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## Evaluation of the Cytotoxic Efficacy of Thymoquinone and Capsaicin in the SH-SY5Y Neuroblastoma Cell Line

Ayhan Çetinkaya\*<sup>1</sup>, Şeyda Karabörk<sup>2</sup>, Hümeysra Çelik<sup>3</sup>, İbrahim Ethem Torun<sup>4</sup>

<sup>1</sup>Bolu Abant İzzet Baysal University, Faculty of Medicine, Department of Physiology, Bolu, Turkey

[orcid.org/0000-0002-8212-7149](https://orcid.org/0000-0002-8212-7149)

<sup>2</sup>Bolu Abant İzzet Baysal University, Faculty of Medicine, Department of Microbiology, Bolu, Turkey

[orcid.org/0000-0002-9026-4485](https://orcid.org/0000-0002-9026-4485)

<sup>3</sup>Bolu Abant İzzet Baysal University, Faculty of Medicine, Department of Physiology, Bolu, Turkey

[orcid.org/0000-0002-3394-2438](https://orcid.org/0000-0002-3394-2438)

<sup>4</sup>Bolu Abant İzzet Baysal University, Faculty of Medicine, Department of Physiology, Bolu, Turkey

[orcid.org/0000-0001-7038-3336](https://orcid.org/0000-0001-7038-3336)

\*Corresponding author: [cetinkayaayhan@hotmail.com](mailto:cetinkayaayhan@hotmail.com)

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### Abstract

In our study, it was aimed to examine the effects of thymoquinone, the active ingredient of *Nigella sativa*, which has known anticancer activities, and capsaicin, which is an important part of the endocannabinoid system, on the neuroblastoma cell line SH-SY5Y cells. SH-SY5Y cells were grown in culture in conventional culture flasks in DMEM medium at 37 °C and 5% CO<sub>2</sub>. When the cells were 70-80% confluent, morphological changes were examined under an inverted microscope. The cells were passaged into 96 microplates, and after passage, different concentrations of thymoquinone (2.5; 5; 10; 25; 50; 100; 200; 300 µM/ml) and capsaicin (0.675; 1.25; 2.5; 5; 10; 20; 50; 100 µM/ml) were applied to the cells. After administration, cytotoxic effect and proliferation rates/cell proliferation were analyzed by the MTT method. When compared to the control group, cultured cells treated with 200 and 300 µM thymoquinone and 100 µM capsaicin had reduced cell proliferation at statistically significant levels ( $p < 0.05$ ). Demonstration of antiproliferative activity of thymoquinone and capsaicin on neuroblastoma shows that phytotherapeutic approaches can be evaluated in cancer.

**Key words:** Cell culture techniques, Cytotoxicity test, MTT formazan, Thymoquinone, Capsaicin, Neuroblastoma

### 1. Introduction

Neuroblastoma is the most common extracranial solid tumor among childhood cancers that originates from the neural crest and develops anywhere in the sympathetic nervous system (Maris, 2010). An average of 700 children are diagnosed with neuroblastoma each year in the United States (Ward et al., 2014). In

neuroblastoma cases, about half of which are metastatic at the time of diagnosis (Tolbert and Matthay, 2018), the disease may regress spontaneously or may progress aggressively (Chung et al., 2020). As well as a fully cured treatment is still not available today, modern protocols including induction chemotherapy, surgical resection, high-dose chemotherapy combined with autologous stem cell transplantation, external beam radiotherapy, immunotherapy, and differentiating agents have produced results that increase the three-year survival rate to 60% (Park et al., 2016). Besides high drug resistance (Marengo et al., 2018), most patients with neuroblastoma, of whom 50% survive despite multimodality therapy, are severely affected by treatment complications. At this stage, patients with neuroblastoma urgently need new treatment targets with reduced toxicity and a high treatment rate (Park et al., 2020).

The SH-SY5Y neuroblastoma cell line is frequently used in neurodegenerative disease studies (Molina-Jimenez et al., 2003). Because it is unethical to obtain a primary human neuron, SH-SY5Y allows the study of human-specific proteins not found in rodent cultures, making the cell line valuable (Encinas et al., 2000). It is a unique experimental material for new molecule studies to be developed for neuroblastoma.

Phytochemicals are widely used in cancer as an antioxidative defense mechanism against DNA breaks caused by oxidative damage induced by oxidant radicals (Ames et al., 1993; Rajput and Mandal, 2011). *Nigella sativa* extract thymoquinone, which is frequently used in phytotherapy, is known to inhibit the proliferation, migration and invasion of cancer cells (Imran et al., 2018). Similarly, a different phytotherapeutic molecule, capsaicin (trans-8-methyl-N-vanillyl-6-nonenamide), is the primary and natural component of red pepper species (Cordell and Araujo, 1993). Capsaicinoids, which are included in a group of chemicals known as vanilloids, have many pharmacological and physiological activities such as anti-inflammatory, anti-apoptotic, antioxidant, anti-cancer, and neuroprotective properties (Chaudhary et al., 2019; Bujak et al., 2019; Zhang et al., 2019). Within the framework of these features, it is important to examine and compare thymoquinone and capsaicin in neuroblastoma cell lines to increase awareness of phytotherapeutic approaches to neurological cancers. Although they have been shown to be effective in many types of cancer: breast (Adinew et al., 2021; Dupoirion et al., 2022), prostate (Alshyarba et al., 2021; Sanchez et al., 2022), colorectal (Karim et al., 2022; Hou et al., 2019) and neuroblastoma (Firdaus et al., 2018; Baek et al., 2008), there is no study in the literature comparing the effectiveness of the two.

