphology of the eye, wing, and hair [7]. In later studies, it was found that the disorder of Notch expression is associated with some diseases, for example, T-cell acute lymphoblastic leukemia (T-ALL) patients, CADASIL syndrome (cerebral autosomal-dominant arteriopathy with subcortical infarcts and leukoencephalopathy) and Alagille syndrome [8-10]. The Notch signaling pathway plays a significant role both in embryonic development and the determination of cell fate in organ homeostasis [11]. The main function of the Notch pathway is the regulation of bi-directional cell fate decisions. Furthermore, it has been shown that Notch activation in cancer cells causes abnormal cell proliferation. Mammals have four Notch receptors, Notch1, Notch2, Notch3 and Notch4. There are also five Notch ligands including Jagged1, Jagged2, Delta 1, Delta 3 and Delta 4. In addition, the ligands for the Notch receptor are divided into two groups: Delta-like ligands (DLL1, DLL3, and DLL4) and Serrate-like ligands (JAG1 and JAG2). The Notch gene encodes a 300 kD single-pass transmembrane receptor. The Notch receptor consists of two parts, one intracellular and one extracellular. The large extracellular region contains 36 consecutive EGF (Epidermal Growth Factor)-like repeats and 3 cysteine-rich LIN 12 repeats. EGF repeats allow the Notch receptor to interact with ligands. The intracellular part of the Notch receptor is also called Notch Intracellular Domain (NICD) [12-14].

Many cancer pathways have been investigated to date [15,16]. Recently, studies have been examining the Notch pathway in terms of prostate carcinogenesis, with no full elucidation [17]. Therefore, in this study, we aimed to investigate the frequency and clinicopathologic features associated with Notch1 gene expression using immunohistochemistry and real-time-polymerase chain reaction in patients with prostate cancer.

Materials and methods

Tissue samples

Tissue samples from prostate carcinoma patients who underwent surgery between 2010 and 2017 were selected from the archives of Department of Pathology. The study was performed in accordance with the principles of Declaration of Helsinki and approved by the Ethics Committee of Akdeniz

University (Date:30.11.2016, number:625). This study was supported by Akdeniz University Research Fund Accounting (BAP Project no: TTU-2017-2289). All our patients were sampled with suspicion of prostate carcinoma. The blocks with sufficient tumor tissue for the study were selected in historical order since 2017. Hematoxylin-Eosin stained preparations obtained from formalin-fixed, paraffin-embedded blocks were removed from the archive and re-examined. Sixty of our patients had prostate carcinoma; 33 samples were obtained with tru-cut biopsy, 16 with radical prostatectomy and 11 with prostate TUR. Twenty patients with benign prostatic hyperplasia were used as the control group. Notch1 gene expression was examined both immunohistochemically and by RT-PCR in 60 patients with prostate cancer and 20 with benign prostatic hyperplasia. The optimal blocks for immunohistochemistry and real-time-polymerase chain reaction (RT-PCR) were selected. The anamnesis, stage, diagnosis history, PSA values and treatment follow-up times for the cases were obtained from our hospital's automation system, patient follow-up files, and from the patients themselves or their relatives via telephone.

Immunohistochemistry

In retrospectively selected cases, sections with a thickness of 5 μ m were obtained from the appropriate paraffin blocks. Immunohistochemistry was performed using Cell Signaling Technology rabbit monoclonal Notch1(D1E11) XP[®] antibody (Cell Signaling Technology, Danvers, MA, USA) at 1:50 dilution for two hours. The staining procedure was performed on the DAKO Omnis autostainer (Agilent, Santa Clara, CA 95051, USA). Sections were examined using light microscopy (Zeiss, Oberkochen, Germany). Negative controls were performed by replacing the primary antibody with the nonimmune IgG in the same dilutions as the specific antibodies. Positive staining for Notch1 was observed in the cytoplasm and nucleus.