For this purpose, we aim to investigate the anti-proliferative effects of thymoquinone and capsaicin in SH-SY5Y neuroblastoma cells isolated from human tissue to develop treatment alternatives in neuroblastoma.

## **2. Material and Methods**

Our research was carried out under in vitro conditions by using the Cell Culture Laboratory in Bolu Abant İzzet Baysal University Faculty of Medicine, Department of Physiology. Cell culture studies do not require ethics committee approval.

## **2.1. Drugs and reagents**

Thymoquinone (Sigma-Aldrich Chemical Co., cat no: 490-91-5, St. Louis, Missouri, USA) and capsaicin (Tocris, cat no: 545395-94-6, UK) were acquired to be used in the study. Penicillin-streptomycin as antibiotic/antimycotic (Capricorn Scientific Ebsdorfergrund, lot no: CP21-4278, Germany), fetal bovine/bovine serum for cell culture experiment (Sigma-Aldrich, Schnelldorf, lot no: BCBT7617, Germany), dimethyl sulfoxide (DMSO, Pan-Biotek, Aidenbach, cat no: P60-36720100), trypsin/EDTA solution (Hyclone, Logan, UT, cat no: SH30042.01, USA), phosphate buffered water (PBS, Sigma-Aldrich Chemical Co., lot no: 6730915, St. Louis, Missouri, USA), and Dulbecco's Modified Eagle Medium/F-12 (DMEM/F-12) medium (Pan-Biotech, Aidenbach, cat no: P04-41250, Germany) were used. In order to obtain the final concentrations of thymoquinone and capsaicin in the in vitro experiment, thymoquinone was dissolved and capsaicin was diluted in ethanol at different concentrations: For thymoquinone 2.5, 5, 10, 25, 50, 100, 200, 300  $\mu\text{M}$  /ml; for capsaicin (0.675, 1.25, 2.5, 5, 10, 20, 50, 100). We used our work control groups for the MTT assay. For the metabolic activity assays of control groups were untreated cells (Cel-) exposed to the maximum solvent concentrations with ethanol and DMSO. Both our working and control groups are presented as the mean  $\pm$  SD of each with two replicates. Each treatment was compared to the 100% metabolic activity of the negative control (untreated cells).

## **2.2. Cell culture**

The SH-SY5Y human neuroblastoma cell line purchased from ATCC (ATCC CRL-2266, Manassas, VA, USA) was used in the study. The cell culture protocol was completed with reference to previously published studies (Shipley et al., 2016). In brief, cells were quickly thawed in a 37 °C water bath and then centrifuged at 3000 rpm for 4 minutes. Next, all cells were cultured in 75 cm<sup>2</sup> flasks/flasks having fortified DMEM/F12 mixture/full medium supplemented with 10% heat-inactivated FBS and 1% penicillin-streptomycin at 37 °C in a humidified incubator/oven with 5% CO<sub>2</sub>. One day later, the growth medium was rapidly changed to take away any DMSO that may be present in the freezing medium. Cells were maintained at the log phase and supplemented/replaced with fresh medium every 3-4 days. Cell morphologies and growth rates of the cell line were monitored daily under an inverted microscope (Olympus CKX41, Tokyo, Japan) and passaged by separation with 0.25% Trypsin-EDTA when cells reached 70-80% confluence.

## **2.3. Thymoquinone and capsaicin treatment**

After reaching the appropriate confluence, the cells were passaged. Thymoquinone at concentrations of 2.5; 5; 10; 25; 50; 100; 200; 300  $\mu\text{M}$ /ml and capsaicin at concentrations of 0.675; 1.25; 2.5; 5; 10; 20; 50; 100  $\mu\text{M}$ /ml were prepared and given to the cells in equal volumes as 100  $\mu\text{l}$  and incubated for the defined times (24 hours) used. For both drugs and each dose, 2 wells were inoculated and the cells were incubated at 37 °C in a 5% CO<sub>2</sub> air mixture in a cell culture incubators, very high relative (around 90-95%) humidity environment. Humidity is homogeneous and created by evaporation from the water tank inside the incubator.

## **2.4. MTT assay**

Trypan blue was used to establish cell viability and cell quantification. BioRad TC20 Automated Cell Counter was used to calculate cell viability and cell numbers. After cell counting in 96-well plates, nearly  $1.5 \times 10^4$  cells/well were seeded in a total volume of 200  $\mu$ L. Plates were then incubated at 37 °C for 24 hours for cell attachment.

When the cells in the flask reached 70- 80% confluence, thymoquinone (2.5; 5; 10; 25; 50; 100; 200; 300  $\mu$ M/ml) and capsaicin (0.675; 1.25; 2.5; 5; 10; 20; 50; 100  $\mu$ M/ml) in different dilutions were added to the 96-well plate(s) and kept in incubation for 24 and 48 hours. Cells incubated in 10% FBS were used as positive control and the viability of the cells was determined by MTT Method (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl- tetrazolium bromide) (Serva, Germany). The MTT test is a colorimetric method used to find out cell viability. It is based on the ability of NADPH-dependent cellular oxidoreductase enzymes to alter the tetrazolium MTT into insoluble formazan, which is purple. Briefly, 10  $\mu$ l of MTT reagent was added to each well and the 96-well plate was incubated at 37 °C for 4 hours, then DMSO was added to the cells. Absorbance values/Change in Color were read at 570 nm with a colorimetric reader (spectrophotometer) (Epoch BioTek Instruments, Inc., Highland Park). The most suitable proliferative and inhibitory doses of thymoquinone and capsaicin were determined according to the cells. Thus, the effect of thymoquinone and capsaicin on the viability of cultured cells and the period of the effective dose were determined.

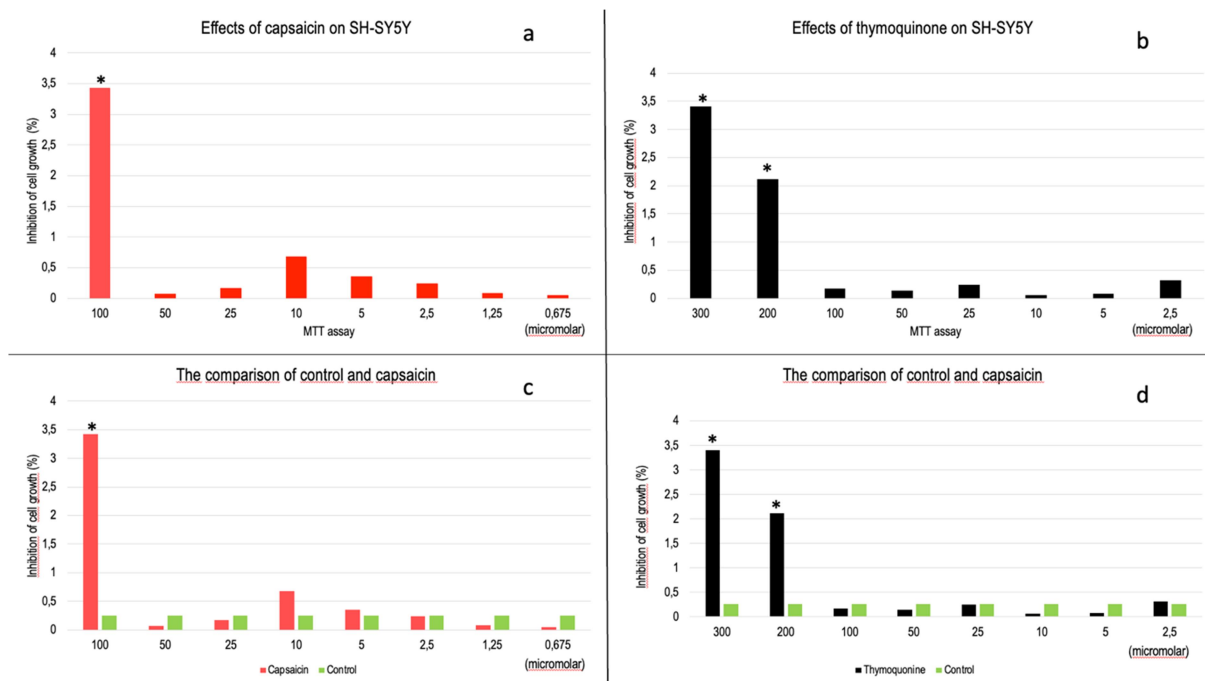
## **2.5. Statistical analysis**

Statistical analyzes were made with SPSS 25.0 package statistics program. Cytotoxicity experiments were performed in duplicate and GraphPad Prism 5.0 software (GraphPad Software, San Diego, USA) was used for data analysis. Descriptive statistics were given as number of units (n), percentage (%), mean  $\pm$  standard deviation ( $\bar{x} \pm SD$ ) median (Q1-Q3) values. The normal distribution of the datasets were evaluated with the Kolmogorov-Smirnov (K-S) Test. Mann-Whitney U Test was used for nonparametric distributed variables as a multiple comparison test.  $p < 0.05$  was considered statistically significant.