RNA Extraction and RT-PCR (Reverse Transcription Polymerase Chain Reaction)

Total RNA was extracted from FFPE tissue using Purelink FFPE RNA Isolation Kit (Cat No: K156002; Invitrogen) according to the manufacturer's instructions. The amount of total RNA in each sample is measured using NanoDrop (Maestrogen, Taiwan). All samples are diluted to a concentration of 25 ng/ μ l. The reverse transcription reaction was performed using High Capacity cDNA Reverse Transcription Kit (Appliedbiosystems by Thermo Fisher Scientific). Complementary DNA was prepared from 25 ng of isolated total RNA, with 10xRT Random Primers, according to the manufacturer's instructions. The program was as follows: 10 minutes at 25 °C, 2 hours at 37 °C, 5 minutes at 85 °C and at 4 °C thereafter on a polymerase chain reaction thermocycler (Applied Biosystems). The resulting cDNA was diluted to 1/5.

qPCR

The relative expression levels of Notch1 gene were determined by qPCR with TaqMan[®] Gene Expression Master Mix in Applied Biosystems 7900HT Real-Time PCR System and normalized to 18S. Primers and FAM-MGB hydrolysis probes were TaqMan[®] Gene expression assays on demand for Notch11 Hs01062014_m1 and 18S (Hs99999901_s1) (Applied biosystems by Thermo Fisher Scientific). All samples were

performed in duplicates. The PCR amplification program was as follows: 20 seconds at 95 °C, 40 cycles of 1 second at 95 °C and 20 seconds at 60 °C. In addition, as the non-template control, ddH₂O was analyzed for every plate. To assure that the amplification efficiencies for each real-time-PCR run and each gene were similar, the slope was adjusted for each sample. The data obtained from the qPCR was analyzed by the $\Delta\Delta C_t$ -method.

Statistical analysis

Data were evaluated with the SPSS 22® program. Descriptive statistics were presented as mean (standard deviation) and median (minimum-maximum) for normally and non-normally distributed values, respectively. Nominal variables were shown as number (n) and percentage (%). As there were two groups, the significance of the difference between the groups was evaluated with the Student's t-test or the Mann Whitney U-test. Categorical variables were evaluated with the Pearson Chi-square test. A value of $P < 0.05$ was considered statistically significant.

Results

Eighty cases, diagnosed at the Akdeniz University Medical Faculty Pathology Department between 2010 and 2017, were selected for this study. Of the patients with prostate adenocarcinoma, 20, 10, 14, 8 and 8 patients had Gleason scores of 6, 7, 8, 9 and 10, respectively. In the 80 cases we evaluated, the youngest and oldest patients were 46 and 81 years old, with a mean age of 65 years.

PSA values ranged between a minimum of 0.16 ng/ml and a maximum of 260 ng / ml. The Gleason score in our prostate adenocarcinoma cases ranged from a minimum of 6, to a maximum of 10. ISUP grade group were scored in 21 patients as ISUP-1, in eight patients as ISUP-2, in one patient as ISUP-3, in 14 patients as ISUP-4 and in 16 patients as ISUP-5. Notch1 gene expression was detected in 17 out of 60 patients by RT-PCR (Table 1). Thirty-three specimens were obtained with tru-cut biopsy, 16 with radical prostatectomy and 11 with prostate TUR material.

In immunohistochemical examination, cytoplasmic and nuclear staining in malignant gland epithelium was considered positive. In 17 of our 60 prostate carcinoma patients, a positive reaction was observed in the malignant gland epithelium. In our control group with benign prostatic hyperplasia, focal staining with Notch-1 was observed in the basal cells and smooth muscles in the stroma. In our control group, no staining was observed in the prostate gland epithelium. Immunohistochemically, going by the RT-PCR results, preexisting cytoplasmic and nuclear immunoreaction was observed in 17 of 60 patients (Figure 1, 2). A significant relationship was found between Notch1 gene expression, increase in Gleason scores ($P=0.007$), and ISUP grade groups (Figure 3).

The Notch1 gene expression of our patients with prostate adenocarcinoma were compared with those with benign prostatic hyperplasia, as evaluated with RT-PCR. Out of 60 patients with prostate adenocarcinoma, 17 had significantly higher levels of Notch1 gene expression than patients with benign prostatic hyperplasia.

In addition, in our study, 10 out of 17 patients with Notch1 gene expression were metastatic, and nine metastatic

distant organs were detected. We could not perform a statistical evaluation because our data on metastasis were incomplete. We found that Notch1 gene expression was significantly associated with Gleason score, ISUP grade group increase and PSA elevation (Table 2).