## **3. Results and Discussion**

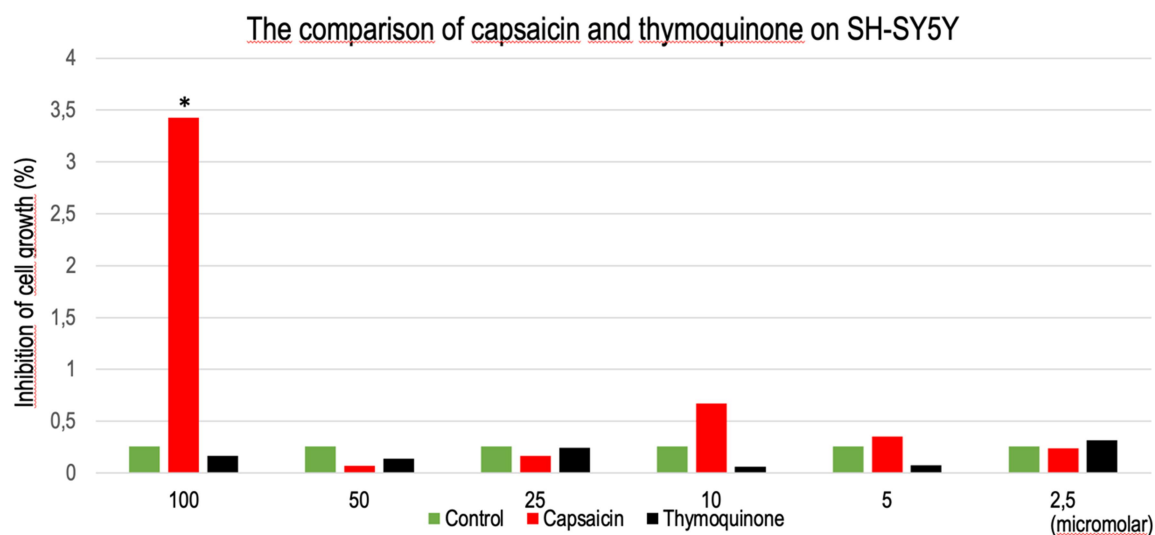
Thymoquinone (200 and 300  $\mu$ M) decreased ( $p < 0.05$ ) the cell viability, while the other concentrations of thymoquinone did not alter the viability compared to medium-treated cells (Figure 1). Capsaicin (100  $\mu$ M) decreased ( $p < 0.05$ ) the cell viability, while the other concentrations of capsaicin did not alter the viability compared to medium-treated cells (Figure 1).

The same doses of thymoquinone and capsaicin were compared. It was observed that capsaicin was found to be more cytotoxic when both drugs are administered at the same molarity in different wells: 5; 10; 100  $\mu$ M on SH-SY5Y cells. Besides it was observed that thymoquinone was found to be more cytotoxic when both drugs are administered at the same molarity in different wells: 2,5; 25; 50 on SH-SY5Y cells (Figure 2).



**Figure 1.** a. Effect of different doses of capsaicin on SH-SY5Y cell line. b. Effect of different doses of thymoquinone on SH-SY5Y cell line. c. Comparison of the control and capsaicin SH-SY5Y cell line. d. Comparison of the control and thymoquinone SH-SY5Y cell line.

\*:  $p < 0.05$ . There are significantly differences on 100  $\mu\text{M}$  capsaicin-treated cells and 200 and 300  $\mu\text{M}$  thymoquinone-treated cells versus non-treated control cells, each with the two replicates.



**Figure 2.** Comparison of the same doses of capsaicin and thymoquinone in SH-SY5Y cell line, each with the two replicates.

In the present study, the effectiveness of capsaicin and thymoquinone molecules on cell viability in the SH-SY5Y neuroblastoma cell line was evaluated, and it was observed that a 100  $\mu\text{M}$  dose of capsaicin and 200 and 300  $\mu\text{M}$  doses of thymoquinone decreased the number of neuroblastoma cells.

While phytotherapy was evaluated in the shadow of synthetic drugs in the 20<sup>th</sup> century (Kueete and Efferth, 2011) due to the recommendation of non-health professionals, it has come to the fore again in recent years due to the growing interest in medicinal plants (Falzon and Balobanova, 2017) so much so that today many drugs are obtained from plants. The fact that it has against cancer toxicity and improves health constitutes makes it a potential drug target. In this respect, intensive study of phytochemicals in medicine by researchers led to the award of the Nobel Physiology Prize to a phytotherapeutic product in 2015; a scientist named Youyou Tu introduced artemisinin in the plant *Artemisia annua*, known in Chinese medicine, to the literature by recommending it for the treatment of malaria (Efferth et al., 2015).

Due to surgical difficulties, chemotherapeutic side effects, and neurodegenerative processes, the demonstration of the effects of phytochemicals is important today to increase patient survival, especially in the field of neurological cancers. Thymoquinone, one of the active components of *Nigella sativa*, which has a strong traditional use among medicinal plants, is widely studied due to its pharmacological properties (Samarghandian et al., 2018). Although the molecular action pathways have not been fully revealed yet, thymoquinone has immunomodulatory, antioxidant, anticancer, antiapoptotic (Akter et al., 2021) and anti-inflammatory properties (Amin and Hossainzadeh, 2016) in healthy cells, and antiproliferative effects in cancerous cells (Aslan et al., 2021). It also has curative effects in neurological diseases such as epilepsy (Beyazçiçek et al., 2016), Alzheimer's (Elibol et al., 2020), depression (Safhi et al., 2019), and Parkinson Disease (Uddin et al., 2021). Thymoquinone has been shown to be effective in breast (Talib, 2017), prostate (Kolli-Bouhafs et al., 2012), blood (Hansen et al., 2003), bone (Thummuri et al., 2015), and colon (Lei et al., 2015) cancers. In the present study, 200 and 300  $\mu\text{M}$  doses of thymoquinone in the neuroblastoma SH-SY5Y cell line effectively reduced cell viability and supported the anticancer property of thymoquinone.

Capsaicin is the primary and naturally occurring component found in red pepper species and is responsible for the pungency of peppers (Cordell and Araujo, 1993). It shows neuroprotective effects in neurodegenerative diseases as an analgesic in the treatment of neuropathic pain (Ilie et al., 2019) by reducing beta-amyloid accumulation in Alzheimer's Disease (Wang et al., 2020) and oxidative stress in Parkinson's Disease (Siddique et al., 2018). It has antiproliferative and cytotoxic effects on many peripheral cancer cells such as breast (Chen et al., 2021), prostate (Zhu et al., 2020), colon (Nisari and Eroz, 2020) and oral cancers (Chang et al., 2020). It is also known to exert apoptotic effects on brain-derived tumors such as glioblastoma (Xie et al., 2016). In our study, the use of capsaicin in neuroblastoma SH-SY5Y cell line with the cytotoxic effect of 100  $\mu\text{M}$  dose in neurological cancers was also demonstrated.

Description of the action pathways of thymoquinone and capsaicin, which have been the potential drugs for the future, will allow to know both molecules better. Thymoquinone, has been shown to increase reactive oxygen products in brain cancer cell lines and trigger apoptosis by reducing the mitochondrial membrane potential (Firdaus et al., 2018). Similarly, it is known that capsaicin triggers the formation of superoxide radicals and initiates apoptosis by disrupting the intracellular mitochondrial membrane status in many cancer cell lines (Hail and Lotan et al., 2002). The similarity of the action mechanisms of both capsaicin and thymoquinone against

SH-SY5Y cells led us to compare the two molecules. Our study shows that the dose-response assessment of both drugs is not linear, as capsaicin at 100, 10, and 5 micromolar and thymoquinone at 2.5, 25 and 50 micromolar are more cytotoxic to SH-SY5Y cells when similar doses are compared. The limitation is that the signaling pathways of capsaicin and thymoquinone have not been evaluated by molecular, histopathologic and serologic techniques.

#### **4. Conclusion**

We anticipate that information from this research article would be capitalized for the future development of thymoquinone and capsaicin as potential therapeutic agents to be applied for neuroblastoma and the formulations of popular phytochemicals to promote optimum health and reduce cancers. In addition, it will be valuable for further studies to investigate the potential beneficial effects of thymoquinone and capsaicin in all aspects with animal experiments and clinical studies.

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#### **Conflicts of Interests**

There is no conflict of interest between authors.

#### **Statement contribution of the authors**

This study's experimentation, analysis and writing, etc. all steps were made by the authors.

#### **References**

- Adinew, G. M., Taka, E., Mochona, B., Badisa, R. B., Mazziro, E. A., Elhag, R., & Soliman, K. F. A. (2021). Therapeutic potential of thymoquinone in triple-negative breast cancer prevention and progression through the modulation of the tumor microenvironment. *Nutrients*, *14*, 79. <https://doi.org/10.3390/nu14010079>
- Akter, Z., Ahmed, F. R., Tania, M., & Khan, M. A. (2021). Targeting inflammatory mediators: an anticancer mechanism of thymoquinone action. *Current Medicinal Chemistry*, *28*, 80-92. <https://doi.org/10.2174/0929867326666191011143642>
- Alshyarba, M., Otifi, H., Al Fayi, M., A Dera, A., & Rajagopalan, P. (2021). Thymoquinone inhibits IL-7-induced tumor progression and metastatic invasion in prostate cancer cells by attenuating matrix metalloproteinase activity and Akt/NF- $\kappa$ B signaling. *Biotechnology and Applied Biochemistry*, *68*, 1403-1411. <https://doi.org/10.1002/bab.2062>
- Ames, B. N., Shigenaga, M. K., & Gold, L. S. (1993). DNA lesions, inducible DNA repair, and cell division: three key factors in mutagenesis and carcinogenesis. *Environmental Health Perspectives*, *101*, 35-44. <https://doi.org/10.1289/ehp.93101s535>