Table 1: Notch-1 gene expression of 60 patients with prostatic adenocarcinoma compared to patients with benign prostatic hyperplasia (Bold texts: Notch-1 positive patients)

Patient number	Notch1 CT	18SrRNA CT	Δ CT	$\Delta\Delta$ CT (8.595)	Fold difference ($2^{-\Delta\Delta$ CT)
1	31.98	21.64	10.34	1.745	0.298
2	27.64	15.43	12.21	3.615	0.082
3	33.95	26.14	7.81	-0.785	1.72
4	36.146	29.16	6.99	-1.605	3.042
5	29.22	14.78	14.44	5.845	0.017
6	34.37	27.69	6.68	-1.915	3.771
7	34.85	27.88	6.97	-1.625	3.08
8	30.93	25.50	5.43	-3.165	8.96
9	33.89	26.42	7.47	-1.125	2.181
10	35.11	24.58	10.53	1.935	0.262
11	29.44	16.01	13.43	4.835	0.035
12	35.69	26.76	8.93	0.335	0.79
13	33.97	21.83	12.14	3.545	0.09
14	29.98	16.18	13.8	5.205	0.027
15	33.214	25.079	8.135	-0.46	1.376
16	31.89	18.37	13.52	4.925	0.033
17	31.86	18.04	13.82	5.255	0.026
18	30.197	17.895	12.30	3.705	0.0389
19	34.38	25.15	9.23	0.635	0.64
20	36.45	27.96	8.49	-0.105	1.08
21	33.81	23.85	9.96	1.365	0.388
22	29.89	22.14	7.75	-0.845	1.80
23	34.18	28.63	5.55	-3.045	8.25
24	30.73	17.14	13.59	4.995	0.031
25	34.11	26.13	7.98	-0.615	1.53
26	35.241	27.249	7.99	-0.605	1.52
27	29.531	17.173	12.36	3.765	0.074
28	30.285	16.336	13.95	5.355	0.024
29	28.76	16.75	12.01	3.415	0.09
30	30.999	17.42	13.58	4.985	0.0316
31	29.536	16.135	13.40	4.805	0.058
32	34.501	24.496	10	1.405	0.378
33	31.473	17.895	13.58	4.985	0.032
34	29.86	15.97	13.89	5.295	0.03
35	31.861	20.559	11.30	2.705	0.153
36	31.793	18.893	12.9	4.305	0.051
37	30.1	17.87	12.23	3.635	0.081
38	33.63	23.63	10	1.405	0.38
39	34.43	24.38	10.05	1.455	0.36
40	33.918	25.714	8.20	-0.395	1.315
41	33.626	22.241	11.39	2.795	0.144
42	35.059	27.177	7.88	-0.715	1.64
43	31.431	19.075	12.36	3.765	0.074
44	35.19	27.78	7.41	-1.185	2.27
45	34.39	24.88	9.51	0.915	0.53
46	28.99	15.56	13.43	4.835	0.04
47	30.93	17.03	13.09	4.495	0.044
48	33.84	22.63	11.21	2.615	0.163
49	35.61	27.62	7.99	-0.605	1.52
50	31.08	15.99	15.09	6.495	0.011
51	30.0	18.29	11.71	3.115	0.12
52	29.099	19.392	9.71	1.115	0.461
53	30.338	15.53	14.81	6.215	0.013
54	30.508	17.225	13.28	4.685	0.0389
55	29.98	16.98	13	4.405	0.047
56	30.167	18.165	12	3.405	0.094
57	33.51	23.50	10.01	1.415	0.38
58	29.392	15.245	14.15	5.555	0.0213
59	29.60	18.76	10.84	2.245	0.22
60	29.299	21.406	7.89	-0.705	1.63