- Amin, B., & Hosseinzadeh, H. (2016). Black cumin (*Nigella sativa*) and its active constituent, thymoquinone: an overview on the analgesic and anti-inflammatory effects. *Planta Medica*, 82, 8-16. <https://doi.org/10.1055/s-0035-1557838>
- Aslan, M., Afşar, E., Kırımlıoğlu, E., Çeker, T., & Yılmaz, Ç. (2021). Antiproliferative effects of thymoquinone in MCF-7 breast and HepG2 liver cancer cells: possible role of ceramide and ER stress. *Nutrition and Cancer*, 73, 460-472. <https://doi.org/10.1080/01635581.2020.1751216>
- Baek, Y. M., Hwang, H. J., Kim, S. W., Hwang, H. S., Lee, S. H., Kim, J. A., & Yun, J. W. (2008) A comparative proteomic analysis for capsaicin-induced apoptosis between human hepatocarcinoma (HepG2) and human neuroblastoma (SK-N-SH) cells. *Proteomics*, 8, 4748-67. <https://doi.org/10.1002/pmic.200800094>
- Beyazcicek, E., Ankarali, S., Beyazcicek, O., Ankarali, H., Demir, S., & Ozmerdivenli, R. (2016). Effects of thymoquinone, the major constituent of *Nigella sativa* seeds, on penicillin-induced epileptiform activity in rats. *Neurosciences (Riyadh)*, 21, 131-137. <https://doi.org/10.17712/nsj.2016.2.20150781>
- Bujak, J. K., Kosmala, D., Szopa, I. M., Majchrzak, K. & Bednarczyk, P. (2019). Inflammation, Cancer and Immunity-Implication of TRPV1 Channel. *Frontiers in Oncology*, 9, 1087. <https://doi.org/10.3389/fonc.2019.01087>
- Chang, C. F., Islam, A., Liu, P. F., Zhan, J. H., & Chueh, P. J. (2020). Capsaicin acts through tNOX (ENOX2) to induce autophagic apoptosis in p53-mutated HSC-3 cells but autophagy in p53-functional SAS oral cancer cells. *American Journal of Cancer Research*, 10, 3230-3247. <https://doi.org/10.1016/j.jff.2022.104934>
- Chaudhary, A., Gour, J. K. & Rizvi, S. I. (2019). Capsaicin has potent anti-oxidative effects in vivo through a mechanism which is nonreceptor mediated. *Archives of Physiology and Biochemistry*, 128, 141-147. <https://doi.org/10.1080/13813455.2019.1669056>
- Chen, M., Xiao, C., Jiang, W., Yang, W., Qin, Q., Tan, Q., Lian, B., Liang, Z., & Wei, C. (2021). Capsaicin inhibits proliferation and induces apoptosis in breast cancer by down-regulating FBI-1-Mediated NF-κB pathway. *Drug Design, Development and Therapy*, 15, 125-140. <https://doi.org/10.2147/DDDT.S269901>
- Chung, C., Boterberg, T., Lucas, J., Panoff, J., Valteau-Couanet, D., Hero, B., Bagatell, R., & Hill-Kayser, C. E. (2021). Neuroblastoma. *Pediatric Blood & Cancer*, 2, 28473. <https://doi.org/10.1002/pbc.28473>
- Cordell, G. A., & Araujo, O. E. (1993). Capsaicin: identification, nomenclature, and pharmacotherapy. *The Annals of Pharmacotherapy*, 27, 330-336. <https://doi.org/10.1177/106002809302700316>
- Dupoiron, D., Jubier-Hamon, S., Seegers, V., Bienfait, F., Pluchon, Y. M., Lebec, N., Jaoul, V., & Delorme, T. (2022). Peripheral neuropathic pain following breast cancer: Effectiveness and tolerability of high-concentration capsaicin patch. *Journal of Pain Research*, 15, 241-255. <https://doi.org/10.2147/JPR.S341378>
- Efferth, T., Zacchino, S., Georgiev, M. I., Liu, L., Wagner, H., & Panossian, A. (2015). Nobel Prize for artemisinin brings phytotherapy into the spotlight. *Phytotherapy*, 22, 1-3. <https://doi.org/10.1016/j.phymed.2015.10.003>
- Elibol, B., Beker, M., Terzioglu-Usak, S., Dalli, T., & Kilic, U. (2020). Thymoquinone administration ameliorates Alzheimer's disease-like phenotype by promoting cell survival in the hippocampus of amyloid beta<sub>1-42</sub> infused rat model. *Phytotherapy*, 79, 153324. <https://doi.org/10.1016/j.phymed.2020.153324>