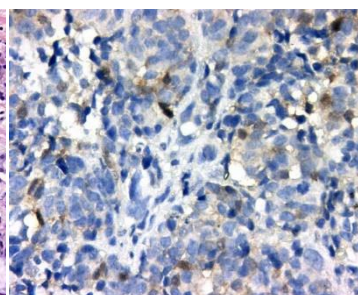
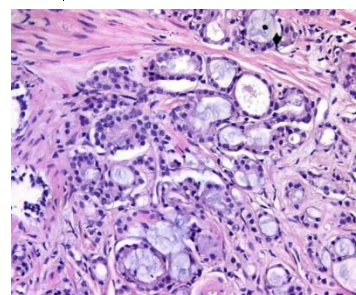


Figure 1: Prostate adenocarcinoma (H&E 20X)

Figure 2: Prostate adenocarcinoma positive with Notch1 (40X)

Table 2: Mean age, PSA, ISUP and Gleason scores of scores in Notch-1 positive and negative patients

	Notch-1 positive (n=17)		Notch-1 negative (n=43)		P-value
	mean	SD	mean	SD	
Age	67	9	65	8	0.407
ISUP Grade Group	3.82	1.47	2.58	1.67	0.008
Gleason Score	8.35	1.41	7.23	1.32	0.007
Serum PSA Level	184.51	432.84	24.67	45.10	0.018

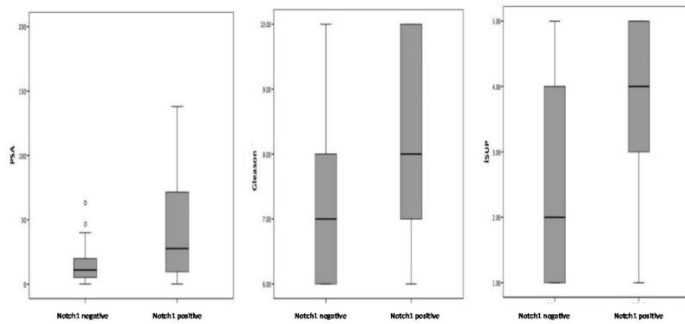


Figure 3: A significant relationship was found between Notch1 gene expression and Gleason score increase, ISUP score increase, and PSA elevation

Discussion

The aim of this study was to investigate the frequency and clinicopathologic features associated with Notch1 gene expression. We demonstrated that high Gleason scores, high ISUP grade group scores and high PSA levels significantly correlated with Notch1 gene expression.

It is well known that Notch signaling regulates normal and cancer development in many tissues including the prostate. The expression of Notch pathway elements clearly shows the regulation of Notch signaling pathway in prostate cancer in cases of established cancer cell lines, transgenic mouse models, and clinical tumor specimens.

The role of Notch ligands and Notch receptors in prostate tumorigenesis is not well defined. In a study in 2006, Notch1 mRNA expression has been reported to be significantly down-regulated in prostate cancers, thus suggesting a potential tumor suppressor role for Notch1 in prostate cancer [18]. This study also examined the expression of the remaining members of the Notch receptor family in prostate cancer but found no significant differences in mRNA levels for Notch-2-3-4 between benign and malignant prostate specimens [18]. Conversely, a study in 2007 revealed that Notch1 protein was overexpressed in malignant prostates compared to benign controls [19]. This suggests a potential carcinogenic role of Notch1 in prostate tumorigenesis. Notch1 protein expression, rather than Notch1 mRNA expression, is a more reliable indicator of Notch1 functionality. Researchers in a study showed that the Notch signal is associated with prostate development and cancer cell growth, and they correlated high grade localized PCa with the gene expression of the Notch signal. Because of the role of basal epithelial cells in the development of prostate carcinoma, they have suggested that PCa and basal epithelial cell relationship should be investigated and discussed [20]. A review of Notch pathway and prostate tissue revealed that Notch pathway was involved in prostate differentiation in benign prostate tissue and associated with lethal potential in prostate cancers [21]. The expression of Immunohistochemical Notch-1 in normal prostate tissue, prostatic intraepithelial neoplasia and PCa was investigated. As a result of this study, stronger staining was observed in prostatic intraepithelial neoplasia than prostatic adenocarcinomas [22].

Prostate cancer most frequently metastasizes to bone, brain, and lymph nodes. Therefore, the relationship between the metastasis of prostate cancers and Notch pathway was investigated [22-29]. As a result of these studies, metastatic prostate cancer was shown to have increased expression levels of

the Notch1 receptor ligand, when compared to benign prostate tissue and localized prostate cancer [22-28].