- Encinas, M., Iglesias, M., Liu, Y., Wang, H., Muhaisen, A., Cena, V., Gallego, C., & Comella, J. X. (2000). Sequential treatment of SH- SY5Y cells with retinoic acid and brain-derived neurotrophic factor gives rise to fully differentiated, neurotrophic factor-dependent, human neuron-like cells. *Journal of Neurochemistry*, *75*, 991-1003. <https://doi.org/10.1046/j.1471-4159.2000.0750991.x>
- Falzon, C. C., & Balabanova, A. (2017). Phytotherapy: An introduction to herbal medicine. *Primary Care*, *44*, 217-227. <https://doi.org/10.1016/j.pop.2017.02.001>
- Firdaus, F., Zafeer, M. F., Anis, E., Ahmad, F., Hossain, M., Ali, A., & Afzal, M. (2018). Evaluation of phyto-medicinal efficacy of thymo- quinone against Arsenic induced mitochondrial dysfunction and cytotoxicity in SH-SY5Y cells. *Phytomedicine*, *15*, 224-230, <https://doi.org/10.1016/j.phymed.2018.09.197>
- Hail, N., & Lotan, R. (2002). Examining the role of mitochondrial respiration in vanilloid- induced apoptosis. *Journal of the National Cancer Institute*, *94*, 1281-1292. <https://doi.org/10.1093/jnci/94.17.1281>
- Hansen, J. T., Benghuzzi, H., Tucci, M., & Cason, Z. (2003). The role of black seed in the proliferation and biochemical marker levels of Hep-2 cells. *Biomedical Sciences Instrumentation*, *39*, 371-376.
- Hou, N., He, X., Yang, Y., Fu, J., Zhang, W., Guo, Z., Hu, Y., Liang, L., Xie, W., Xiong, H., Wang, K., & Pang, M. (2019). TRPV1 induced apoptosis of colorectal cancer cells by activating Calcineurin-NFAT2-p53 signaling pathway. *Biomed Research International*, *30*, 6712536. <https://doi.org/10.1155/2019/6712536>
- Ilie, M. A., Caruntu C., Tampa M., Georgescu S. R., Matei C., Negrei C., Ion R. M., Constantin C., Neagu M., & Boda D. (2019). Capsaicin: physicochemical properties, cutaneous reactions and potential applications in painful and inflammatory conditions. *Experimental and Therapeutic Medicine*, *18*, 916-925 <https://doi.org/10.3892/etm.2019.7513>
- Imran, M., Rauf, A., Khan, I. A., Shahbaz, M., Qaisrani, T. B., Fatmawati, S., Abu-Izneid, T., Imran, A., Rahman, K. U., & Gondal, T. A. (2018). Thymoquinone: A novel strategy to combat cancer: A review. *Biomedicine & Pharmacotherapy*, *106*, 390-402. <https://doi.org/10.1016/j.biopha.2018.06.159>
- Karim, S., Burzangi, A. S., Ahmad, A., Siddiqui, N. A., Ibrahim, I. M., Sharma, P., Abualsunun, W. A., & Gabr, G. A. (2022). PI3K-AKT pathway modulation by thymoquinone limits tumor growth and glycolytic metabolism in colorectal cancer. *International Journal of Molecular Sciences*, *23*, 2305. <https://doi.org/10.3390/ijms23042305>
- Kolli-Bouhaf, K., Boukhari, A., Abusnina, A., Velot, E., Gies, J. P., Lugnier, C., & Rond, P. (2012). Thymoquinone reduces migration and invasion of human glioblastoma cells associated with FAK, MMP-2 and MMP-9 down-regulation. *Investigational New Drugs*, *30*, 2121-2131. <https://doi.org/10.1007/s10637-011-9777-3>
- Kuete, V., & Efferth, T. (2011). Pharmacogenomics of Cameroonian traditional herbal medicine for cancer therapy. *Journal of Ethnopharmacology*, *137*, 752-766. <https://doi.org/10.1016/j.jep.2011.06.035>
- Lei, X., Lv, X., Liu, M., Yang, Z., Ji, M., Guo, X., Dong, W. (2012). Thymoquinone inhibits growth and augments 5-fluorouracil-induced apoptosis in gastric cancer cells both in vitro and in vivo. *Biochemical and Biophysical Research Communication*, *417*, 864-868. <https://doi.org/10.1016/j.bbrc.2011.12.063>
- Marengo, B., Monti, P., Miele, M., Menichini, P., Ottaggio, L., Foggetti, G., Pulliero, A., Izzotti A., Speciale, A., Garbarino, O., Traverso, N., Fronza, G., & Domenicotti, C. (2018). Etoposide-resistance in a neuroblastoma

- model cell line is associated with 13q14. 3 mono-allelic deletion and miRNA-15a/16-1 down-regulation. *Scientific Reports*, 8, 1-15. <https://doi.org/10.1038/s41598-018-32195-7>
- Maris, J. M. (2010). Recent advances in neuroblastoma. *The New England Journal of Medicine*, 362, 2202-2211 <https://doi.org/10.1056/NEJMra0804577>
- Molina-Jimenez, M. F., Sanchez-Reus, M. I., & Benedi, J. (2003). Effect of fraxetin and myricetin on rotenone-induced cytotoxicity in SH-SY5Y cells: comparison with N-acetylcysteine. *European Journal of Pharmacology*, 472, 81-87. [https://doi.org/10.1016/s0014-2999\(03\)01902-2](https://doi.org/10.1016/s0014-2999(03)01902-2)
- Nisari, M., & Eröz, R. (2020). Does capsaicin have therapeutic benefits in human colon adenocarcinoma? Selection of the most reliable dose via AgNOR. *Turkish Journal of Medical Sciences*, 50, 1076-1081. <https://doi.org/10.3906/sag-2003-251>
- Park, J. R., Kreissman, S. G., London, W. B., Naranjo, A., Cohn, S. L., Hogarty, M. D., Tenney, S. C., Haas-Kogan, D., Shaw, P. J., Geiger, J. D., Doski, J. J., Gorges, S. W., Khanna, G., Voss, S. D., Maris, J. M., Grupp, S. A. & Diller, L. (2016). A phase III randomized clinical trial (RCT) of tandem myeloablative autologous stem cell transplant (ASCT) using peripheral blood stem cell (PBSC) as consolidation therapy for high risk neuroblastoma (HR-NB): a Children's Oncology Group (COG) study. *Journal of Clinical Oncology*, 34, LBA3-LBA. [https://doi.org/10.1200/JCO.2016.34.18\\_suppl.LBA3](https://doi.org/10.1200/JCO.2016.34.18_suppl.LBA3)
- Park, J. A., & Cheung, N. V. (2020). Targets and antibody formats for immunotherapy of neuroblastoma. *Journal of Clinical Oncology*, 38, 1836-1848. <https://doi.org/10.1200/JCO.19.01410>
- Rajput, S., & Mandal, M. (2012). Antitumor promoting potential of selected phytochemicals derived from spices. *European Journal of Cancer Prevention*, 21, 205-215. <https://doi.org/10.1097/CEJ.0b013e32834a7f0c>
- Safhi, M. M., Qumayri, H. M., Masmali, A. U. M., Siddiqui, R., Alam, M. F., Khan, G., & Anwer, T. (2019). Thymoquinone and fluoxetine alleviate depression via attenuating oxidative damage and inflammatory markers in type-2 diabetic rats. *Archives of Physiology and Biochemistry*, 125, 150-155. <https://doi.org/10.1080/13813455.2018.1443141>
- Samarghandian, S., Farkhondeh, T., & Samini, F. (2018). A review on possible therapeutic effect of Nigella sativa and thymoquinone in neurodegenerative diseases. *CNS & Neurological Disorders-Drug Targets*, 17, 412-420. <https://doi.org/10.2174/1871527317666180702101455>
- Sánchez, B. G., Bort, A., Mora-Rodríguez, J. M., & Díaz-Laviada, I. (2022). The natural chemotherapeutic capsaicin activates AMPK through LKB1 Kinase and TRPV1 receptors in prostate cancer cells. *Pharmaceutics*, 14, 329. <https://doi.org/10.3390/pharmaceutics14020329>
- Shipley, M.M., Mangold, C.A., & Szpara, M.L. (2016). Differentiation of the SH-SY5Y human neuroblastoma cell line. *Journal of Visualized Experiments*, 108, 53193 <https://doi.org/10.3791/53193>
- Siddique, Y. H., Naz, F., & Jyoti, S. (2018). Effect of capsaicin on the oxidative stress and dopamine content in the transgenic Drosophila model of Parkinson's disease. *Acta Biologica Hungarica*, 69, 115-124. <https://doi.org/10.1556/018.69.2018.2.1>
- Talib W. H. (2017). Regressions of breast carcinoma syngraft following treatment with piperine in combination with thymoquinone. *Journal of Environmental Sciences*, 3, 85. <https://doi.org/10.3390/scipharm85030027>