Surgical treatment and chemotherapy are used in the treatment of prostate cancers. Although chemotherapy agents used in prostate carcinomas have exceptionally good response at first, it is known that a rapid resistance to chemotherapeutic agents develops soon. Therefore, resistance mechanisms against chemotherapeutic agents have been investigated. Since the notch pathway is known to play a role in prostate carcinomas, its effect on chemotherapy resistance has been researched. It is concluded that the notch pathway is effective in the progression, metastasis, and chemotherapy resistance of prostate cancer [24,28].

To fully understand the role of Notch1 in prostate carcinogenesis, it may be necessary to assess the possibility that primary tumor growth and metastasis progression may be affected differently by Notch signaling. There is also a need for further study, more mouse models, and cell lines in this regard. If the role of Notch1 in prostate tumorigenesis is better understood, therapeutic strategies for Notch1 can be developed. Therefore, this is an important and urgent need, given the proposed use of anti-Notch compounds for the treatment of cancer, including prostate cancer.

Limitations

Our study had some limitations. Due to our financial limitation, we were able to study a small number of patients and evaluate Notch1 only.

Conclusions

The current study showed that a high Gleason score, high ISUP scores, and high PSA levels and Notch1 gene expression are significantly correlated. Due to the small number of studies related to this topic and the inadequate data obtained, it is important to conduct further studies with wider series to determine the prognosis of PCa patients and manage their treatment.

References

- Balistreri CR, Candore G, Lio D, Carruba G. Prostate cancer: from the pathophysiological implications of some genetic risk factors to translation in personalized cancer treatments. *Cancer Gene Therapy*. 2014;21(1):2.
- Mandair D, Rossi RE, Pericleous M, Whyand T, Caplin ME. Prostate cancer and the influence of dietary factors and supplements: a systematic review. *Nutrition & Metabolism*. 2014;11(1):30.
- Sherwood ER, Theyer G, Steiner G, Berg LA, Kozlowski JM, Lee C. Differential expression of specific cytokeratin polypeptides in the basal and luminal epithelia of the human prostate. *The Prostate*. 1991;18(4):303-14.
- Ghagane SC, Nerli RB, Hiremath MB, Wagh AT, Magdum PV. Incidence of prostate cancer at a single tertiary care center in North Karnataka. *Indian Journal of Cancer*. 2016;53(3):429.
- Humphrey PA. Histological variants of prostatic carcinoma and their significance. *Histopathology*. 2012;60(1):59-74.
- Morgan TH. "The theory of the gene." *The American Naturalist*. 1917;51:609:513-44.
- Kidd SIMON, Kelley MR, Young MW. Sequence of the notch locus of *Drosophila melanogaster*: relationship of the encoded protein to mammalian clotting and growth factors. *Molecular and cellular biology*. 1986;6(9):3094-108.
- Ellisen LW, Bird J, West DC, Soreng AL, Reynolds TC, Smith SD, et al. TAN-1, the human homolog of the *Drosophila* notch gene, is broken by chromosomal translocations in T lymphoblastic neoplasms. *Cell*. 1991;66(4):649-61.
- Pear WS, Aster JC, Scott ML, Hasserjian RP, Soffer B, Sklar J, et al. Exclusive development of T cell neoplasms in mice transplanted with bone marrow expressing activated Notch alleles. *Journal of Experimental Medicine*. 1996;183(5):2283-91.
- Oda T, Elkahoul AG, Pike BL, Okajima K, Krantz ID, Genin A, et al. Mutations in the human Jagged1 gene are responsible for Alagille syndrome. *Nature genetics*. 1997; 16(3):235.
- Artavanis-Tsakonas S, Rand MD, Lake RJ. Notch signaling: cell fate control and signal integration in development. *Science*. 1999;284(5415):770-6.
- Kopan R. Notch: a membrane-bound transcription factor. *Journal of Cell Science*. 2002;115(6):1095-1097.
- Ohishi K, Varnum-Finney B, Flowers D, Anasetti C, Myerson D, Bernstein ID. Monocytes express high amounts of Notch and undergo cytokine specific apoptosis following interaction with the Notch ligand, Delta-1. *Blood*. 2000;95(9):2847-54.
- Yağcı E, Güneş HV. Notch Sinyal Yoluğu ve Karsinogenez/Notch Signaling Pathway and Carcinogenesis. *Osmangazi Journal of Medicine*. 2017;39(1):109-16.
- Park JW, Lee JK, Phillips JW, Huang P, Cheng D, Huang J, et al. Prostate epithelial cell of origin determines cancer differentiation state in an organoid transformation assay. *Proceedings of the National Academy of Sciences*. 2016;113(16):4482-7.