- Thummuri, D., Jeengar, M. K., Shrivastava, S., Nemani, H., Ramavat, R. N., Chaudhari, P., & Naidu, V. G. (2015). Thymoquinone prevents RANKL-induced osteoclastogenesis activation and osteolysis in an in vivo model of inflammation by suppressing NF-KB and MAPK signalling. *Pharmacological Research*, 99, 63-73. <https://doi.org/10.1016/j.phrs.2015.05.006>
- Tolbert, V. P., & Matthay, K. K. (2018). Neuroblastoma: clinical and biological approach to risk stratification and treatment. *Cell and Tissue Research*, 372, 195-209. <https://doi.org/10.1007/s00441-018-2821-2>
- Uddin, M. N., Hoq, M. I., Jahan, I., Siddiqui, S. A., Clinton, C. D., Ibrahim, M., Islam, M. S., & Jakaria, M. (2021). The mechanistic role of thymoquinone in parkinson's disease: Focus on neuroprotection in pre-clinical studies. *Current Molecular Pharmacology*, 14, 1083-1092. <https://doi.org/10.2174/1874467214666210105140944>
- Wang, J., Sun, B. L., Xiang, Y., Tian, D. Y., Zhu, C., Li, W. W., Liu, Y. H., Bu, X. L., Shen, L. L., Jin, W. S., Wang, Z., Zeng, G. H., Xu, W., Chen, L. Y., Chen, X. W., Hu, Z., Zhu, Z. M., Song, W., Zhou, H. D., Yu, J. T., & Wang, Y. J. (2020). Capsaicin consumption reduces brain amyloid-beta generation and attenuates Alzheimer's disease-type pathology and cognitive deficits in APP/PS1 mice. *Translational Psychiatry*, 10, 230. <https://doi.org/10.1038/s41398-020-00918-y>
- Ward, E., De Santis, C., Robbins, A., Kohler, B., & Jemal, A. (2014). Childhood and adolescent cancer statistics. *CA: Cancer Journal for Clinicians*, 64, 83-103. <https://doi.org/10.3322/caac.21219>
- Xie, L., Xiang, G. H., Tang, T., Tang, Y., Zhao, L. Y., Liu, D., Zhang, Y. R., Tang, J. T., Zhou, S. & Wu, D. H. (2016). Capsaicin and dihydrocapsaicin induce apoptosis in human glioma cells via ROS and Ca<sup>2+</sup>-mediated mitochondrial pathway. *Molecular Medicine Reports*, 14, 4198-4208. <https://doi.org/10.3892/mmr.2016.5784>
- Zhang, S., Wang, D., Huang, J., Hu, Y. & Xu, Y. (2019). Application of capsaicin as a potential new therapeutic drug in human cancers. *Journal of clinical pharmacy and therapeutics. Journal of Clinical Pharmacy and Therapeutics*, 45, 16-28. <https://doi.org/10.3892/mmr.2016.5784>
- Zhu, M., Yu, X., Zheng, Z., Huang, J., Yang, X., Shi, H. (2020). Capsaicin suppressed activity of prostate cancer stem cells by inhibition of Wnt/ $\beta$ -catenin pathway. *Phytotherapy Research*, 34, 817-824. <https://doi.org/10.1002/ptr.6563>