- 16.Schulz W. Molecular biology of human cancers: an advanced student's textbook. Springer Science & Business Media. 2005.
- 17.Koch U, Radtke F. Dual Function of Notch Signaling in Cancer: Oncogene and Tumor Suppressor. In Targeting Notch in Cancer. Springer, New York, NY. 2018;55-86.
- 18.Wang XD, Leow CC, Zha J, Tang Z, Modrusan Z, Radtke F, et al. Notch signaling is required for normal prostatic epithelial cell proliferation and differentiation. *Developmental Biology*. 2006;290(1):66-80.
- 19.Brown MD, Gilmore PE, Hart CA, Samuel JD, Ramani VA, George NJ, et al. Characterization of benign and malignant prostate epithelial Hoechst 33342 side populations. *The Prostate*. 2007;67(13):1384-96.
- 20.Shou J, Ross S, Koeppen H, de Sauvage FJ, Gao WQ. Dynamics of notch expression during murine prostate development and tumorigenesis. *Cancer Research*. 2001;61(19):7291-7.
- 21.Carvalho FL, Simons BW, Eberhart CG, Berman DM. Notch signaling in prostate cancer: a moving target. *The Prostate*. 2014;74(9):933-45.
- 22.Soylu H, Acar N, Ozbey O, Unal B, Koksall IT, Bassorgun I, et al. Characterization of Notch signalling pathway members in normal prostate, prostatic intraepithelial neoplasia (PIN) and prostatic adenocarcinoma. *Pathology & Oncology Research*. 2016;22(1):87-94.
- 23.Kron KJ, Murison A, Zhou S, Huang V, Yamaguchi TN, Shiah YJ, et al. TMRSS2-ERG fusion co-opts master transcription factors and activates NOTCH signaling in primary prostate cancer. *Nature Genetics*. 2017;49(9):1336.
- 24.Zhu H, Zhou X, Redfield S, Lewin J, Miele L. Elevated Jagged-1 and Notch-1 expression in high grade and metastatic prostate cancers. *American Journal of Translational Research*. 2013;5(3):368.
- 25.Wang Z, Li Y, Ahmad A, Banerjee S, Azmi AS, Kong D, et al. Down-regulation of Notch-1 is associated with Akt and FoxM1 in inducing cell growth inhibition and apoptosis in prostate cancer cells. *Journal of Cellular Biochemistry*. 2011;112(1):78-88.
- 26.Zayzafoon M, Abdulkadir SA, McDonald JM. Notch signaling and ERK activation are important for the osteomimetic properties of prostate cancer bone metastatic cell lines. *Journal of Biological Chemistry*. 2004;279(5):3662-70.
- 27.Sethi N, Kang Y. Notch signalling in cancer progression and bone metastasis. *British Journal of Cancer*. 2011;105(12):805.
- 28.Domingo-Domenech J, Vidal SJ, Rodriguez-Bravo V, Castillo-Martin M, Quinn SA, Rodriguez-Barrueco R, et al. Suppression of acquired docetaxel resistance in prostate cancer through depletion of notch-and hedgehog-dependent tumor-initiating cells. *Cancer Cell*. 2012;22(3):373-88.
- 29.Çayır D, Bozkurt M, Gültekin S, Turan A. Complete scintigraphic resolution of a bone metastasis after androgen-deprivation therapy. *Journal of Surgery and Medicine*. 2019;3(12):899-8.

This paper has been checked for language accuracy by JOSAM editors.

The National Library of Medicine (NLM) citation style guide has been used in this paper.